

BEST MANAGEMENT PRACTICES FOR MAINE GOLF COURSES

JANUARY 2021



BEST MANAGEMENT PRACTICES FOR MAINE GOLF COURSES



This manual is an adaptation of the Best Management Practices for New England Golf Courses. The text contained in this document was originally authored for the New England document and adapted to meet the specific needs and requirements of Maine.



Disclaimer: The information contained in this document is provided on an “as is” basis with no guarantees of completeness, accuracy, usefulness or timeliness and is solely at the discretion of and/or the opinion of the author. The opinions expressed in this publication are those of the authors. They do not purport to reflect the opinions or views of the GCSAA, USGA, PGA TOUR.

Cover photos: Sunday River Golf Club © Tim O’Wril (top), Val Halla Golf Club © Toby Young (left), © Samoset Golf Club, Rockport, ME (right).

FOREWORD

I commend the Maine Golf Course Superintendents Association for this excellent manual. It's an important contribution to our ongoing work to protect Maine's environment.

Maine's golf courses are gaining wide recognition as a valued recreational resource. At the same time, our communities want to know that golf courses are correctly managing their water resources, including protecting valuable wetlands and employing good IPM and nutrient management methods. This manual provides extensive guidance, developed first at the New England regional level and subsequently refined specifically for Maine, and includes excellent case studies focused on a pair of Maine courses. Doing it right is the Maine way—and this manual provides a great blueprint to do just that.

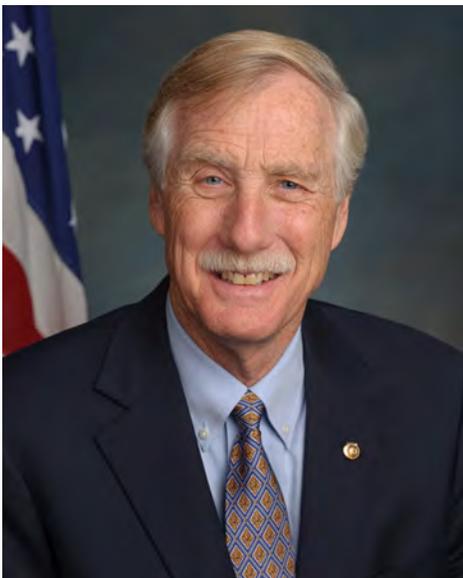
I am especially glad that the manual emphasizes the opportunity for golf courses to provide valuable wildlife habitat, including pollinator-friendly gardens, grasses, and tall meadow areas. Not only do most golfers value seeing a fox slowly crossing a fairway or a hawk guarding a green, but Maine citizens in general are now coming to realize that golf courses can connect wildlife corridors and enhance critical habitat for many species.

In addition, you oversee an important part of Maine's economy, with our state's golf courses generating (from a UMaine study) about \$300 million in revenue, 5,000 full and part-time jobs paying \$100 million to your employees, and making \$2 million in charitable donations!

This manual should not only assist your good work; I hope it will also inspire you to reach out to your local Maine communities to demonstrate the value of their nearby golf course. My Scottish ancestors who brought us this great game of golf already appreciate everything you do!

Angus S. King, Jr.

United States Senate



ACKNOWLEDGMENTS

The preparation of this document was a collaborative effort among authors, external reviewers, and the Maine Chapter of the Golf Course Superintendents Association.

The Maine Golf Course Superintendents Association worked with the other five state golf course superintendents' associations in New England, in collaboration with the New England Regional Turfgrass Foundation and the Universities of Connecticut and Massachusetts, to produce a manual of best management practices to foster improved environmental sustainability and stewardship on New England golf courses. The *Best Management Practices for New England Golf Courses* document was published through this collaborative effort in February 2020. The Maine Golf Course Superintendents Association then took the additional step in adapting this regional Manual to create a state-specific Manual for golf course management in Maine. This is part of a larger effort across the country to create a specific guide for each state. The Steering Committee responsible for this Best Management Practices for Maine Golf Courses manual deserves recognition for their dedication to this document. Steering Committee members include:

Brian Bickford, Maine State Golf Association

Scott Cybulski, Certified Golf Course Superintendent (CGCS), Martindale Country Club

Kevin Doyle, Northeast Regional Representative, Golf Course Superintendents Association of America

Gene Pierotti, Superintendent, Riverside Municipal Golf Course

Robert Searle, Superintendent, Abenakee Club

Individuals from the state of Maine were selected by the Superintendents Association to serve as external reviewers and provide expertise in their field to the applicable chapter within this manual. The development of a state-specific manual would not be possible without their contributions.

© Portland Country Club, Falmouth, ME.



The following lists each chapter in the Manual, the original New England chapter author, and the stakeholder reviewer(s) who generously assisted the steering committee with adopting the New England Manual to conditions in Maine. Thank you to all of the course superintendents and collaborators who provided us beautiful photos of Maine courses to supplement this manual.

Stakeholder Review Team

Chapter 1 - Terrence DeWan FASLA, Maine Licensed Landscape Architect, TJD&A

Chapter 2 - Warren Michaelsen, Environmental Solutions

Chapter 3 - Jeff Dennis, Kristin Feindel, Kathy Hoppe, Addie Halligan, Amanda Pratt, and Marianne Senechal, Maine Department of Environmental Protection (Maine DEP)

Chapter 4 - Jeff Dennis, Kristin Feindel, Kathy Hoppe, Addie Halligan, Amanda Pratt, and Marianne Senechal, Maine DEP

Chapter 5 - Bruce Hoskins, University of Maine Soil Testing Laboratory

Chapter 6 - Gary Fish, State Horticulturist, State of Maine Board of Pesticides Control

Chapter 7 - Kathy Murray, IPM Entomologist, Maine Department of Agriculture, Conservation and Forestry

Chapter 8 - Gary Fish, State Horticulturist, Maine Department of Agriculture, Conservation and Forestry

Chapter 9 - Jennifer Lund, State Apiarist, Maine Department of Agriculture, Conservation and Forestry

Chapter 10 - Chris Baldwin, District Engineer, Cumberland County Soil and Water Conservation District

Chapter 11 - Al Lappin, Maine Landscape and Nursery Association

Chapter 12 - Greg Smith, Director of Business Analytics & Data Services, Competitive Energy Services

New England Manual Authors

Chapter 1 - Mark Mungeam, Mungeam Cornish Golf Design

Chapter 2 - Bob Healey, Irrigation Management & Services and Michelle DaCosta, Ph.D. University of Massachusetts

Chapter 3 - Sarah Stearns, PWS, Beals + Thomas

Chapter 4 - Sarah Stearns, PWS, Beals + Thomas

Chapter 5 - Karl Guillard, Ph.D. & John Inguagiato, Ph.D., University of Connecticut

Chapter 6 - Jason Henderson Ph.D. & Steven Rackliffe, CGCS, University of Connecticut

Chapter 7 - Stan Swier Ph.D., New England Pest Consulting

Chapter 8 - John Inguagiato, Ph.D., University of Connecticut & Dr. Pat Vittum, University of Massachusetts

Chapter 9 - Pat Vittum, Ph.D. University of Massachusetts

Chapter 10 - Golf Course Superintendents Association of America

Chapter 11 - Vickie Wallace, University of Connecticut

Chapter 12 - Amelia Gulkis, Ensave



Golf Course Superintendents Association of America

The Golf Course Superintendents Association of America (GCSAA) is the professional association for the men and women who manage and maintain the game’s most valuable resource — the golf course. Today, GCSAA and its members are recognized by the golf industry as one of the key contributors in elevating the game and business to its current state. Since 1926, GCSAA has been the top professional association for the men and women who manage golf courses in the United States and worldwide. From its headquarters in Lawrence, Kansas, the association provides education, information and representation to more than 17,000 members in more than 72 countries. GCSAA’s mission is to serve its members, advance their profession and enhance the enjoyment, growth and vitality of the game of golf.



Environmental Institute for Golf

The Environmental Institute for Golf (EIFG) fosters sustainability by providing funding for research grants, education programs, scholarships and awareness of golf’s environmental efforts. Founded in 1955 as the GCSAA Scholarship & Research Fund for the Golf Course Superintendents Association of America, the EIFG serves as the association’s philanthropic organization. The EIFG relies on the support of many individuals and organizations to fund programs to advance stewardship on golf courses in the areas of research, scholarships, education, and advocacy. The results from these activities, conducted by GCSAA, are used to position golf courses as properly managed landscapes that contribute to the greater good of their communities. Supporters of the EIFG know they are fostering programs and initiatives that will benefit the game and its environment for years to come.



United States Golf Association

The United States Golf Association (USGA) provides governance for the game of golf, conducts the U.S. Open, U.S. Women’s Open and U.S. Senior Open as well as 10 national amateur championships, two state team championships and international matches, and celebrates the history of the game of golf. The USGA establishes equipment standards, administers the Rules of Golf and Rules of Amateur Status, maintains the USGA Handicap System and Course Rating System, and is one of the world’s foremost authorities on research, development and support of sustainable golf course management practices.

SPECIAL ACKNOWLEDGMENTS

The GCSAA and EIFG wish to thank the University of Florida, Institute of Food and Agricultural Sciences, faculty, Dr. J. Bryan Unruh, Dr. Travis Shaddox, Dr. Jason Kruse, and Mr. Don Rainey, who worked on this project, providing their knowledge and expertise to help the golf course industry; the USGA for their grant to fund this important project; the volunteers who served on the task group to review BMP and provide technical assistance; and the Florida Department of Environmental Protection for permission to copy its publication, “Best Management Practices for the Enhancement of Environmental Quality on Florida Golf Courses”.

Table of Contents

FOREWORD.....	iii
ACKNOWLEDGMENTS	iv
CHAPTER 1: PLANNING, DESIGN, AND CONSTRUCTION.....	1
Planning	2
Design	8
Construction	18
Turfgrass Establishment.....	21
External Programs	24
Resources	24
CHAPTER 2: IRRIGATION	25
Water Conservation and Efficient Use Planning.....	26
Irrigation Management Decision-Making.....	29
Irrigation Scheduling.....	31
Irrigation Water Sources	34
Irrigation Water Suitability	35
Irrigation System Design	36
Pump Stations.....	39
Irrigation System Installation	40
Irrigation System Maintenance and Performance.....	40
Metering.....	43
Irrigation Leak Detection.....	43
Irrigation System Renovation	44
Irrigation Record Keeping	44
CHAPTER 3: WATER QUALITY MANAGEMENT AND PROTECTION	45
Environmental Fate and Transport	46
Pollution Prevention	48
Stormwater Management.....	49
Buffers.....	52
Wetlands.....	53
Lake and Pond Management	54
CASE STUDY: PROTECTING WATER QUALITY ON ABENAKEE GOLF COURSE.....	58
CHAPTER 4: WATER QUALITY MONITORING	60

Review Existing Data	61
Groundwater Monitoring.....	61
Surface Water Monitoring.....	62
Things to Consider	62
Water Quality Analysis	63
CHAPTER 5: GOLF TURF FERTILIZATION AND NUTRIENT MANAGEMENT	66
Essential Elements for Turfgrass and Plant-Available Ionic Forms.....	67
Basis for Fertilization.....	68
Soil Testing.....	71
Plant Tissue Analysis.....	79
Fertilizers Used in Golf Course Management	81
Slow-Release Nitrogen Sources.....	84
Phosphorus.....	90
Potassium.....	91
Soil pH.....	93
CHAPTER 6: CULTURAL PRACTICES.....	97
Turfgrass Selection	98
Mowing	101
Cultivation.....	107
Topdressing	108
Plant Growth Regulators	110
CHAPTER 7: INTEGRATED PEST MANAGEMENT	111
Monitoring Pests and Recording Information.....	113
Identifying and Understanding Pests	115
Determining Pest Threshold Levels	118
Control Methods.....	118
Reduced-Risk Pesticides and Biopesticides.....	121
Evaluation and Record Keeping.....	123
CHAPTER 8: PESTICIDE MANAGEMENT	124
Human Health Risks.....	125
Environmental Fate and Transport	128
Pesticide Risk Assessment Tools	130
Pesticide Storage	132
Pesticide Inventory.....	135
Pesticide Mixing/Washing.....	135

Personal Protective Equipment.....	137
Pesticide Container Management.....	137
Emergency Preparedness and Spill Response	138
Sprayer Calibration.....	139
Sprayers and Nozzles.....	140
Pesticide Record Keeping.....	141
CHAPTER 9: POLLINATORS	142
Pesticides and Pollinators.....	143
Pesticide Application Practices to Protect Pollinators	146
Pollinator-Related Communication	148
Enhancing Pollinator Habitat.....	149
CASE STUDY: MAKING WAY FOR POLLINATORS AT RIVERSIDE GOLF COURSE	151
CHAPTER 10: MAINTENANCE OPERATIONS.....	153
Storage and Handling of Fertilizers.....	154
Equipment Storage and Maintenance.....	156
Equipment Washing.....	157
Fueling Facilities	158
Waste Handling.....	160
Emergency Preparedness and Spill Response	161
CHAPTER 11: SUSTAINABLE LANDSCAPING IN OUT OF PLAY AREAS.....	162
Benefits of Sustainable Areas on the Golf Course	163
Sustainable Landscaping Concepts.....	164
Sustainable High Visibility Areas.....	166
Sustainable Naturalized Areas	168
Habitat Corridors	169
Meadows/Tall Grass Areas	170
Plant Selection	172
CHAPTER 12: ENERGY MANAGEMENT.....	175
General Energy Efficiency Considerations.....	175
Buildings and Amenities	177
Course Management.....	180
Renewable Energy.....	181
Funding Resources.....	183
REFERENCES	184



CHAPTER 1: PLANNING, DESIGN, AND CONSTRUCTION

Belgrade Lakes Golf Club, Belgrade, ME © Tim O'Wril

PREFACE

The development of a new golf course or the renovation of an existing facility requires consideration of numerous environmental, economic, and site suitability factors. These factors must be carefully considered during each step of planning, design, and construction of both new and existing courses, to ensure that the final project is viable, sustainable, and ecologically sensitive (Table 1-1). The thoughtful use of best management practices (BMPs) throughout the planning, design and construction of the course are important to a successful result. Use of BMPs should not be considered a regulatory burden. Instead, they improve and protect natural resources, wildlife habitats, and water quality for current and future generations. Furthermore, thinking holistically and building facilities to maximize sustainability will have a positive impact on maintenance, operations, landscaping, and energy efficiency.



Samoset Golf Club, Rockport, ME. © Forrest Bell

Planning

Design Team. The implementation of a golf course project typically benefits from the use of experienced professional consultants familiar with both local and state¹ and federal² (U.S. Army Corps of Engineers) land use regulations as well as the intricacies of golf course design. The consultants needed depends on the scope and complexity of the proposed work and the environmental constraints present at the project site. Most projects commence with the hiring of a qualified golf course architect. The team would also include a civil engineer and an environmental consultant. For most renovation projects, this may be the extent of consultants required for planning. Depending upon the nature of the project, additional expertise may be required.

Site Analysis. The first step in planning is the development of a detailed site analysis – an existing conditions plan identifying property boundaries, topography, vegetation limits, roads, wetlands, vernal pools, drainage patterns, soil conditions, and other jurisdictional areas. The site analysis is critical in planning a project to take advantage of opportunities, avoid negative environmental impacts, and determine the feasibility of achieving project goals. This initial step will identify site constraints – along with an evaluation of a suitable water source (for new courses) – may determine that a site is unsuitable for the intended project before expensive planning and permitting begins.

Key Players. Once the suitability is confirmed through generation of preliminary concept plans and cost estimates, a team is generally assembled to guide the project. The golf course architect and civil engineer may be helpful in assembling the permitting team.

Professional, experienced judgment is crucial when implementing BMPs in the planning, design and construction phases of the project. An experienced golf course superintendent is integral to the planning process for any golf project. For course renovation projects, with their extensive knowledge of the site, they can assist in determining the most suitable design and can inform the design team of issues that may impact maintenance of the course or player enjoyment of the facility. The superintendent's knowledge of the BMPs and their direct participation in planning and construction can greatly affect the success and ultimately the environmental sustainability of the project.

Table 1-1. Overview of the steps involved in golf course planning, design, and construction.

Planning	
Step	Description
<i>1. Assemble Team</i>	The team should include, but not be limited to, a golf course architect, golf course superintendent, clubhouse architect, irrigation engineer, environmental engineer, energy analyst, economic consultant, civil engineer, soil scientist, golf course builder, biologist or ecologist, and a legal team. Professionals involved in the design of the course (e.g., landscape architects, architects, civil engineers) should be licensed in the State of Maine.
<i>2. Define Objectives</i>	Identify realistic goals, formulate a timeline, etc.
<i>3. Conduct a Feasibility Study</i>	Evaluate finances, environmental issues, availability of water for irrigation and domestic use, and energy, materials, and labor needs. Identify applicable government regulations, permitting requirements, and timelines for securing permits.
<i>4. Select and Analyze Site</i>	Site should meet project goals and expectations. Identify all strengths and weakness of each potential site. During site selection, any site constraints, such as the presence of listed species, significant wildlife habitat (e.g., vernal pools), or invasive species should be identified.
Design	
<i>5. Retain a Project Manager/Superintendent</i>	This person is responsible for integrating sustainable practices in the development, maintenance, and operation of the course. The superintendent will work with the project team, including the architect, civil engineers, and more.
	The project manager/superintendent is responsible for meeting the municipal and state planners early in the process to understand local and state regulations that may affect the design of the course and permitting process.

Design	
Step	Description
6. Design the Course	Existing native landscapes (mature vegetation, steep slopes, major drainage patterns, stone walls, rock outcrops, etc.) should remain intact as much as possible. Consider supplemental native vegetation to enhance existing vegetation alongside lengthy fairways and out-of-play areas. Nuisance, invasive, and exotic plants should be removed and replaced with native species suitable to the area.
	Integrate unique site characteristics into the planning of the course to add variety, visual interest, and to minimize site disturbance. Consider viewscapes, and aesthetically pleasing plant species when designing both the in- and out-of-play areas of a course.
	Structural BMPs: Incorporate structural BMPs into the design plan, identifying opportunities to detain and infiltrate stormwater and to improve water quality through stormwater volume reduction, and water filtration through both physical and biological processes.
	Greens: Should have plenty of sunlight and be well drained. Greens should be big enough to have several hole locations that can handle expected traffic. Root zone material in the greens should be selected with United States Golf Association (USGA) specifications in mind, as published in A Guide to Constructing The USGA Putting Green . Physical testing of the root zone materials should be completed by an accredited laboratory prior to use is recommended.
	Grass Selection: Species should be selected based on climate, environmental, and site conditions, including disease resistance, drought tolerance, spring greenup, and traffic tolerance.

Martindale Country Club, Auburn, ME. © Scott Cybulski, CGCS



Design	
Step	Description
6. <i>Design the Course (continued)</i>	<p>Bunkers: The number and size of bunkers depend on many design considerations, as well as the resources available for daily maintenance. For each bunker consider:</p> <ul style="list-style-type: none"> • The need for drainage • Entry/exit points and how these will affect wear-and- tear patterns • The proper color, size and shape of bunker sands to meet needs
	<p>Vegetative Filters: Vegetative filters (conservation buffers, vegetated filter strips, drainage swales, etc.) can be used throughout the golf course to act as natural biofilters to reduce stormwater flow and pollutant load. Turf areas are also effective filters.</p>
7. <i>Design Irrigation System</i>	<p>Hire a professional irrigation designer to design the irrigation system. Keep in mind the different water needs of greens, tees, fairways, roughs, and native areas. Consider the topography, prevalent wind speeds, and wind direction when spacing the heads. Choose the most efficient type of irrigation system considering available resources.</p>
Construction	
8. <i>Select Qualified Contractors</i>	<p>Use only qualified contractors who are experienced in the special requirements of golf course construction, such as members of the Golf Course Builders Association of America.</p>

Construction	
Step	Description
9. <i>Safeguard Environment</i>	Follow all design phase plans and environmental laws. Utilize the Maine DEP BMP Technical Design Manual (https://www.maine.gov/dep/land/stormwater/stormwaterbmps/index.html) to select appropriate stormwater controls during construction to minimize erosion and sediment transport during construction. Identify all invasive species present on the property. Prevent the spread of invasive species during construction by removing the invasive species prior to construction (if possible) and avoiding these areas with equipment if invasive species are still present. Inspect all equipment and material being introduced to the site to avoid introduction of new invasive species. See chapter 2 for more details on selecting the appropriate stormwater controls during construction to minimize erosion and sediment transport.
10. <i>Install Irrigation System</i>	Installation should consider the need to move equipment and bury pipe while maintaining the original soil surface grade to minimize the potential for erosion.
11. <i>Establish Turfgrass</i>	Turfgrass establishment methods and timing should allow for the most efficient progress of work, while optimizing resources and preventing erosion from bare soils before grass is established.

Martindale Country Club, Auburn, ME. © Scott Cybulski, CGCS



WATER SOURCES

Golf courses are perceived as using a lot of water. Work with the local municipality to determine the amount of water a course can use for irrigation, and the source of that water. Local officials and/or Maine DEP/LUPC may require that the course utilize recycled water, or limit the amount of water taken from wells, streams or lakes. Create a contingency plan for drought conditions to ensure back-up water supplies are in place. For some courses that lack their own supply, the only option is to purchase costly municipal water. These requirements can severely impact the economic feasibility of a golf project. For more information on water sources, see the Irrigation chapter.

WETLANDS AND STREAMS

The Maine DEP defines “Freshwater wetlands” as freshwater swamps, marshes, bogs and similar areas that are inundated or saturated by surface or groundwater at a frequency and for a duration sufficient to support, and which under normal circumstances do support, a prevalence of wetland vegetation typically adapted for life in saturated soils; and, not considered part of a great pond, coastal wetland, river stream or brook. (Maine DEP 2019)

The boundaries and buffer zones of any wetlands, vernal pools, coastal zones, water bodies, intermittent streams and rivers should be identified, flagged and mapped in accordance with local, state and federal regulations by a qualified wetland scientist. Regulated activities, such as draining, dredging, clearing and filling within or in proximity to wetlands, streams, or rivers, generally will require a Natural Resource Protection Act (NRPA) permit³ from Maine DEP. A professional consultant should be utilized to determine permitting needs and provide design assistance to reduce impacts. In some cases, the scope of a project can be changed to eliminate work in a regulated area. If not, the design may be altered to reduce impacts or generate other environmental improvements.

FLOODPLAINS

Golf courses can be a compatible use of floodplain zones, depending on the frequency and severity of flooding. When persistent floods could result in frequent course closures, turf loss and/or significant sediment removal and bunker repair, then use for golf may not be sustainable without improvements. As part of the site analysis, determine whether any part of the property lies in the 100-year flood identified on a Flood Insurance Rate Map (FIRM) prepared by the Federal Emergency Management Agency. Before committing to use an area that may be affected by a 100-year flood, check with the municipality and Maine DEP/LUPC to see if there are any local or state regulations that would apply and possibly restrict use.

LISTED SPECIES AND HABITATS

The presence of either listed species or habitats of concern within the site boundaries or in proximity to the project site could limit the goals and objectives for the project. In addition to identifying wetlands and floodplains before intensive planning, the absence of any listed species or habitats of concern

³ <https://www.maine.gov/dep/land/nrpa/index.html>

should be verified with the Maine Natural Areas Program⁴, part of the Department of Agriculture, Conservation, and Forestry. The Program provides a variety of maps, data, technical assistance, and site review to assist in the evaluation of properties. In addition, as part of the site selection and evaluation process, golf course developers should contact Beginning with Habitat (BwH), a collaborative program of federal, state, and local agencies and non-governmental organizations. BwH is a habitat-based approach to conserving wildlife and plant habitat on a landscape scale. For every town in Maine, BwH has prepared a collection of maps that depict and describe various habitats of statewide and national significance, designed to be used in local land use planning efforts. BwH is designed to help local decision-makers create a vision for their communities and to help developers create plans that provide habitat for all species while balancing future development with conservation.

Best Management Practices for Planning

- Assemble a qualified team with expertise in site evaluation, golf course development, engineering practices, and environmental permitting.
- Determine objectives and complete an environmental and economic feasibility study of the project.
- Select an appropriate site that is capable of achieving the needs of stakeholders and identify strengths and weakness of the selected site.
- Determine water source and supply information for the site.
- Prepare a site inventory and analysis plan.
- Host preliminary meetings with state and local regulators.
- Have a qualified specialist accurately identify wetland boundaries.
- Identify any listed species or habitat of concern on the site.

Design

It is important to establish clear and achievable goals and objectives at the commencement of any proposed project. Project goals should be focused on the overall vision of the course while project objectives should be a measurable set of activities that can be accomplished to achieve the goal. The professional team can assist the course owner or developer in refining the goals and objectives by providing concept plans and cost estimates for the work in the preliminary phase of design. This information helps to determine what changes are appropriate for the site, the financial feasibility of the project and the anticipated schedule for implementation. Although the process of developing goals and objectives varies depending on the complexity of the proposed work, projects are most successful when a clear scope of work is defined and thoughtfully implemented.

A good design meets the needs of the stakeholders, protects the location's environmental resources, and is economically sustainable. The design should address a number of site issues, such as the ones discussed below.

⁴ <https://www.maine.gov/dacf/mnap/>

ENVIRONMENTAL IMPACTS

Consider how to develop the land responsibly and view site limitations as opportunities for unique site design. Design with nature to avoid impacts from the initiation of the project and to make the course fit into the natural landscape to minimize environmental impacts. The design should avoid or minimize impacts to sensitive environmental issues that may have been identified during the site review and preliminary planning phase. When impacts are unavoidable, the design should identify the scope of the impact and address how future use and maintenance of the course will be undertaken to lessen negative impacts. The Maine DEP Site Location of Development Permit⁵ will require the project design to meet applicable standards addressing areas such as stormwater management, groundwater protection, infrastructure, wildlife and fisheries, noise, and unusual natural areas.

WETLANDS DEFINED

The United States Army Corps of Engineers (USACE) defines wetlands as places “Inundated or saturated by surface or groundwater at a frequency and for duration to support, and which under normal circumstances do support, a prevalence of wetland vegetation typically adapted for life in saturated soil”.

Wetlands in Maine fall under four general categories—marshes, swamps, bogs, and fens. Marshes are typically open-canopy wetlands dominated by herbaceous vegetation. Examples include cattail (*Typha* sp.) marshes and tussock sedge (*Carex stricta*) meadows. Swamps are wetlands that are forested, like red maple (*Acer rubrum*) swamps and pocket swamps. Bogs are ombrotrophic freshwater wetlands, meaning they receive all their water input from rain. They are characterized by thick, spongy peat deposits, evergreen trees and shrubs, and a floor covered by a thick carpet of sphagnum moss. Fens are similar to bogs, but are minerotrophic, meaning they receive water mainly from groundwater.

Identifying wetland areas and locating their boundaries is not always straightforward. To identify and delineate wetlands many states (including Maine) use the method developed by the United States Army Corps of Engineers which is based on a three-factor approach involving indicators of hydrophytic vegetation, hydric soil, and wetland hydrology. The USACE classifies wetlands according to Cowardin et al.'s (1979) “Classification of Wetlands and Deepwater Habitats of the United States” which divides wetlands into five main systems: Marine (salt water), Estuarine (brackish water), Riverine (river-associated), Lacustrine (lake-associated), and Palustrine (all other freshwater wetlands less than 20 acres in size with water less than two meters deep). These systems are further broken down into Subsystems, Classes, and Subclasses. Laypersons can readily identify wetland types at the wetter end of the spectrum, such as cattail marshes. However, wetlands that do not have surface water for all or a portion of the year may go unrecognized.

Wetlands provide many societal benefits: food and habitat for fish and wildlife, including threatened and endangered species; water quality improvement; flood storage; shoreline erosion control; economically beneficial natural products for human use; and opportunities for recreation, education, and research.

5 <https://www.maine.gov/dep/land/sitelaw/index.html>

MAINE WETLANDS OF SPECIAL SIGNIFICANCE

In Maine, Wetlands of Special Significance are regulated by the Maine Department of Environmental Protection (DEP). All coastal wetlands and great ponds (inland bodies of water >10 acres in size) are classified as wetlands of special significance. In addition, a freshwater wetland may be considered one of special significance if it: (1) contains a natural community that is critically imperiled or imperiled as defined by the Maine Natural Areas Program; (2) contains significant wildlife habitat; (3) is located within 250 feet of a coastal wetland; (4) is located within 250 feet of a great pond; (5) contains at least 20,000 square feet of aquatic vegetation, emergent marsh vegetation, or open water; (6) is inundated with floodwater during a 100-year flood event based on flood insurance maps; (7) is or contains peatlands; and (8) is located within 25 feet of a river, stream or brook. More information on Wetlands of Special Significance is available at [http://www.beginningwithhabitat.org/pdf/Wetland%20_definitions.pdf](http://www.beginningwithhabitat.org/pdf/Wetland%20definitions.pdf).

MAINE STREAMS

The Maine Natural Resources Protection Act provides the following definition. “River, stream or brook” means a channel between defined banks. A channel is created by the action of surface water and has two or more of the following characteristics:

- A. It is depicted as a solid or broken blue line on the most recent edition of the U.S. Geological Survey 7.5-minute series topographic map or, if that is not available, a 15-minute series topographic map.
- B. It contains or is known to contain flowing water continuously for a period of at least 6 months of the year in most years.
- C. The channel bed is primarily composed of mineral material such as sand and gravel, parent material or bedrock that has been deposited, or scoured by water.
- D. The channel contains aquatic animals such as fish, aquatic insects, or mollusks in the water or, if no surface water is present, within the stream bed.
- E. The channel contains aquatic vegetation and is essentially devoid of upland vegetation.
- F. “River, stream or brook” does not mean a ditch or other drainage way constructed, or constructed and maintained, solely for the purpose of draining stormwater or a grassy swale.

VERNAL POOLS IN MAINE

Vernal pools are also referred to as vernal ponds, ephemeral pools, seasonal woodland pools, or seasonal wetlands. They are naturally-occurring, seasonal bodies of water that lack permanent fish populations. The amount of time that pools hold water varies from pool to pool and from year to year, depending largely on the amount of precipitation in a given year. Vernal pools provide breeding habitat for one or more of Maine’s vernal pool indicator species: spotted salamanders (*Ambystoma maculatum*), blue-spotted salamanders (*Ambystoma laterale*), wood frogs (*Lithobates sylvaticus*), and fairy shrimp (Order Anostraca).

A “classic” vernal pool looks essentially like a giant puddle in the woods. However, they can be found in a variety of wetland settings. For example, a given vernal pool could also be correctly classified as marsh, shrub-swamp, or forested wetland. Vernal pools also occur in floodplains, or may be embedded in larger wetland complexes. For example, an area of pooled water within a larger forested wetland complex may function as a vernal pool. For these reasons, vernal pools are best defined by the species that breed in them, rather than by their physical structure.

In Maine, vernal pool is considered to be “significant” by regulatory authorities if it has a high habitat value, either because (1) a state-listed threatened or endangered species, such as a spotted turtle, or a rare species, such as a ribbon snake, uses it to complete a critical part of its life history, or (2) there is a notable abundance of specific wildlife, such as the blue spotted salamander, wood frog, or fairy shrimp.

WETLAND REGULATION IN MAINE

Congress established federal regulatory power regarding wetlands under Section 404 of the Clean Water Act. In Maine, the 1988 Natural Resources Protection Act (NRPA) established state regulatory authority over wetlands. The federal, state, and local, and private entities involved with wetlands regulation are described below.

Army Corps of Engineers (Federal): The USACE is responsible for overseeing impacts to wetlands and waterways (“waters of the United States” under the Clean Water Act) resulting from drain and fill activities to wetlands (e.g., areas drained, flooded, fragmented, cleared, or excavated).

US Fish and Wildlife Service (FWS) and US Environmental Protection Agency (EPA) (Federal): Both these agencies provide input to the USACE on proposals for development that will have impacts on wildlife habitat, including wetlands.

Maine Department of Environmental Protection (State): The DEP is responsible for permitting and enforcement associated with wetland alteration activities within the state.

Maine Department of Inland Fisheries and Wildlife (MDIFW) (State): MDIFW maps high value wildlife habitats and provides technical expertise in regard to areas deemed Significant Wildlife Habitat (including certain wetland types). MDIFW maintains a mapped database of Significant Vernal Pools and makes recommendation as to pool status to DEP.

Town Code Enforcement Officers and Planning Boards (Local): Code Enforcement Officers administer and enforce municipal zoning, building, and similar ordinances, and, especially when local standards overlap with state and federal regulations, they encourage adherence to state and federal regulations such as protection of natural resources, including wetlands.

Environmental Consulting Firms (Private): Environmental consulting firms may be hired to delineate wetlands, identify vernal pools and determine their significance status, and help landowners apply for permits and adhere to regulations.

FLOODPLAINS

Any planned substantial disturbance to a floodplain, including clearing and grading, generally requires an engineering analysis to demonstrate minimal impact on the base flood elevation in accordance with local ordinances. Depending on the complexity of the encroachment, this analysis may be as simple as a comparison of cut and fill quantities within the floodplain or as complex as a detailed floodplain model of the entire watershed. A complex analysis may require a Federal Emergency Management Agency (FEMA) review along with potential revision to the floodplain mapping. As noted above in Planning, if the project site and the proposed activity falls within a designated floodplain, contact the municipal planning office to determine what restrictions (if any) may apply.

Floodplain restoration is the reestablishment of natural water systems to help mitigate flooding and control stormwater. During the planning and design phase, floodplain restoration activities can address vertical and lateral stream migration, which causes unstable banks, flooding, reductions in groundwater recharge, and high sediment and nutrient loads. When incorporating floodplain restoration into the course design or renovation, land use decisions and engineering standards must be based on the latest research science available. Where appropriate and if land is available, installing or enhancing stream buffers may also help restore or enhance natural water flows and flooding controls as well as providing wildlife habitat.

STORMWATER MANAGEMENT

Although golf courses are typically large properties ranging from 60 to 200 acres, they are just one link in a stormwater management chain. Generally, a quantity of stormwater enters the golf course area, supplemented by what falls on the golf course proper, and then the stormwater leaves the golf course. As part of a larger watershed area, it is important that precipitation entering the course is addressed through stormwater management techniques on the course to avoid impacts to downstream properties or uses.

When golf courses are designed, their drainage capability concept is guided by an average rainfall event of a given frequency. For example, a golf course drainage system is typically designed to detain a two- or five-year rain event. In other words, when that rain event happens, the golf course will be able to be reasonably drained in a matter of hours, as excess water not absorbed by the soil flows through the drainage system, is temporarily held, and finally leaves the property.

Maine DEP requires basins and ponds designed to control flows so as to meet the flooding standard must have principal spillways capable of controlling runoff from 24-hour storms of the 2-year, 10-year, and 25-year frequencies. Basins and ponds designed to provide channel protection detention must have principal spillways capable of providing extended detention of twelve hours for runoff from a 24-hour storm of a one-year frequency.

To meet the flooding standard, the applicant must demonstrate that a project's stormwater management systems will meet the following:

1. The system must detain, retain, or result in the infiltration of stormwater from 24-hour storms of the 2-year, 10-year, and 25-year frequencies such that the peak flows of stormwater from the project site do not exceed the peak flows of stormwater prior to undertaking the project;
2. The design of piped or open channel systems must be based on a 10-year, 24-hour storm without overloading or flooding beyond channel limits;
3. The areas expected to be flooded by runoff from a 10-year or 25-year, 24-hour storm must be designated in the application, and no buildings or other similar facilities may be planned within such areas. This does not preclude the use of parking areas, recreation areas, or similar areas from use for the detention of storms greater than the 10-year, 24-hour storm. The applicant shall secure drainage easements from any downstream property owners across whose property may be flooded by runoff pursuant to Section 4(G)(2)(a);
4. Runoff from the project may not flood the primary access road to the project and any public roads bordering the project as a result of a 25-year, 24-hour storm.

The ability to detain large amounts of water requires accurate engineering and extensive construction to prevent physical or financial damage to the facility.

BMPs are intended to prolong the detention process as long as practical, harvest as much of the stormwater in surface or underground storage as reasonable, and to improve the quality of water leaving the property when possible. Methods of stormwater management include infiltration chambers that allow water to better enter the ground and recharge aquifers, retention basins that slow the flow of water off the property during heavy rain events while also trapping sediments, and installation of swales with check dams to slow runoff.

In Maine, if a project disturbs one acre or more of land area it will require a stormwater permit pursuant to the Stormwater Management Law 38 M.R.S. §420-D. Additional stormwater standards and requirements, such as those involving the Maine Pollutant Discharge Elimination System (MEPDES) program⁶ and the Municipal Separate Storm Sewer System (MS4) program⁷ may also apply in addition to the standards and requirements set forth in this Chapter.

“Low impact development” or “green infrastructure” means site planning and design strategies intended to replace or replicate predevelopment hydrology through the use of source control and relatively small-scale measures integrated throughout a site to disconnect impervious surfaces and enhance filtration, treatment, and management of stormwater runoff as close to its source as possible. Low impact development strategies may be either nonstructural or structural, except that low impact development strategies utilizing structural stormwater management techniques shall be limited to an impervious contributing drainage area equal to or less than 1 acre. Low impact development strategies include, but are not limited to: bioretention filters, grass swales and channels, vegetated filter strips, permeable pavements, rain gardens and vegetated rooftops.

Low impact development strategies can reduce stormwater storage volume requirements through the use of non-structural stormwater management techniques that minimize impervious cover, thereby reducing both the size and cost of stormwater management structures. The use of low impact development strategies is optional and voluntary for all projects, but projects using this credit are eligible to reduce the portion of the project’s impervious or developed acreage that must be treated.

Groundwater quality monitoring may be required by the MaineDEP if necessary, to demonstrate that the infiltration system will operate in compliance with the Water Classification Program. Groundwater quality monitoring will generally be required for activities infiltrating water from areas of heavy turf-chemical use, such as golf courses and certain athletic fields, and large connected impervious areas, such as parking lots and runways.

DRAINAGE

Adequate drainage is necessary for healthy turfgrass. Good surface drainage is the most reliable method for removal of water from course play areas. On gently sloped sites, surface drainage may be adequate to remove water without concern for erosion. In general, putting surfaces should be tilted at no less than one-percent (1%) slope and fairways and roughs no less than two-percent (2%) to achieve adequate surface drainage.

⁶ <https://www.maine.gov/dep/water/wd/general.html>

⁷ <https://www.maine.gov/dep/water/wd/ms4/index.html>

Where the ground is very flat, subsurface drainage may be needed to assist in expedient removal. Where the ground is steep, subsurface drainage is helpful in reducing erosion by limiting the length and velocity of overland flow. Subsurface drainage is also installed to control a water table or to interrupt subsurface seepage or flow. Wherever possible, drainage should be directed into vegetative areas for biological filtration or into infiltration basins to minimize potential adverse effects of fertilizers and pesticides on adjacent wetlands or other natural areas.

Drainage is only as good as the system's integrity. Damaged, improperly installed, or poorly maintained drainage systems negatively impact play and increase risks to water quality. The drainage system should be routinely inspected to ensure proper function. Roots and animal activity can easily clog drains and prevent proper functioning.

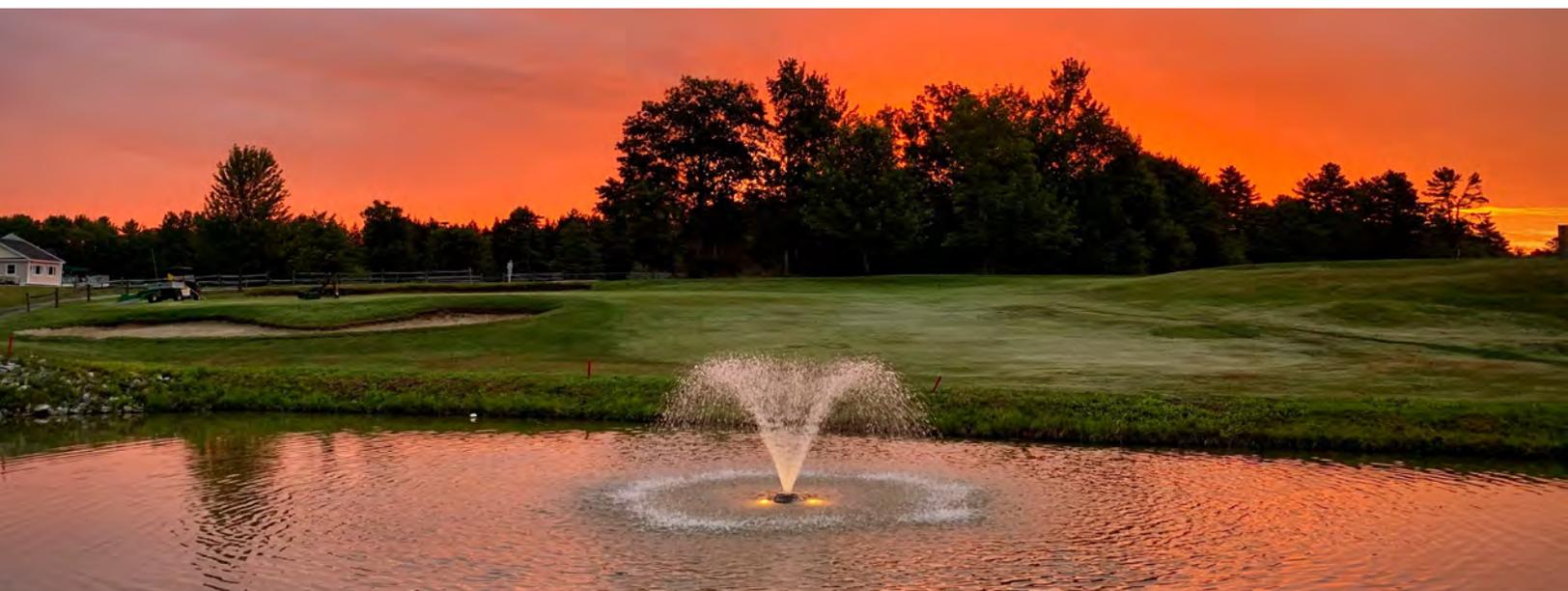
POND LOCATION AND DESIGN

Designing a new pond requires considerations such as the size of the drainage area, water supply, soil types, and water depth. In addition to potentially serving as an irrigation water source, ponds support aquatic life. Therefore, the design of ponds should consider the needs of aquatic ecosystems, such as discouraging excessive growth of aquatic vegetation, supplying sufficient dissolved oxygen (DO) to support aquatic species, etc. Careful design may significantly reduce future operating expenses for pond and aquatic plant management. In addition, water resources should be managed to control or limit the spread of aquatic invasive species, such as submerged plants, fish or invertebrates.

Ponds often have the primary purpose of drainage and stormwater management and are also often a source of irrigation water. When the golf course is properly designed, rain and runoff captured in water hazards and stormwater ponds may provide most or all of the supplemental water necessary under normal conditions, though backup sources may be needed during times of drought.

Furthermore, when designing a new pond or expanding an existing pond, refer to Maine DEP permitting guidelines on pond construction. (See the “Maine Stormwater Design Manual” for technical details.⁸)

Val Halla Golf Course, Cumberland, ME. © Toby Young



8 <https://www11.maine.gov/dep/land/stormwater/stormwaterbmps/vol3/volume%20III%20May%202016.pdf>

HABITAT CONSERVATION

In addition to adhering to regulations that protect listed species, maintaining habitat to the extent possible during all phases of planning, design, and construction helps maintain biodiversity. Natural habitats provide food and shelter for numerous species, including mammals, birds, fish, amphibians, reptiles, insects, and native plants. The “Pollinator” and “ Sustainable Landscaping in Out-of-Play Areas” chapters of this document provide additional recommendations and BMPs for enhancing habitat on the golf course.

© Bob Searle



Best Management Practices for Design

- Involve a qualified golf course superintendent/project manager at the beginning of the design process to integrate sustainable maintenance practices in the development, maintenance, and operation of the course.
- Design the proposed changes to minimize or eliminate alteration to sensitive existing native landscapes. The plans should review alternate designs to determine the concept plan that best meets the objectives with the least disturbance.
- Design the changes to retain as many natural site characteristics as possible.
- Consider potential wear patterns in turf areas and create adequate space for ingress/egress at greens, tees, fairways and bunkers.
- Define play and non-play maintenance boundaries.
- Select a greens location that has adequate sunlight to meet plant specific needs and provides sufficient drainage, or design site improvements to reduce shade and improve drainage characteristics.
- Choose a green size and sufficient number of hole locations large enough to accommodate traffic and play damage, but not so large that it is not sustainable with your resources
- Consider the placement of bunkers in relation to circulation patterns at greens so as not to concentrate turf wear.
- Consider the number, size and style of sand bunkers as they relate to resources available for daily maintenance.
- Be aware of bunker design as it relates to cost of construction and future maintenance. Make sure bunkers have suitable machine entry and exit points.
- Select the proper color, size, and shape of bunker sand that meets the course needs.
- Ensure that wetlands have been properly delineated by a professional consultant before working in and around any wetlands.
- Ensure that proper permitting has been obtained before disturbing any tidal or non-tidal wetland or the regulated buffer zone.
- Establish and maintain an appropriately sized buffer around wetlands, springs, and spring runs.
- Install stream buffers to restore natural water flows and flooding controls.
- Install buffers in play areas to stabilize and restore natural areas that can attract wildlife species.
- Install detention basins to store water and reduce flooding at peak flows.
- Use a swale and berm system to allow for resident time (ponding) for water to infiltrate through the root zone to reduce lateral water movement to surface water bodies.

Best Management Practices for Design (continued)

- When constructing drainage systems, pay close attention to engineering details such as subsoil preparation, the placement of gravel, slopes, and backfilling and placement of drainage gravel.
- Discharge subsurface drainage through pretreatment zones and/or vegetative buffers to help remove nutrients and sediments.
- The drainage system should be routinely inspected to ensure proper function.
- Install berms and swells to capture pollutants and sediments from runoff before it enters irrigation storage ponds.
- Monitor pond water level for water loss (seepage) to underground systems. If seepage is occurring, it may be necessary to line or seal irrigation ponds or install pumps to relocate water.
- Install water-intake systems that use horizontal wells placed in the subsoil below the storage basin. Use a post pump filter to remove particulate matter.
- Remove excess sediments in irrigation ponds to reduce irrigation system failures.
- Select an appropriate site for irrigation levels to be maintained, including in times of drought.
- Design ponds so as to avoid peninsular projections and long, narrow fingers, which may prevent water mixing. Ponds that are too shallow may promote algal growth, excess sedimentation, and exhibit high temperatures and low DO levels.
- Reverse-grade around the perimeter of ponds to control surface water runoff and reduce nutrient loads.
- Construct random small dips and ridges (micro topography) on shorelines of a few inches to a foot to promote diversity for the aquatic plant community and provide a healthier and more productive littoral zone.
- Consult with a Maine licensed professional engineer (civil or geotechnical) when constructing a dam.
- Protect and enhance habitat by:
 - Identifying the different types of habitat specific to the site.
 - Identifying habitat requirements (food, water, cover, space) for wildlife species.
 - Identifying and preserving regional wildlife and migration corridors by avoiding or minimizing crossings. Design unavoidable crossings to accommodate wildlife movement.
 - Designing out-of-play areas to retain or restore existing native vegetation where possible.
 - Removing nuisance and exotic/invasive plants and replacing them with native species that are adapted to a particular site.

Best Management Practices for Design (continued)

- Maintaining clearance between the ground and the lowest portion of any fences or walls to allow wildlife to pass, except in areas where feral animals need to be excluded.
- Retaining dead tree snags for nesting and feeding sites, provided they pose no danger to people or property.
- Constructing and placing birdhouses, bat houses, bee boxes, etc. in out-of-play areas.
- Design and locate cart paths to minimize environmental impacts. Construct the paths with permeable materials, if possible.

Construction

DOCUMENTS

Prior to starting construction, contract documents (plans and specifications) that clearly communicate the scope of work are needed to communicate all aspects of the project to stakeholders to ensure that all parties understand the project. All critical data from the environmental resource inventory as well as key notes regarding construction processes should be included in the construction documents, as well as any conditions imposed in the permitting process. Documents should include sediment and erosion control and the stormwater management plans that were established in the design phase. During the bidding process, experienced contractors may be able to provide additional insight on appropriate BMPs to accomplish the objectives of the plan.

Challenges often arise in construction that were not foreseen in planning and design. Development of clear and thoughtful plans and specifications can minimize costly changes during the work and will assist those involved in the project in responding to any unforeseen challenges. Proper planning can reduce the number of issues, but it is best if the responsible contractor has significant experience with golf course renovation or new construction. A well-qualified contractor will also be familiar with best management practices to employ during construction.

CONSTRUCTION ACTIVITIES

Construction should begin with the project team meeting with the contractor(s) to review construction protocols. The purpose of this meeting is to define lines of communication, review the scope of work, establish reasonable timetables, review methods for reducing environmental impacts, and for the contractor(s) to become familiar with permit requirements. Any natural resource areas impacted by the project should be clearly delineated prior to the meeting, field reviewed during the meeting, and permit requirements discussed that may impact those areas.

The golf course architect, engineer, irrigation designer and other key consultants should remain involved through the construction phase to ensure plans and specifications are followed and permit conditions

are met. The consultant's role in construction should be defined at the start of work. The leadership structure of the project team should be outlined at the beginning of the project to identify the point of contact for the contractors regarding change orders, de minimis changes, modifications to permitting and approval, etc.

During construction, the site should be kept as stable as possible to reduce erosion and stabilize sediments. For large renovation and new course projects, the contractor should attempt to limit the amount of disturbed area at one time, which may require completing and stabilizing a portion of the site prior to starting on a new area. On smaller projects, rather than phasing, the best method to prevent environmental impacts may be a narrow construction window. The construction schedule and work limitations should be tailored to each site and project. The emphasis during construction must be on performing the work with quality and care to minimize the potential for future problems.

CONSTRUCTION TECHNIQUES

Sound construction techniques include those processes and practices that control soil erosion and stormwater runoff. *Examples* of such techniques include the following (Refer to the Maine Erosion and Sediment Control BMPs Handbook for more details on the appropriate construction technique.):

- Installation of silt fences, erosion control mix berms, or filter socks to minimize erosion prior to any land disturbance.
- Locating construction staging and fueling operations at least two-hundred feet (250') from any water body, wetland or sensitive area.
- Checking barriers prior to a predicted rainfall and removing excess siltation and repairing barriers immediately following a storm.
- Protection of drain inlets with gravel and silt fabric.

SEDIMENT STABILIZATION

The loss of topsoil from a site can be a problem for numerous reasons. Soil and sediment carried by wind and water transports contaminants with it. For example, erosion can enrich surface waters, where phosphorus, and to a lesser extent nitrogen, can cause eutrophication. When sediments and soils enter water, they can also increase turbidity, which can have harmful effects on aquatic plants and animals.

Therefore, control measures need to be documented in an erosion and sediment control plan, put in place prior to any soil disturbance, and properly maintained. Wherever possible, a vegetative cover should be kept on the site until it is ready for construction.

As soon as possible after construction activities are completed, the areas should be planted, sodded, or otherwise covered to prevent stabilize sediments and prevent erosion.

If soils are stockpiled on site during construction, they should be covered with an erosion control cover. Additionally, a sediment barrier should be installed along the downgradient edge to collect runoff and sediments. (See page 15 of the Maine Erosion and Sediment Control BMP Manual for details on BMPs for stockpiled sediments.)

CONSTRUCTION MONITORING

The contractor, owner's representative or a hired consultant should be responsible for monitoring the construction process and providing any reports required by project permits. The level of diligence invested in monitoring can significantly influence the environmental and financial sustainability and viability of the project.

Best Management Practices for Construction

- Conduct a pre-construction conference with stakeholders.
- Use a qualified golf course contractor, such as a member of the Golf Course Builders Association of America, or one with significant local golf course construction experience.
- Consider limiting the area of disturbance at one time. Finish and stabilize one area before starting another area.
- Construction should be scheduled to maximize turfgrass establishment and site drainage.
- Monitor construction activities, maintain a construction progress report, and communicate the report to the proper permitting agencies.
- When constructing drainage systems, pay close attention to engineering details such as subsoil preparation, the placement of gravel, slopes, and backfilling to stabilize soils.
- Discharge subsurface drainage systems through pretreatment zones and/or vegetative buffers, where possible, to help remove sediments and nutrients.
- Develop and implement strategies to effectively control sediment, minimize the loss of topsoil, protect water resources, and reduce disruption to wildlife and plant communities.
- Integrate hydro-mulching, erosion blankets or straw mulch into the seeding process to enhance soil stabilization. Avoid using hay as mulch which may introduce unwanted weed seeds that may become a future problem.
- Check and repair erosion control barriers after every rain event.
- See the Maine Erosion and Sediment Control BMP Manual for more.

Turfgrass Establishment

Turfgrass establishment is a unique phase in turfgrass growth and requires greater quantities of water and nutrients than maintaining established turfgrasses. To this end, the establishment phase should be considered carefully to minimize environmental risk. Selection of turfgrass species or cultivar is one of the most important decisions a manager can make to ensure a healthy turfgrass stand. Prior to establishing turf, turfgrass managers should select grass species and cultivars based on the existing site conditions and the intended use of the turf, as described in the Cultural Practices chapter.

Golf course managers can choose to establish turfgrass via seeding or sodding. Selection of each method will be highly dependent on material cost, and site suitability. Each method is described in detail, below.

SEEDBED PREPARATION

Proper seedbed site preparation can help avoid long-term problems, such as weed encroachment, diseases, and drought susceptibility. Debris should be removed that could hinder root growth and limit access to water and nutrients. Any drainage issues should be corrected through grading and installation of drainage technologies.

Additionally, soil testing should be performed at a certified laboratory to understand the seedbed needs (e.g. amendments, topsoil additions). See Chapter 5 on “Fertilization and Nutrient Management” for details.

SODDING

Sod should be topdressed to fill in the gaps between the pieces to speed establishment and create a smoother surface. During dry weather (summer or winter), light and frequent irrigation is required until the sod takes root. Check for rooting by lightly pulling the corner of the sod.

Irrigation frequency can be reduced when the sod cannot be pulled from the soil surface.

SEEDING

New England’s cool-season turfgrass species (bluegrass, fescues, and ryegrass) should be seeded in late summer, except creeping bentgrass. This timing is generally ideal because soils are warm, nights are cool, and disease and weed pressure are reduced. However, research at the University of Connecticut has found July or early- August to be optimal for seeding creeping bentgrass to minimize annual bluegrass competition which typically germinates during late-summer and fall. Cool-season grasses can also be dormant seeded in late fall through the winter.

Cool-season species can be seeded in the spring following winterkill. However, pre-emergence herbicides should be used to increase the success of grow-in. Without a pre-emergence herbicide, spring seeding success is significantly reduced because of the summer annual weed pressure.

Herbicide labels should be reviewed to ensure that the product is labeled for use during establishment. A drop-type spreader should be used for uniform seed dispersal. Lightly raking the soil or using specialized “slit seeders” improves seed-to-soil contact.

During grow-in and establishment, more water is required than for established stands. Until the sod begins to root down or the seedlings start to establish, water should be applied lightly and frequently. The goal is to keep the surface moist until germination. Irrigation frequency should then be reduced, though the amount of water applied should be slightly increased until the first mowing. Turfgrasses with relatively large seeds (i.e., tall fescue, perennial ryegrass, and fine fescue) generally need fewer irrigation events during establishment than finer-textured seeds (i.e., Kentucky bluegrass and creeping bentgrass).

In phosphorus-deficient soils, phosphorus should be applied to the soil before seeding or sodding at a rate ranging from 0.5 to 1.5 lbs. P_2O_5 per 1,000 square feet (22 to 65 lbs. P_2O_5 per acre). A follow-up application of phosphorus fertilizer may be required four to eight weeks after seedling germination or sodding. A second application is justified if the turf has symptoms of phosphorus deficiency (such as purple-blue color, thin canopy, poor nitrogen response). Soils with a pH greater than 7.5 are also at greatest risk of phosphorus deficiency during establishment. Therefore, higher rates and a second application of phosphorus fertilizer are recommended for high pH soils.

Nitrogen (N) management is essential during establishment. For highly maintained turf stands or turf growing on sand or sand-based soils, soluble sources of N fertilizer should be applied every 7 to 14 days. Fertilizer applications should continue until the turf canopy has achieved 100% cover. Single application rates should not exceed 0.5 lbs. of N per 1,000 square feet. Slow-release nitrogen sources can also benefit establishment regardless of soil type. Higher N application rates may be used with products containing more slow-release nitrogen. Fertilizers with 50% quick-release and 50% slow-release N provide uniform nitrogen release for a period of 6 to 10 weeks, depending on the formulation.

Micronutrient fertilizers can also be beneficial during establishment, especially on sandy or high pH soils. Nutrients such as iron and manganese can sometimes be limiting in these soil conditions. The recommendations found in soil test results are for established turfgrass. For establishing a new stand, these recommendations are not as beneficial.

Instead, deficiencies can be diagnosed through small applications of fertilizer to a section of the turf. Lack of a response indicates that the nutrient is not limiting and that application to the entire area is not warranted.

MOWING

For a majority of turfgrass areas, mowing should begin as soon as the turf height reaches the desired mowing height for that area. An exception may be putting greens. While some managers will start mowing to standard putting green heights immediately (0.15” or less), most managers start mowing at 0.4” and slowly reduce the height of cut as the stand matures. Regular mowing promotes new tiller formation and stimulates the transition from juvenile to mature plants.

Best Management Practices for Turfgrass Establishment

- The area to be established should be properly prepared and cleared of pests (e.g. weeds and pathogens).
- Select cultivars that are adapted to the desired use, taking note of disease resistance, drought, traffic, and shade tolerance, and other traits such as texture and color.
- Ensure erosion and sediment control devices are in place and properly maintained.
- Use mulch (e.g., hydromulch, loose straw from a clean source, straw mats) for soil stabilization.
- Prepare seed/sod bed to maximize success.
- Fill gaps in sod seams with soil or sand to provide a uniform surface.
- Use selective pre-emergence herbicides to reduce weed competition and improve the chance of success with seeding establishment during the spring.
- Apply a fertilizer containing phosphorus at seeding. An additional application should be applied if turf displays symptoms of phosphorus deficiency.
- Nitrogen and sufficient water are essential during establishment. Light and frequent applications of nutrients are most desirable, unless a slow-release nitrogen source is used.
- Mow turf to the desired mowing height as soon as practical to promote density and maturation. Never remove more than one-third of the turf leaf at mowing.

Mowing using hybrid mower technology with reduced fuel consumption on Cape Arundel Golf Club, Kennebunkport, ME. © Brendan Parkhurst



External Programs

Golf courses can gain valuable recognition for their environmental education and certification efforts. Examples of external designations include Audubon International's [Cooperative Sanctuary Program for Golf](#) and the Groundwater Foundation's [Groundwater Guardian Green Site](#) program.

See page 150 in this manual for a Case Study on how a Maine Golf Course (Riverside Municipal Course in Portland) achieved Audubon certification. Six courses in Maine have achieved Audubon certification (Bangor Municipal Course, Castine Golf Club, Mingo Springs Golf Club, Portland Country Club, Riverside Municipal Golf Course, and Sable Oaks Golf Club (recently closed)).

Resources

- **Stormwater Management Law and Chapter 502:** *Direct Watersheds of Lakes Most at Risk from new Development, and Urban Impaired Streams.* Available from the Bureau of Land and Water Quality (287-3901) or on the DEP website at <https://www.maine.gov/dep/land/stormwater/storm.html>
- Maine erosion and sediment control practices field guide for contractors: https://www.maine.gov/dep/land/erosion/escbmps/esc_bmp_field.pdf. *The field guide is also available in a convenient printed wire-bound format. To order a hardcopy of this manual, contact [John Maclaine](#) (207) 615-3279. There is a charge for a hardcopy.*
- Maine erosion and sediment control best management practices (bmps): <https://www11.maine.gov/dep/land/stormwater/stormwaterbmps/vol3/volume%20iii%20may%202016.pdf>



CHAPTER 2: IRRIGATION

Martindale Country Club, Auburn, ME. © Scott Cybulski, CGCS

PREFACE

The judicious use of supplemental water keeps turfgrass and landscape plants healthy, while providing the firm, fast playing surfaces that golfers desire. BMPs that conserve and protect these water resources are integral to facility management. Conservation and efficiency-related efforts consider the strategic use of course and irrigation design, computerized and data-integrated scheduling, and alternative water supply options that support plant health and reduce the potential for negative impacts on natural resources.

Irrigation BMPs may also provide an economic, regulatory compliance, and environmental stewardship advantage to courses that integrate them into an irrigation management plan. BMPs are not intended to increase labor or create undue burden. If applied appropriately, irrigation-related BMPs can help stabilize labor costs, extend equipment life, reduce repairs, and limit overall personal and public liability while protecting and conserving natural resources. Additional comprehensive information that includes detailed irrigation-related BMPs can be found in [Best Management Practices for Golf Course Water Use](#) (Connecticut DEP, 2006).

Water Conservation and Efficient Use Planning

Potable water supplies in many areas of the United States are limited, and demand continues to grow. The challenge is to find solutions to maintain the quality of golf while using less water. Opportunities to conserve water exist when courses are initially designed and during renovation, during irrigation system design and use, and by incorporating the use of management zones. For example, some new courses are designed using a “target golf” concept that minimizes the acreage of irrigated turfgrass and improves the use of the water applied. Similarly, hand-watering specific areas of stress-prone turfgrass can result in significant water savings. If properly designed, water hazards and stormwater ponds can capture rain and runoff that may provide supplemental water under normal conditions, though backup sources may be needed during severe drought.

WATER BUDGETS

The development of a water budget establishes a benchmark for golf course water requirements that can be compared with actual water use, ultimately confirming whether water is being used efficiently or whether changes in management strategy are needed. Using a water budget to accurately estimate a course’s water requirements can translate into improved playing conditions for golfers, lower operating and maintenance costs, and improved resource management. Water budgets take into account the size of the property, historic climate data, effective rainfall, and plant factors. This helps managers make informed decisions about their current water use and the effectiveness of programs designed to reduce water use. The water-budget approach is recognized by the United States Environmental Protection Agency (USEPA) and other federal and state agencies as a science-based approach for estimating landscape water requirements. The United States Golf Association (USGA) provides an online [Water Budget Calculator](#) along with step-by-step instructions for assistance in creating a facility-specific water budget. Precision water management is one of the most important practices for maintaining high-quality golf turf while conserving water resources, as discussed in detail later in this chapter.

Precision water management can be achieved through efficient irrigation practices that replace only the amount of irrigation water needed to maintain healthy turf in playing areas. It requires an efficient and properly functioning irrigation system and regular cultural practices that increase the water-holding capacity of soil.

TURFGRASS SELECTION

Turfgrass selection is an important component of a water conservation efforts. The increased availability of improved turfgrass species and varieties provide an excellent opportunity to select the most well adapted turf to specific site conditions. If selected for drought tolerance, some turfgrass varieties require less water to survive and maintain playability. The [National Turfgrass Evaluation Program](#) (NTEP) provides information on top performing cultivars for various desirable turfgrass traits, including tolerance to drought, traffic and diseases.

OUT-OF-PLAY AREAS

In addition to utilizing well-adapted cultivars for in-play areas, existing golf courses can convert out-of-play area turfgrass to native plants, grasses, or ground covers to reduce water use and augment the site's aesthetic appeal. Native plant species also provide wildlife with habitat and food sources, such as native flower areas that benefit pollinators. After establishment, site-appropriate plants normally require little to no irrigation. [The Native Plant Trust](#) (formerly New England Wild Flower Society) provides information on native plants in the region, and additional lists for drought-resistant landscapes can be found through state university extension programs, including the [University of Massachusetts](#) and [University of New Hampshire](#). See also the “Pollinators” and “Sustainable Landscaping in Out-of-Play Areas” chapters for more information on native and drought-tolerant plants.

WETTING AGENTS

Wetting agents can be useful for managing a number of water-related issues, such as improving irrigation efficiency, assisting in the retention of water in the soil profile, aiding in infiltration, preventing and treating localized dry spot (LDS), or serving as a spray adjuvant when applying pesticides or plant growth regulators (PGRs).

Research shows preventative applications can increase soil water uniformity and sustain high visual turfgrass quality at very low levels of irrigation (30% potential evapotranspiration) [Kostka et al., 2005]. Preventative applications of wetting agents can also increase irrigation precision, which reduces water use while maximizing playing conditions. Late fall applications may reduce water repellency in soils well into the spring, reducing the potential for LDS in the spring.

Wetting agents are especially useful in restoring the wettability of hydrophobic (water repellent) sand-based soils. Turfgrass grown on sand-based root zones can develop severe localized dry spots especially

© Bob Searle



when the stand is irrigated deep and infrequently (wet and dry cycles). Wetting agents help promote water infiltration and retention in these hydrophobic areas by reducing the surface tension of water and restoring the polar attraction of water to soils.

In addition to a variety of chemistries available for wetting agent products, natural options to improve water movement in the soil include yucca extracts and gypsum (calcium sulfate).

DROUGHT PLANNING AND RESPONSE

Besides operating the facility in a manner that promotes water conservation, superintendents should identify water-conserving measures in time of severe shortages before water usage restrictions are enacted at a state or local level. Water conservation plans should identify opportunities to achieve a 10%, 30%, and 50% reduction in water use. The USGA publication [BMPs and Water-Use Efficiency/Conservation Plan For Golf Courses: Template and Guidelines](#) can facilitate the creation of the plan. In addition, superintendents should monitor drought status when needed. Drought conditions for each state can be accessed through the National Integrated Drought Information System on the [U.S. Drought Portal](#).

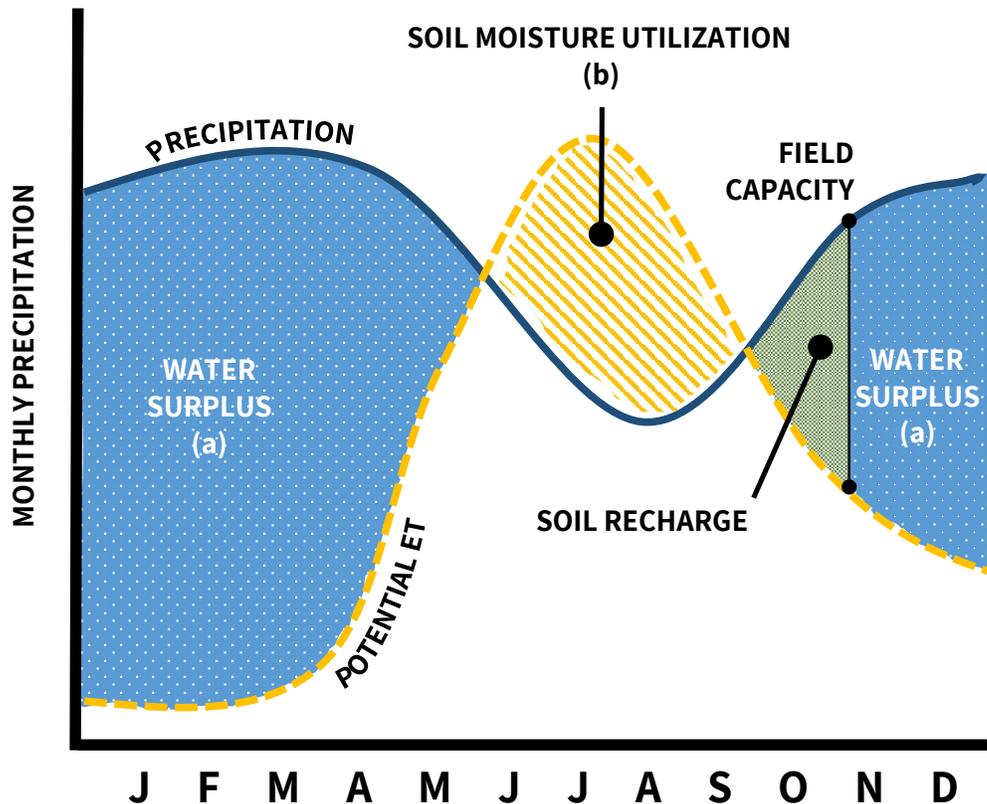
Communication should be maintained with water managers, golf club members, and the public to explain these water conservation efforts as a proactive approach to addressing water-related issues.

Best Management Practices for Water Conservation and Efficient Use Planning

- Develop a water budget for the course.
- Select drought-tolerant varieties of turfgrass to minimize water use.
- Utilize hand watering or targeted irrigation to conserve water.
- Control invasive plants or plants that use excessive water.
- Reduce the amount of area on the golf course that is irrigated, if possible, such as non-play areas.
- Utilize wetting agents to increase soil water uniformity, minimize localized dry spot, and sustain high visual turfgrass quality at very low levels of irrigation.
- Water-in wetting agents sufficiently.
- Identify opportunities to achieve water use reductions before mandatory water restrictions are enacted in times of drought.
- Develop a drought plan for the property based on the USGA Drought Plan Template that includes all aspects of facility management.
- During a drought, monitor the state's drought status to ensure compliance with restrictions.

Irrigation Management Decision-Making

An irrigation system should be operated based only on the moisture needs of the turfgrass -- or to water in a fertilizer or chemical application as directed by the label. Irrigation scheduling must take plant water requirements and soil intake capacity into account to prevent excessive water use that could contribute to leaching and runoff. Plant water needs are determined by several factors including evapotranspiration (ET) demands, recent rainfall, recent temperature extremes, and soil moisture. ET rates and soil moisture replacement should serve as primary factors to help determine the irrigation schedule rather than a calendar-based schedule.



Understanding natural water budgets in your region is critical to proper irrigation. Course superintendents must recognize when they are experiencing a water surplus (a) and deficit (b).

EVAPOTRANSPIRATION

Evapotranspiration describes the water lost through soil evaporation and plant transpiration and is influenced by the climatic conditions such as solar radiation, temperature, and relative humidity. Evaporative demands (and ET) increase with increasing solar radiation, high temperatures, and decreasing relative humidity. The irrigation of turf based on ET replacement is effective in preventing leaching losses and therefore eliminating potential sources of waste, which is an important water conservation strategy. The scheduling of irrigation using ET replacement also helps to quantify water in terms of the “amount” in inches to be applied to turf. Deficit irrigation is another way ET can be used to avoid overwatering and even reduce water consumption. In this approach, turf is irrigated with only a portion of the calculated ET (e.g., 60 to 80% ET) for a period of days until wilt becomes too difficult

to manage, or a rain event occurs. In either case, soil water levels are restored to facilitate turf recovery either through irrigation or natural precipitation. In the latter case, a water savings is accomplished by applying less water daily, allowing for natural rainfall to provide necessary water for turf maintenance. Accuracy of ET calculations, and therefore their potential to reduce water consumption, can be enhanced through the use of crop coefficients (Kc) that account for variations in water use among different plant species. Research conducted at the University of Massachusetts determined Kc values for creeping bentgrass, Kentucky bluegrass, and perennial ryegrass at fairway and rough mowing heights. Detailed information on how to correctly use [ET-based irrigation scheduling](#) for cool-season grasses can be found as part of the Turf Irrigation Series, University of Massachusetts.

It is important to note that because electric/mechanical clocks cannot automatically adjust for changing ET rates, frequent adjustment is necessary to compensate for the needs of individual turfgrass areas using these older systems.

SOIL INFILTRATION RATE AND PLANT AVAILABLE WATER

The rate of infiltration depends on soil texture. Sandy soils, with their higher porosity, have greater infiltration rates than silty or clay soils. Plant available water (PAW) represents the amount of water (expressed in inches) available per inch of soil depth that a plant can access for transpiration. A soil moisture probe indicates the total volumetric water content, which is greater than the PAW for a soil. The PAW can be estimated with a soil moisture meter by subtracting the current soil moisture content from the moisture content when the turfgrass wilts. Plant available soil moisture and infiltration rates are provided in Table 2-1.

Table 2-1. Available soil moisture and infiltration rates for common soil textures.

Soil Texture	Soil Type	Typical plant- available moisture per foot of soil depth (inches)	Infiltration rate (inches h ⁻¹)
Light, sandy	Coarse sand	0.25 – 0.75	Fast (0.5 – 6+)
	Fine sand	0.75 – 1.00	
Medium, loamy	Loamy sand	1.10 – 1.20	Moderate (0.25 – 0.5)
	Sandy loam	1.25 – 1.40	
	Fine sandy loam	1.50 – 2.00	
	Silt loam	2.00 – 2.50	
Heavy, clay	Silty clay loam	1.80 – 2.00	Slow (0.1 – 0.25)
	Silty clay	1.50 – 1.70	
	Clay	1.20 – 1.50	

ROOTZONE DEPTH

The depth of effective turfgrass rooting should be determined with a soil probe or spade. Golf greens and tees have the majority of roots in the top several inches of soil, while fairways and roughs will typically have deeper roots. Exact root depths depend on grass species and time of year. The soil infiltration rate and root zone depth should be used together to estimate the amount of water that needs to be available to the root system to avoid wilting. The rooting depth is multiplied by the PAW to estimate the total amount of water available to the turfgrass.

SOIL MOISTURE

To accurately measure local precipitation, the proper use of rain gauges, rain shut-off devices, soil moisture sensors (especially sensors utilizing time domain reflectometry [TDR] technology), and other irrigation management devices should be incorporated into the site's irrigation schedule. Monitoring of soil moisture, in addition to calculating

ET rates and visual observations of turfgrass, assists in meeting turfgrass water needs while conserving water resources.

Irrigation Scheduling

Proper irrigation can sustain plant energy reserves, increase root mass and depth, and reduce thatch accumulation. Irrigation should be applied as necessary to prevent wilt without oversaturating the soil/rootzone and without compromising playing conditions. In general, it is appropriate to water deeply and infrequently to promote root growth. In some cases, such as when watering-in products, it is important not to water past the rootzone to prevent leaching of products, including fungicides and wetting agents. It is also important to be mindful of how various areas of the course may require a different approach because of soil type (sand-based greens, etc.).

The goal of successful irrigation management is to limit excessive soil moisture while preventing wilt. Golf course managers strive to precisely apply water so PAW is only slightly greater than predicted ET. For many highly maintained turfgrass areas, like greens, small amounts of water are applied every night to replace what was lost the prior day. Soil moisture sensors can help further improve irrigation precision. These technologies can guide irrigation run times and identify locations that might benefit from additional hand watering.

During periods of sufficient natural precipitation, stress pre-conditioning through deficit irrigation can improve tolerance to future drought, heat, and cold stress. Deficit irrigation is the practice of limiting irrigation to slowly deplete soil PAW until the soil moisture approaches wilting points.

Computerized irrigation systems provide many advantages and most Maine courses utilize them. However, non-computerized technologies are still in use on some Maine courses. Computerized systems should allow a superintendent to remotely cancel the program if the course has received adequate rainfall. Wi-Fi controllers connected to weather stations can adjust for changing ET. (Clock-controlled irrigation systems preceded computer-controlled systems; many are still in use today and do not automatically adjust for changing ET rates. Therefore, frequent adjustment is necessary to compensate for the needs of individual turfgrass areas.)

MAINTAINED TURF AREAS

The irrigation system should be designed and installed so that the putting surface, slopes, and surrounding areas can be watered independently. Precision irrigation scheduling of these areas is based on soil infiltration rates, soil water-holding capacity, plant water-use requirements, the depth of the rootzone, and desired level of turfgrass appearance and performance.

NON-PLAY AND LANDSCAPE AREAS

Courses should map any environmentally sensitive areas such as sinkholes, wetlands, or flood-prone areas, and should identify any listed species or habitats of concern.

Natural vegetation should be retained and enhanced for non-play areas to conserve water. The most efficient and effective watering method for non-turf landscape is micro-irrigation. Older golf courses may have more irrigated and maintained acres than are necessary. With the help of a golf course architect, golf course superintendent and other key personnel, the amount of functional turfgrass can be evaluated and transitioned into non-play areas. For more information on non-play area landscaping, see the “Sustainable Landscaping in Out-of-Play Areas” and “Pollinator” chapters of this document.

Best Management Practices for Irrigation Scheduling

- The irrigation system should be designed and installed so that the putting surface, slopes and surrounding areas can be watered independently.
- Install part-circle heads to conserve water and reduce unnecessary stress to greens and surrounds.
- Avoid use of a global setting. Make adjustments to watering times per head based on turf species and soil and slope characteristics.
- Base water times on actual site conditions for each head and zone.
- Adjust irrigation run times based on current local meteorological data.
- Use computed daily ET rate to adjust run times and manually adjust automated ET data to reflect wet and dry areas on the course.
- Install rain switches to shut down the irrigation system if enough rain falls in a zone.
- Use soil moisture sensors to bypass preset irrigation schedules or to create on-demand schedules.
- Permanent irrigation sprinklers and other distribution devices should be spaced according to the manufacturer’s recommendations.
- Spacing should be based on average wind conditions during irrigation.
- Reducing dry spots and soil compaction improves water infiltration, which in turn reduces water use and runoff in other areas.
- When possible, irrigation should occur in the early morning hours before air temperatures rise and relative humidity drops.
- Base plant water needs on ET rates, recent rainfall, recent temperature extremes, and soil moisture.

Best Management Practices for Irrigation Scheduling (continued)

- Use mowing, verticutting, aeration, wetting agents, nutrition, and other cultural practices to maximize infiltration and minimize water loss as runoff.
- Visually monitor for localized dry conditions or hot spots to identify poor irrigation efficiency or a failed system device.
- Install inground (wireless) soil moisture sensors in the rootzone and use in conjunction with handheld moisture meters for each irrigation zone to enhance scheduled, timer-based run times.
- Wireless soil moisture systems should also be installed to prevent damage from aeration. (Wired-in soil moisture systems can also be installed but require close monitoring to prevent damage from aeration.)
- Designate 50% to 70% of the non-play area to remain in natural cover according to “right-plant, right-place” principle of plant selection that favors limited supplemental irrigation and on-site practices.
- Incorporate natural vegetation in non-play areas.
- Use micro-irrigation and low-pressure emitters in non-play areas to supplement irrigation.
- Routinely inspect non-play irrigation systems for problems related to emitter clogging, filter defects, and overall system functionality.

The Ledges Golf Course, York, ME. © Forrest Bell



Irrigation Water Sources

Golf course designers and managers should endeavor to manage water resources to minimize impacts on freshwater drinking supplies, while also promoting plant health and protecting the environment. Studies of water supplies are recommended for irrigation systems, as are studies of waterbodies or flows on, near, and under the property. Water sources for irrigation must be dependable and offer sufficient resources to accommodate turf grow-in needs and ongoing maintenance. [Environmental Best Management Practices for Virginia's Golf Courses](#) describes the methodology and provides example calculations to determine water requirements using a seasonal and maximum bulk water requirement analysis (see pages 37 and 38).

The opportunity to identify and use alternative water supply sources may also be appropriate, depending on the availability of infrastructure and additional management costs associated with non-potable water.

When using groundwater, the area around the wellhead should be protected, and safe land-use practices should be instituted to protect aquifers from accidental contamination. This includes protecting wellheads from physical impacts, keeping them secure, and sampling wells according to the monitoring schedule required by the regulating authority. Before installing new wells, the local regulatory authorities should be contacted to determine the permitting and construction requirements and the isolation distances required from potential sources of contamination. New wells should be located up-gradient as far as possible from potential pollutant sources, such as petroleum storage tanks, septic tanks, chemical mixing areas, or fertilizer storage facilities.

Best Management Practices for Irrigation Water Sources

- Identify appropriate water supply sources that meet seasonal and bulk water allocations for grow-in and routine maintenance needs.
- When developing new water sources, incorporate surface storage (lined ponds) with wellhead withdrawals to conserve water by conservation of rainfall, site drainage, and runoff as a supplemental water source.
- Use alternative water supplies/sources that are appropriate and sufficiently available to supplement water needs and follow guidelines for use.
- Ensure that reclaimed, effluent, and other non-potable water supply mains have a thorough cross-connection and backflow prevention device in place and are operating correctly. Adhere to cross-connection regulations.
- Post signs in accordance with local utility and state requirements when reclaimed water is in use.
- Surround new wells with bollards or a physical barrier to prevent impacts to the wellhead.
- Maintain records of new well construction and modifications to existing wells.

Best Management Practices for Irrigation Water Sources (continued)

- Obtain a copy of the well log for each well to determine the local geology and well depth. These factors will have a bearing on how vulnerable the well is to contamination. Sample wells for contaminants according to the schedule and protocol required by various state agencies.
- Inspect wellheads and the well casing at least annually for leaks or cracks. Make repairs as needed.
- Use backflow-prevention devices at the wellhead, on hoses, and at the pesticide mix/load station to prevent contamination of the water source. Adhere to various state cross-connection regulations.
- Properly plug abandoned or flowing wells.
- Never apply a fertilizer or pesticide next to or near a wellhead.
- Never mix and load pesticide next to or near a wellhead if not on a pesticide mix/load pad.

Irrigation Water Suitability

Irrigation water quality must be suitable for plant growth and pose no threat to public health. Because water quality can influence soil quality and turfgrass performance, it is advisable to test irrigation water periodically for factors that can compromise the turf/soil system. This is especially true for non-potable water irrigation sources, such as retention ponds and recycled water. Excess nutrients and/or salts may accumulate to levels that are toxic to plants -- potentially influencing aquatic plant growth in rivers, lakes, and estuaries -- and that contribute to a variety of soil-related problems. For example, irrigation water high in sodium and low in calcium and magnesium applied frequently to clay soils can break down soil structure, cause precipitation of organic matter, and reduce permeability.

Routine analysis should provide the following information: conductivity, pH, Na, Ca, Mg, K, CO_3^{2-} , HCO_3^- , SO_4^{2-} , Cl^- , P, B, nitrate-N, hardness, and sodium adsorption ratio (Landschoot, 2016). The results can be used to address possible issues with soil salinity and plant health caused by poor water quality.

When necessary, water system treatment options should be included in the operating budget to address water quality and equipment maintenance.

Best Management Practices for Water Suitability

- Account for the nutrients in effluent (reuse/reclaimed) water when making fertilizer calculations.
- Test reclaimed water regularly for dissolved salt content.
- Routinely monitor the shallow groundwater table of fresh water for saltwater intrusion or contamination by heavy metals and nutrients.
- Flush with fresh water or use amending materials regularly to move salts out of the root zone and/or pump brackish water to keep salts moving out of the root zone.
- Amend sodic water systems appropriately (with gypsum or an appropriate ion) to minimize sodium buildup in soil.
- Monitor sodium and bicarbonate buildup in the soil using salinity sensors or routine soil tests.
- When drilling supplemental wells adjacent to coastal or brackish waters, consult with a hydrologist to minimize salt water contamination of staging ponds or holding systems.

Irrigation System Design

SITE ASSESSMENT

An assessment of the facility should be conducted prior to developing an irrigation system design. The assessment should include site-specific features, such as water sources, soil types (see the [Web Soil Survey](#)), soil physical properties, microclimates, slopes, sun, wind, and shade exposures, and a seasonal and bulk water requirement analysis.

The site assessment should also evaluate the impact of design elements, such as design features and concepts, planned or existing turfgrass varieties, and planned or existing drainage systems. The system design should include a general irrigation schedule with recommendations and instructions on modifying the schedule to meet these site-specific needs.

DESIGN CONSIDERATIONS

A well-designed irrigation system should operate at peak efficiency and be designed and installed to optimize water use efficiency, focusing on water placement and distribution. The design should maximize water use, reduce operational cost, conserve supply, and protect water resources. Detailed BMPs for irrigation system design are published by the Irrigation Association in [2014 Landscape Irrigation Best Management Practices](#).

The irrigation system design should meet the site-specific needs identified by the water quantity analyses and the site assessment. The system's capacity to deliver water should not exceed the infiltration of the soils on site, as that will lead to runoff. Though the design of an irrigation system is

complex, some of the most important design decisions that influence the efficiency and effectiveness of water usage include those related to sprinkler and piping placement, sprinkler coverage and spacing, and communication options.

SPRINKLERS

Multi-row sprinkler systems provide the most efficient use of water and can respond to specific moisture requirements of selected areas. Newer designs with multiple nozzle configurations provide increased flexibility and improved distribution uniformity. Single row systems do not uniformly distribute water and increase the risk of runoff. Double- row systems offer improved efficiency over single-row coverage, although manual watering or other types of supplemental watering may be needed outside the fairway area and into the extended rough. Sprinkler layouts can be specific to each area. For example, part-circle sprinklers can be arranged to avoid overspray of impervious surfaces and to apply water only to the green surface or in heavy traffic areas. Manual quick-coupler valves can be an important conservation element and should be installed near greens, tees, and bunkers so these can be hand-watered during severe droughts. Irrigation systems strive to provide uniform water distribution and to achieve distribution uniformity (DU) values near 80%. After installation, nozzles and irrigation head runtimes should be optimized to maintain uniform soil moisture distribution. That can be easily monitored with a soil moisture probe.

COMMUNICATION

For precise irrigation control, advanced irrigation control systems that schedule each green, tee, and fairway separately allow course managers to adjust for differences in microclimates and root zones. Weather stations that calculate and automatically program water replacement schedules also provide opportunities for more precise irrigation, as do soil moisture sensors placed in multiple locations. Additional features may include internet connectivity to allow for remote adjustment or cancellation of schedules as well as rain stop safety switches that either shut down the system in the event of rain or adjust schedules based on the amount of precipitation.

Best Management Practices for Irrigation System Design

- Seek assistance from irrigation professionals, such as from Certified Golf Course Irrigation System designers and WaterSense-certified irrigation consultants.
- New and upgraded irrigation system designs should deliver water with maximum efficiency, focusing on precision water placement and distribution.
- Design and/or maintain a system to meet a site's peak water requirements under normal conditions with the flexibility to adapt to extreme conditions or local restrictions.
- Design should account for optimal distribution efficiency and effective root-zone moisture coverage. Target 80% or better DU.
- Design should allow the putting surface, slopes, and surrounds to be watered independently.

Best Management Practices for Irrigation System Design (continued)

- The design package should include a general irrigation schedule with recommendations and instructions on modifying the schedule for local climatic, soil, and growing conditions. It should include the base ET rate for the particular location.
- The application rate must not exceed the infiltration rate, which is the ability of the soil to absorb and retain the water applied during any one application. Conduct saturated hydraulic conductivity tests periodically.
- The design operating pressure must not be greater than the available source pressure.
- The design operating pressure must account for peak-use times and supply-line pressures at final buildout for the entire system.
- Turf and landscape areas should be zoned separately. Specific-use areas that should be zoned include greens, tees, primary roughs, secondary roughs, fairways, trees, shrubs and flower beds.
- Design should account for the need to leach out salt buildup from poor-quality water sources by providing access to fresh water.
- Permanent irrigation sprinklers and other distribution devices should be spaced according to the manufacturer's recommendations.
- Spacing should be based on average wind conditions during irrigation.
- Distribution devices and pipe sizes should be designed for optimal uniform coverage and flow rate.
- Distribution equipment, such as sprinklers, rotors, and micro-irrigation devices, in a given zone must have the same precipitation rate.
- Heads for turf areas should be spaced for head-to-head coverage.
- Water supply systems (for example, wells and pipelines) should be designed for varying control devices, rain shut-off devices, and backflow prevention.
- Water conveyance systems should be designed with thrust blocks and air-release valves.
- Flow velocity must be 5 feet per second or less.
- Pipelines should be designed to provide the system with the appropriate flow and pressure required for maximum irrigation uniformity.
- Pressure-regulating or compensating equipment must be used where the system pressure exceeds the manufacturer's recommendations.
- Equipment with check valves must be used in low areas to prevent lowhead drainage.
- Isolation valves should be installed in a manner that allows critical areas to remain functional.

Best Management Practices for Irrigation System Design (continued)

- Manual quick-coupler valves should be installed near greens, tees, and bunkers and in fairways if possible, so that these areas can be hand-watered during severe droughts.
- Use part-circle or adjustable heads to avoid overspray of impervious areas, such as roadways and sidewalks, and surface waters such as lakes, ponds, and wetland margins.
- Update block type sprinkler valves with single head control to conserve water and to enhance efficiency.
- Incorporate multiple nozzle configurations to add flexibility and enhance efficiency and distribution.
- Ensure heads are set level to the ground.
- Provide backup option(s) for loss of power/pumps with PTO, gas/diesel, potable pump or generator to provide irrigation to greens and tees at a minimum.

Pump Stations

Pump stations should be efficient and sized to provide adequate flow and pressure. They should be equipped with control systems that protect distribution piping, provide for emergency shutdown necessitated by line breaks, and allow maximum system scheduling flexibility.

Where feasible, variable frequency drive (VFD) pumps and/or pump station should be used. These systems only expend enough energy to meet the demands of the irrigation pump(s). VFD systems reduce water hammer to fitting, pipe, and sprinklers when systems are pressurized.

Best Management Practices for Pump Stations

- The design operating pressure must account for peak-use times and supply-line pressures at final buildout for the entire system.
- Maintain the air-relief and vacuum-breaker valves by using hydraulic pressure- sustaining valves.
- Install VFD systems to lengthen the life of older pipes and fittings until the golf course can afford a new irrigation system.
- An irrigation system should also have high- and low-pressure sensors that shut down the system in case of breaks and malfunctions.
- Pumps should be sized to provide adequate flow and pressure.
- Pumps should be equipped with control systems to protect distribution piping.

Irrigation System Installation

To ensure maximum efficiency, the irrigation system must be installed per the design and specifications. The installer must ensure that there is qualified supervision of the installation process and that a qualified irrigation specialist inspects and approves the system installation.

Best Management Practices for Irrigation System Installation

- The designer must approve any design changes before construction.
- Construction and materials must meet existing standards and criteria.
- Prior to construction, all underground cables, pipes, and other obstacles must be identified and their locations flagged ([Dig Safe 811](#)).

Irrigation System Maintenance and Performance

CALIBRATION AND AUDITING

Irrigation system maintenance on a golf course involves four major efforts: calibration and auditing, preventive maintenance, corrective maintenance, and record keeping. Personnel charged with maintaining a golf course irrigation system face numerous challenges. This is particularly true for courses with older or outdated equipment.

Irrigation audits can be conducted to assess the system function, ensuring that the irrigation system works reliably and is cost effective. The Irrigation Association has published [irrigation audit guidelines](#).

MAINTENANCE

Good system management starts with good preventive maintenance procedures and record keeping. Corrective maintenance is simply the act of fixing what is broken and may be as simple as cleaning a clogged orifice or as complex as a complete renovation of the irrigation system. As maintenance costs increase, an evaluation of whether a system renovation is needed should be conducted.

WINTERIZATION

Winterizing protects irrigation system pipes from damage due to water expanding and rupturing the pipe walls and fittings. Golf courses need to drain or use compressed air to remove water from lateral and mainline pipes before temperatures drop below freezing, as well as from all sprinkler heads and quick couplers. Many facilities operate an independent irrigation system below the frost line, allowing the

facility to apply water during cold periods to dormant turfgrass in an effort to prevent winter desiccation and winter kill.

SPRING START-UP

Spring start-up of the irrigation system is essentially the reverse of the steps taken to winterize the system. At the time of start-up, the system should be inspected for corrective maintenance issues.

Best Management Practices for Irrigation System Maintenance – Calibration and Auditing

- Examine turf quality and plant health for indications of irrigation malfunction or the need for scheduling adjustments.
- Evaluate pressure and flow to determine that the correct nozzles are being used and that the heads are performing according to the manufacturer's specifications.
- Visually inspect the entire system to identify necessary repairs or corrective actions and make repairs before carrying out other levels of evaluation.
- Conduct an annual irrigation audit to facilitate a high-quality maintenance and scheduling program for the irrigation system.
- Conduct an annual pump test and inspections.
- Submit required withdrawal reports to the appropriate regulatory agency.

Best Management Practices for Irrigation System Maintenance – Preventive Maintenance

- Inspect the system daily for proper operation by checking computer logs and visually inspecting the pump station, remote controllers, and irrigation heads. A visual inspection should be carried out for leaks, misaligned or inoperable heads, and chronic wet or dry spots so that adjustments can be made.
- Observe the system in operation regularly to detect controller or communication failures, stuck or misaligned heads, and clogged or broken nozzles.
- Check filter operations frequently. Keeping filters operating properly prolongs the life of an existing system and reduces pumping costs.
- Monitor the power consumption of pump stations for problems with the pump motors, control valves, or distribution system.
- Increase frequency of routine inspection/calibration of soil moisture sensors that may be operating in high-salinity soils.
- Inspect irrigation pipes and look for fitting breaks caused by surges in the system.
- Install thrust blocks to support conveyances.

- Maintain air-relief and vacuum-breaker valves.
- Have qualified pump personnel perform regular checks of amperage to accurately identify increased power usage that indicates potential problems.
- Check application/distribution efficiencies annually.

Best Management Practices for Irrigation System Maintenance – Winterization

- Flush and drain above-ground irrigation system components.
- Remove water from all conveyances and supply and distribution devices that may freeze. Use compressed air or open the drain valves at the lowest point on the system.
- Change filters, screens, and housing; remove drain plugs and ensure any water is removed from the system. Secure systems and close and lock covers/compartments doors to protect the system from vandalism and from animals seeking refuge.
- Drain any above-ground pump casings that may have “trapped” water.
- Record metering data before closing the system.
- Secure or lock any remote irrigation components, including satellites.
- Perform pump and engine servicing/repair before winterizing.

Best Management Practices for Irrigation System Maintenance – Spring Start Up

- Power up the pump station and pump motors before prior to using the system. By completing this task ahead of recharging the system, the coils inside the motors heat up and remove any moisture that collected during the offseason.
- Keep the water pressure at 60 PSI or lower when priming the lines.
- Operate each of the sprinklers until all excess air is flushed from the irrigation system.
- Check the functionality of air pressure relief valves.
- Inspect the entire system for any corrective maintenance issues.

Metering

Rainfall may vary from location to location on a course; the proper use of rain gauges, rain shut-off devices, flow meters, soil moisture sensors, and/or other irrigation management devices should be incorporated into the site's irrigation schedule. It is also important to measure the amount of water that is actually delivered through the irrigation system, via a water meter or a calibrated flow-measurement device. Knowing the flow or volume will help determine how well the irrigation system and irrigation schedule are working.

Best Management Practices for Metering

- Calibrate equipment periodically to compensate for wear in pumps, nozzles, and metering systems.
- Properly calibrated flow meters, soil moisture sensors, rain shut-off devices, and/or other automated methods should be used to manage irrigation.
- Flow meters should have a run of pipe that is straight enough – both downstream and upstream – to prevent turbulence and bad readings according to manufacturer's requirements.
- Flow meters can be used to determine how much water is applied.

Irrigation Leak Detection

Irrigation systems are complex systems that should be closely monitored to ensure leaks are quickly detected and corrected. An irrigation system should also have high flow, low-pressure sensors that shut down the system in case of breaks and malfunctions. Golf courses without hydraulic pressure-sustaining valves are much more prone to irrigation pipe and fitting breaks because of surges in the system, creating more downtime for older systems.

Best Management Practices for Irrigation Leak Detection

- Monitor water meters or other measuring devices for unusually high or low readings to detect possible leaks or other problems in the system. Make any needed repairs.
- Monitor the system daily for malfunctions and breaks. Log water usage daily.
- Ensure that control systems provide for emergency shutdowns caused by line breaks and allow maximum system scheduling flexibility.

Irrigation System Renovation

Renovating a golf course irrigation system can improve system efficiencies, conserve water, improve playability, and lower operating costs.

Best Management Practices for Irrigation System Renovation

- Determine the age of the system to establish a starting point for renovation.
- Identify problems and their costs to determine which renovations are appropriate.
- Identify system performance improvements that maximize the efficient use of the current system.
- Evaluate the cost of renovation and its return on financial and management benefits.

Irrigation Record Keeping

Careful record keeping is an important part of managing an irrigation system, as well as part of regulatory requirements for reporting water withdrawal.

Best Management Practices for Record Keeping

- Keep records of filter changes, as this could be an early sign of system corrosion, well problems, or declining irrigation water quality.
- Document equipment run-time hours. Ensure that all lubrication, overhauls, and other preventive maintenance are completed according to the manufacturer's schedule.
- Monitor and record the amount of water being applied, including system usage and rainfall and identify areas where minor adjustments can improve performance.
- Document and periodically review the condition of infrastructure, such as pipes, wires, and fittings. If the system requires frequent repairs, it is necessary to determine why these failures are occurring.
- Document all corrective actions.
- Adhere to all regulatory reporting requirements for water withdrawal.



CHAPTER 3: WATER QUALITY MANAGEMENT AND PROTECTION

Riverside Golf Course, Portland, ME © FBE

PREFACE

Protecting water quality involves recognizing the potential fate and transport mechanisms that can carry contaminants into water resources. Stream channel and habitat alteration can also negatively impact the health of aquatic communities, such as the macroinvertebrate and fish communities. If water quality contaminants reach surface or groundwater or stream alteration occurs, the potential water quality impacts can include:

- Drinking water impairment if nitrogen as either nitrate (NO_3) or nitrite (NO_2) exceeds risk values that may adversely affect health.
- Nutrient enrichment of surface water.
- Sedimentation due to eroding soils.
- Toxicity to aquatic life.

- Channel destabilization and erosion.
- Bank failure.
- Loss of habitat diversity and riparian cover and shade.
- Loss of adjacent habitat for oviposition¹ by the adult stages of aquatic insects.

Therefore, aligning water quality management efforts, such as stormwater management and lake and pond management with established, research-based BMPs protects water quality. A water quality monitoring program can be used to verify that water resources are being adequately protected. In addition, monitoring may demonstrate the presence of issues in water before entering a golf course such as upstream contamination.

Overall, protecting water quality includes not only implementing what is contained in this chapter, but what is discussed throughout this document, including:

- Design considerations such as the use of vegetated buffers.
- Fertilizer use (and phosphorus management) and fertilization strategies near surface waters. (Covered in more detail in Chapter 5.)
- Pesticide usage. (Covered in more detail in Chapter 8.)

Environmental Fate and Transport

Understanding contaminant fate and transport mechanisms will help superintendents protect water quality by minimizing the risk of off-movement of nutrients and pesticides applied to golf courses.

Sources of pollution can include:

- Runoff, or the movement of water across the turf and soil surface, typically following a storm event or heavy irrigation. The potential for runoff is greatest on steep slopes and on soils with low infiltration capacity – fine textured or compacted soils and soils with a shallow restrictive layer and/or a high seasonal water table.
- Leaching, or the downward movement of water through the soil and potentially into groundwater. Several variables influence the probability and rate of leaching, such as soil type and structure, vegetation, chemical properties, rate of precipitation, and depth to groundwater.
- Spray drift, or the movement of fine particles, or droplets, through the air while the pesticide is being applied. Droplet size and wind and weather conditions affect the potential for spray drift during pesticide applications.
- Vapor drift, or the movement of pesticide in the form of a gas or vapor during or after application. Pesticide formulation, wind, and atmospheric conditions affect the potential for vapor drift during pesticide applications.
- Volatilization, or the transformation of a pesticide from a solid or liquid to a gas or vapor after a

¹ *Oviposition refers to the process of expulsion of eggs from the oviduct to the external environment.*

pesticide application. Once airborne, volatile pesticides can come into contact with pesticide applicators or move long distances off-site.

- Spills, or unintended releases of chemicals, such as fertilizers, pesticides, hazardous materials, or petroleum products during transportation, storage, routine maintenance, and facility operations.

While most of the fate and transport mechanisms of concern can contribute to nonpoint sources of pollution, spills can be a point source of pollution. On golf courses, point sources of pollution can originate from:

- Storage and maintenance facilities.
- Unintended release of chemicals, such as pesticides, fertilizers, or fuel, during transportation, storage, or handling
- Drainage discharge outlets (e.g., the end of a drainage pipe).

Containment measures can easily prevent chemicals from becoming point sources of pollution during storage and handling. To prevent contamination of surface or groundwater water, any accidental spills of chemicals must be diverted from surface waters and contained and cleaned-up according to state guidelines.

One additional potential contaminant is sediments, though primarily only a concern when bare soil is exposed, as during construction or renovation. Sedimentation is a concern when precipitation and irrigation carry soil particles (sediment) in runoff and deposit them into surface water. Sedimentation can also occur in downstream reaches as a result of channel or bank erosion caused by elevated storm

A silt fence prevents stormwater runoff from carrying sediment offsite. © Bob Searle



flows, stream channel alterations (intentional or unintentional) that destabilize banks, and undersized or misaligned culverts. Too much sediment can cloud surface water, reducing the amount of sunlight that reaches aquatic plants and impairing aquatic species habitat. In addition, sediments can carry fertilizers, pesticides, and other chemicals attached to soil particles and transport them into waterbodies, causing algal blooms that lead to oxygen depletion. Sedimentation is handled through BMPs that control the volume and flow rate of runoff water, maintain adequate turf density, reduce soil transport, and ensure perennial and intermittent stream channels are not altered and remain in their natural condition.

Pollution Prevention

Implementing BMPs can prevent or minimize the effects of a golf course on surface and groundwater, while ensuring and even enhancing public health and environmental quality. Pollution prevention is easier, less expensive, and more effective than addressing problems after they happen. An integrated water quality protection system that incorporates the BMPs found throughout this document is based on a tiered concept as follows:

- Prevention – Stopping problems from occurring.
- Control – Having safeguards in place to handle any problems.
- Detection – Using a monitoring program to detect changes in environmental quality.

At any golf facility, preventive strategies should include combinations of land use controls and source prevention practices. Land use BMPs are engineered and incorporated into the course during golf course design and construction, and can be added and altered throughout the golf course's lifetime as needed. They protect natural resources through primarily mechanical methods, such as retention basins, vegetated swales, and buffer areas around water courses.

Source prevention BMPs are implemented during golf course management operations to prevent sediment, nutrients, or pesticides from being introduced into ecologically sensitive areas. For example, pesticide management BMPs reduce the potential for drift and volatilization during pesticide applications. Irrigation BMPs prevent over-watering and are especially important for minimizing pollutant transport via runoff or leaching.

Cultural practice BMPs maximize the water infiltration and water holding capacity of soils. Safeguards should be incorporated into the facility management to control any problems should they arise, to prevent the contamination of water from spills. For example, many of the BMPs related to pesticide storage and handling as well as maintenance operations can prevent accidental releases of contaminants (pesticides, fertilizers, fuel, etc.) from becoming a source of contamination.

Aligning golf course management practices with BMPs protects water quality on and downstream from the facility. Water quality monitoring on golf courses requires careful consideration of the frequency, duration, and timing of sampling (see following chapter for details). Surface and groundwater chemistries are dynamic and respond to changes in seasonal and daily weather patterns. It is important to consult with a professional regarding the water quality monitoring program to ensure that collected data is reliable and comparable across conditions. Furthermore, providing monitoring information to local, regional, and state regulatory authorities and watershed groups can help foster positive relationships with these stakeholders.

Stormwater Management

Runoff, or the movement of water across the land surface from either precipitation or irrigation that does not infiltrate into the ground, is the conveying force behind nonpoint source pollution. Stormwater management refers to runoff from precipitation but applies to irrigation runoff as well. Stormwater management is the control and use of runoff and includes planning for runoff, maintaining stormwater systems, and regulating the collection, storage, and movement of stormwater.

BMPs reduce stormwater volume, peak flow, and nonpoint source pollution by promoting evapotranspiration, infiltration, detention, and filtering, as well as biological and chemical actions. BMPs help achieve such goals by:

- Keeping stormwater close to where it falls.
- Slowing down stormwater runoff.
- Allowing stormwater to infiltrate into the soil.

Stormwater management is best accomplished by a “treatment train” approach in which water is moved from one treatment to another by conveyances that themselves contribute to the treatment. These treatments include source controls, structural controls, and non-structural controls. An example of this treatment train approach is as follows: Stormwater is directed across vegetated filter strips, through a swale, into a retention pond, then out through another swale to a constructed wetland system.

THE STORMWATER TREATMENT TRAIN



A stormwater treatment train directs stormwater runoff from one treatment to another by conveyance structures. Each “train” provides treatment to the stormwater flowing through it. **1 Source controls** prevent the generation of polluted runoff, **2 structural controls** remove, filter, retain, or reroute potential contaminants, and **3 non-structural controls** mimic the natural hydrology before stormwater leaves the site.

SOURCE CONTROLS

The first car of the BMP treatment train are source controls to help prevent the generation of stormwater runoff or the introduction of pollutants into stormwater runoff. For example, during construction or redesign activities, strict adherence to erosion and sedimentation controls helps to prevent, or at least minimize, the possibility for sediment and nutrients to impact water quality through runoff. After construction, implementation of BMPs can reduce the potential for off-site movement of contaminants such as nutrients and pesticides.

STRUCTURAL CONTROLS

Structural controls are often the next car in the treatment train and are design and engineering features on the course created to remove, filter, retain, or reroute potential contaminants (e.g., nutrients, pesticides, sediments) carried in surface runoff. They may also be combined to increase the treatment of stormwater. For example, sediment forebays can be used to pretreat stormwater before it is discharged to a dry extended detention basin, wet basin, constructed stormwater wetland, or infiltration basin.

Periodic inspection and maintenance of all structural controls are essential to ensure they function as designed. Maintenance includes periodic cleaning of small basins, ponds, and forebays to remove sediments. The disruption and financial outlay of this effort is less than that for dredging an entire body of water.

In and around the clubhouse and other structures, opportunities should be identified to slow down the movement of water from impervious surfaces and allow for infiltration. For example, runoff from gutters and roof drains should flow into permeable areas. Rain gardens near these areas can be incorporated into the landscape design. Maximizing the use of pervious pavements, such as brick or concrete pavers separated by sand and planted with grass, allows stormwater to infiltrate into the soil as opposed to running off. Crushed stone and other permeable products are available for cart paths or parking lots.

NON-STRUCTURAL CONTROLS

Non-structural controls are the last car in the treatment train. Non-structural controls often mimic natural hydrology (e.g., constructed wetlands), hold stormwater, and filter stormwater via vegetative practices (e.g., filter strips and grassed swales). Turfgrass areas are extremely effective in reducing soil losses compared with other cropping systems, due to the architecture of the turf canopy, the fibrous turf root system, and the development of a vast macropore soil structural system that encourages infiltration rather than runoff. Additionally, turf density, leaf texture, rooting strength, and canopy height physically restrain soil erosion and sediment loss by dissipating impact energy from rain and irrigation water droplets. If available, diversion of runoff in well distributed, unchanneled flow into adjacent natural forest vegetation with undisturbed duff layer and ground cover vegetation can provide substantial infiltration.

Best Management Practices for Stormwater Management

- Design stormwater control structures to hold stormwater for appropriate residence times to help remove total suspended solids (TSS). Effective phosphorus and nitrogen removal in a pond system require a multi-week residence time to allow sufficient time for sedimentation of fine particles and conversion of dissolved nutrients to biomass for sedimentation.
- Use a stormwater treatment train (in which water is conveyed from one treatment to another by conveyances that themselves contribute to the treatment).
- Use vegetated swales to slow and infiltrate water from low intensity storms and trap pollutants in the soil, where they can be naturally broken down by soil organisms.

Best Management Practices for Stormwater Management (continued)

- Maximize the use of pervious pavements, such as brick or concrete pavers separated by sand and planted with grass. (Special high-permeability concrete and asphalt products are available for cart paths or parking lots.)
- Minimize directly connected impervious areas to the extent practical.
- Disconnect runoff from gutters and roof drains from impervious areas, so that it flows onto permeable areas that allow the water to infiltrate near the point of generation.
- Use depressed landscape islands in parking lots to catch, filter, and infiltrate water, instead of letting it run off. When hard rains occur, an elevated stormwater drain inlet allows the island to hold the treatment volume and settle out sediments, while allowing the overflow to drain.
- Ensure that no untreated discharges from pipes go directly to waterbodies.

Contained fuel storage prevents any fuel spills from running into nearby surface waters. © Bob Searle



Buffers

Buffers around the shore of surface waters, wetlands, or other sensitive areas filter runoff as it passes across. Buffers are the last line of defense to keep sediment out of streams and to filter out fertilizers and pesticides that might otherwise reach waterways.

Depending upon site-specific conditions, including the amount of available space and in-play versus out-of-play considerations, a range of buffer widths can be considered.

Buffer widths as narrow as 10 feet have been shown to be effective. In most cases, a buffer of at least 100 feet is necessary to protect water quality. Smaller buffers (toward the lower end of this range) still afford some level of protection to the surface waters and are preferable to no buffer at all. Protection of the biological components, such as pool-breeding amphibians and other animals, of wetlands and streams typically requires significantly greater buffer widths.

For vegetated buffer zones, ornamental grasses, wetland plants, or emergent vegetation around the perimeter and edges of surface waters serve as a buffer and wildlife habitat for many aquatic organisms and can be aesthetically pleasing. Use native plants for these plantings whenever possible. See the “Landscaping” chapter for more guidance on plant selection and the benefits of utilizing native plants. Riparian buffers along streams and rivers can be up to three different plant assemblages, progressing from sedges and rushes along the water’s edge to upland species.

Riparian buffers of sufficient width intercept sediment, nutrients, and pesticides in surface runoff and reduce nutrients and other contaminants in shallow subsurface water flow. Woody vegetation in buffers provides food and cover for wildlife, stabilizes stream banks, and slows out-of-bank flood flows. It is important that water reaching riparian buffers enters as non-channelized flow (or is dispersed using BMPs such as level spreaders). Natural riparian buffers should be preserved during course design and construction to eliminate the need for less-effective BMPs in the future.

Buffer strip along a tributary at Cape Arundel Golf Club, Kennebunkport, ME. © Brendan Parkhurst



Best Management Practices for Buffers

- Maintain healthy turf cover and woody riparian buffers adjacent to surface waters to slow sediment accretion and reduce runoff flow rates.
- Vary the width, height, and type of vegetation to meet the specific functions of the buffer and growing conditions at the specific location.
- Encourage clumps of native emergent vegetation at shorelines.
- Mow in-play riparian areas to heights as high as possible (a height of 6 inches or greater is preferred).
- When mowing near buffer areas, return clippings away from the water or collect them (such as for composting in a designated area) so that runoff does not carry vegetation into the water.
- Pesticides may not be applied within 25 feet of wetlands, water bodies, or waterways in the state of Maine.² Keep all chemical applications 25 or more feet away from the water's edge when using rotary spreaders and/or boom sprayer applications.
- When fertilizers or pesticides are needed in the buffer area, spot treat weeds or use drop spreaders or shielded rotary spreaders and boom sprayers to minimize the potential for direct deposit of chemicals into the water.

Wetlands

Natural wetlands serve as filters for pollutant removal and as habitat for many species of birds, insects, fish, and other aquatic organisms, and best management practices for wetland protection are important to protect wetland functionality. Natural wetlands on the property should be identified and delineated by a consulting wetland scientist. Vegetated buffers around the shore of a waterbody or another sensitive area slow, filter, and purify runoff before it can reach surface waters as long as the flow is not channelized. Buffers may increase infiltration and ground water recharge. Existing wetlands on golf course properties should be maintained as a protected area and separated from managed turf areas with native vegetation or structural buffers. Stormwater detention and treatment should be applied prior to discharging stormwater into a natural resource such as a wetland, pond, lake, or stream.

Constructed or disturbed wetlands may require a permit to be an integral part of the stormwater management system.

² https://www.maine.gov/dacf/php/pesticides/documents2/rules/BPC_Rules_11-13-15.pdf, chapter 29, page 3.

Best Management Practices for Wetlands

- Protect and maintain existing vegetation as natural buffers, to the maximum extent possible, during new course design and construction or during course renovation or general maintenance. In Maine, required natural buffer widths depend on local shoreland zoning regulations, the size of the waterbody or waterway, and proximity to the waterbody or wetland. Buffers should be, at minimum 10 feet wide, though ideally, 100 feet or greater to protect water quality.
- Develop, enhance, restore, and protect wetland buffers. Manmade buffers should be designed to improve habitat diversity and include a mixture of fast and slow-growing native trees, shrubs, or grasses to provide a diverse habitat for wildlife. A good mixture of vegetation types also provides a mix of early and late growing plants and a variety of rooting depths which protect the soil and provide a better filter.
- Encourage robust coastal and riparian vegetated buffers along the banks of golf course wetlands, perimeters of storage ponds and other waterbodies, and undeveloped uplands.
- Reduce the frequency of mowing at a waterbody edge. Take clippings to upland areas.

Lake and Pond Management

The management of lakes and ponds should include a clear statement of goals and priorities to guide the development of the BMPs necessary to meet those goals. When working near water resources, course superintendents should contact the Maine DEP to identify necessary permitting or licensing requirements. Some of the particular issues superintendents may need to address to maintain the water quality of golf course lakes and ponds include:

- Pond design.
- Dissolved oxygen (DO) levels.
- Aquatic plant management.
- Near-shore management zones.

POND LOCATION AND DESIGN

Designing a new pond requires considerations such as the size of the drainage area, water supply, soil types, and water depth. In addition to potentially serving as an irrigation water source, ponds support aquatic life. The construction of ponds should consider the needs of aquatic ecosystems, such as discouraging excessive growth of aquatic vegetation and the DO needs for aquatic species. In addition, wet retention ponds should be built to meet Maine DEP stormwater requirements and should incorporate proper outlet design to reduce water temperature prior to discharging to downstream waterbodies. One example of this is a gravel cooling discharge trench which cools the water prior to discharge, preventing negative effects on downstream resources, such as small biota that rely on cold water temperatures to survive. Careful design may significantly reduce future operating expenses for

pond and aquatic plant management.

DISSOLVED OXYGEN

Dissolved oxygen is the amount of oxygen present in water and is measured in milligrams per liter (mg/L). Adequate DO levels are required to sustain life in aquatic organisms and vary by species, the organism's life stage, and water temperature.

The amount of DO that water can hold depends on the physical conditions of the body of water (water temperature, rate of flow, oxygen mixing, etc.) and photosynthetic activity. Dissolved oxygen levels also differ by time of day and by season as water temperatures fluctuate, with warm water holding less DO than colder water. Similarly, a difference in DO levels may occur at different depths in deeper surface waters if the water stratifies into thermal layers. Fast-flowing streams hold more oxygen than impounded water. Lastly, photosynthetic activity also influences DO levels. As aquatic plants and algae photosynthesize during the day, they release oxygen. At night, photosynthesis slows down considerably or even stops, and algae and plants pull oxygen from the water. In impoundments with excessive plant and algae growth, several cloudy days in a row can increase the potential for fish kills due to low DO during warm weather. Therefore, preventing excessive aquatic growth helps to maintain DO levels. The use of artificial aeration (diffusers) can also be used to maintain adequate DO, especially in small impoundments or ponds.

AQUATIC PLANTS

Aquatic plants include algae and vascular plants. Phytoplankton, or algae, give water its green appearance and provide the base for the food chain in ponds. Tiny animals called zooplankton use phytoplankton as a food source. Large aquatic plants (aquatic macrophytes) can grow rooted to the bottom and supported by the water (submersed plants), rooted to the bottom or shoreline and extended above the water surface (emerged plants), rooted to the bottom with their leaves floating on the water surface (floating-leaved plants), or free-floating on the water surface (floating plants).

Aquatic plants are part of aquatic ecosystems. They provide a number of benefits, such as:

- Habitat for aquatic organisms (e.g., food and nesting sites).
- Oxygenation.
- Shoreline stabilization.
- Aesthetic appeal.

Aquatic plants growing on a littoral shelf may help protect receiving waters from the pollutants present in runoff. Ideally, littoral zones should have a slope of about 1 foot vertical to 6-10 foot horizontal to provide the best substrate for aquatic plant growth. In open areas, floating-leaved and floating plants suppress phytoplankton because they absorb nutrients from the pond water and create shade.

Particularly in shallow or nutrient-enriched ponds, aquatic plant growth can become excessive. Non-native plants can aggressively colonize aquatic environments. The excessive growth of any aquatic plant requires management. Following integrated pest management, a number of controls should be

considered to deal with excessive aquatic plant growth, including:

- Prevention, such as reducing nutrient enrichment and avoiding the introduction of invasive species. (See Chapter 5 for specific information on limiting phosphorus enrichment, the limiting nutrient for phytoplankton production, in freshwater resources.)
- Mechanical removal.
- Chemical control, if approved and permitted by the state. (Chemical controls are rare in Maine and require a discharge permit.)

SHORELINE MANAGEMENT

Special management zones should be established around the edges of lakes and ponds. The management specifications should include a setback distance when applying fertilizers, as well as reduced mowing. Grass clippings should be collected near shorelines, as the phosphorus and nitrogen in clippings can impact water quality.

WATERFOWL

The deposits of fecal matter by resident and migrating waterfowl (such as Canada geese) can substantially impact water quality through nutrient enrichment. On golf courses, shallow ponds with significant populations of waterfowl are most likely to be affected. In addition, large numbers of Canada geese can erode shorelines and thin the grass cover on greens and fairways, contributing to the potential for erosion. Efforts to control waterfowl have met with mixed success. Loud sounds, dogs, and hunting have been tried in order to deter them. However, many of these efforts do not lend themselves to golf courses, especially in more urban areas.

Riverside Golf Course, Portland, ME. © FBE



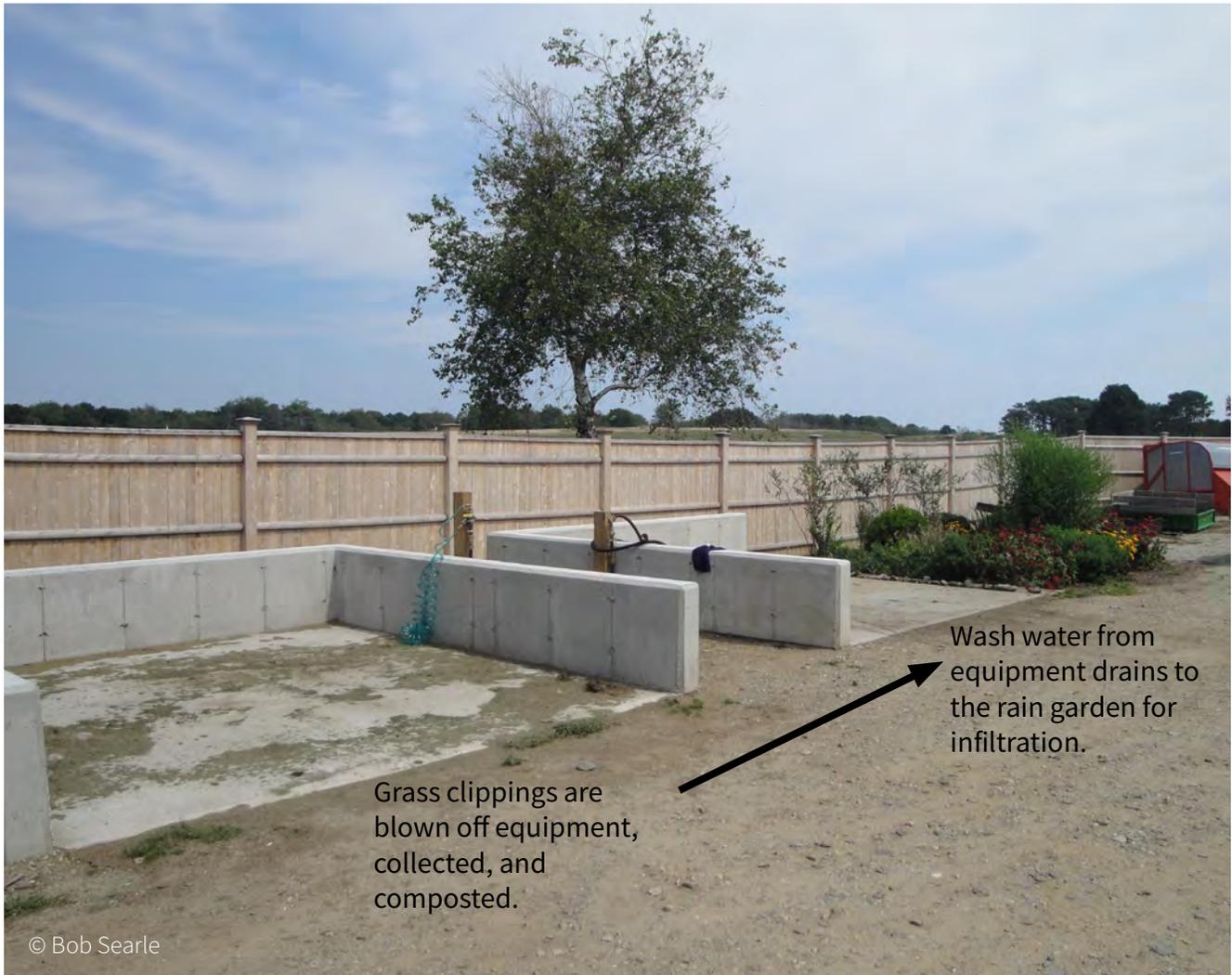
Best Management Practices for Constructed Lake and Pond Management

- Maintain water flow through lakes if they are interconnected.
- Establish wetlands where water enters lakes to slow water flow and trap sediments.
- Maintain appropriate erosion and sedimentation controls on projects upstream to prevent sedimentation and nutrient enrichment to waterbodies.
- With appropriate permitting, dredge or remove sediment before it becomes a problem. When dredging, take extreme measures to avoid discharge of sediment laden water downstream. If feasible, drain the pond and excavate the sediments in a dry environment. Permitting is required.
- Establish DO thresholds to prevent fish kills, which occur at levels of 2-3 mg/L.
- Reduce stress on fish by keeping DO levels above 5 mg/L.
- Manipulate water levels to prevent low levels that result in warmer temperatures and lowered DO levels.
- Use artificial aeration (diffusers), if needed, to maintain adequate DO.
- Develop a comprehensive management plan that includes strategies to prevent and control the growth of nuisance aquatic vegetation.
- Keep phosphorus rich material (e.g., natural or synthetic fertilizers, organic tissues like grass clippings, or unprotected topsoil) from entering surfacewater.
- Install desirable native plants to naturally buffer DO loss and fluctuation.
- To control excessive aquatic plant growth, use an IPM approach that incorporates prevention, cultural practices, and mechanical removal methods in addition to chemical control. A discharge Variance Permit is required³.
- To reduce the risk of DO depletion, use an algaecide containing hydrogen peroxide instead of one with copper or endothall. A Variance Permit is required.
- Reverse-grade around the waterbody perimeters to control surface water runoff and to reduce nutrient loads.
- Discourage large numbers of waterfowl from colonizing golf course waterbodies.
- Use a multi-faceted, IPM approach to control nuisance animals, such as Canada geese. Woody and shrubby buffers are highly effective at eliminating Canada geese congregation in adjacent turf areas.

3 <https://www.maine.gov/dacf/php/pesticides/documents2/forms/VariancePermitCh29-2013.pdf>

Protecting **Water Quality** on the Course

A case study from Abenakee Club



Grass clippings are blown off equipment, collected, and composted.

Wash water from equipment drains to the rain garden for infiltration.

© Bob Searle

Abenakee Club installed wash pads in 2012 to wash vehicles and equipment. Wash pads allow equipment to be cleaned while protecting the environment from contaminants that would otherwise drain into local water sources or cause stormwater contamination.

The photo above shows the two wash pads installed at Abenakee. Residuals (such as grass clippings) collected on the wash pad to the left, are contained and then moved to a compost pile. Residual wash water created on the pad to the right is drained to a rain garden for infiltration.

Photo: wash pad and rain garden installed at the Abenakee Club.

Abenakee Club installed a rain garden to infiltrate water draining from the washpad. Rain gardens are bowl shaped planted areas that collect stormwater from hard surfaces. Captured water slowly seeps into the ground, reducing the amount of pollutants that reach storm drains and waterways.

Abenakee Club designed their rain garden with the assistance of a landscape architect. They planted species including Beebalm, Aster, Iris, Boltonia, Baptisia, and blueberry plants.

Photo: Beebalm (top left), rain garden on the course (top middle), (top right), example rain garden design cross section (bottom).



© almanac.com

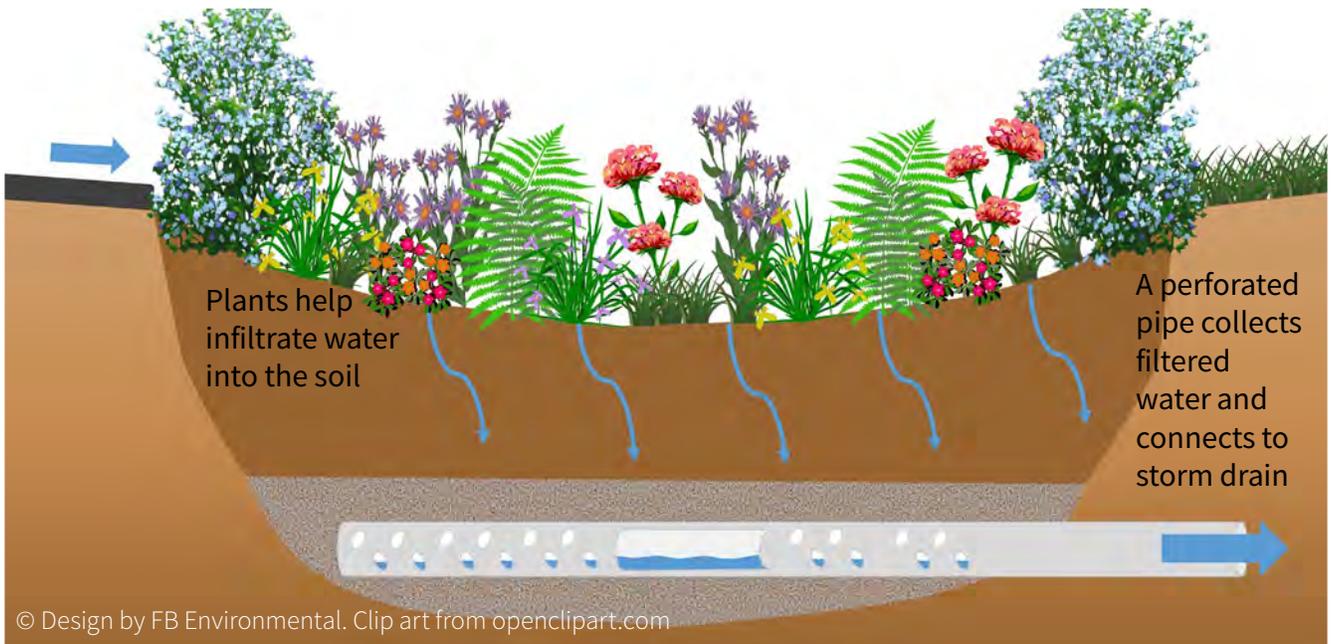


© Bob Searle



© almanac.com

Water from hard surfaces drains to the rain garden





CHAPTER 4: WATER QUALITY MONITORING

Belgrade Lakes Golf Club, Belgrade, ME. © Tim O'Wril

PREFACE

Monitoring can be used to set a pre-construction baseline for water quality. Routine monitoring can be used to measure water quality changes and to identify any areas where corrective actions should be taken. Monitoring can also demonstrate the presence of issues in water as it enters a golf course property not related to any impacts from facility management.

Golf course superintendents wanting to develop and implement a water quality monitoring program should identify a local water quality professional to assist them in the development of a robust and appropriate water quality monitoring program. The section below outlines critical things to consider in developing this program, however, it is important to note that monitoring programs are specific to the conditions and characteristics of each individual waterbody.

Review Existing Data

It is important to first review available baseline water quality data, which can include both groundwater and surface water sampling sites. In Maine, water quality data is stored in the Maine Environmental and Geographic Analysis Database (EGAD). This database includes physical, chemical, biological, and spatial data related to water quality in Maine and includes both groundwater and surface water sampling sites. Data housed in this database has undergone thorough quality assurance and quality control by the Maine DEP. Baseline data can be assessed to determine the likely origin of contaminants, measure the extent of sedimentation and nutrient inputs, and estimate the potential impacts to surface water and groundwater. In addition to monitoring surface and/or groundwater, water quality monitoring of irrigation sources (particularly water supply wells and storage lakes) provides valuable agronomic information that can inform nutrient and liming programs.

Groundwater Monitoring

Groundwater monitoring from wells located at the hydrologic entrance and exit from the course may be the best way to evaluate a golf course's impact on water quality. If groundwater monitoring data from these locations is not available from existing sources, monitoring wells can be installed by private companies. Installing groundwater monitoring wells can be relatively expensive, but the expense may be justified in certain cases where the origin of contamination can only be determined through comparison of water quality entering and exiting the property. To identify the appropriate site for monitoring wells, groundwater flow information is required. If this information is not available, experienced environmental engineering firms or the United States Geological Survey (USGS) can assist in determining suitable monitoring well locations.

Baseline water quality monitoring of adjacent streams, rivers, and wetlands can alert superintendents to changes in water quality that may indicate a contamination. © FBE



Surface Water Monitoring

For new golf courses or renovation projects in the planning stage, baseline water quality levels should be measured prior to construction at points of entry and exit of flowing water sources on or surrounding the golf course and on any surface water. This information can be used to form a baseline of flow and nutrient/chemical levels. For established courses, ongoing, routine water sampling provides meaningful trends over time. Post-construction surface-water quality sampling should begin with the installation and maintenance of golf course turf and landscaping and should continue through the first three years of operation and during the wet and dry seasons every third year thereafter, provided that all required water quality monitoring has been completed and the development continues to implement all current management plans. A single sample is rarely meaningful in isolation. It may also be wise to sample if a significant change has been made in course operation or design that could affect nearby water quality.

Things to Consider

VISUAL INSPECTION

The sections below discuss nuances to a complete water quality monitoring program. However, course superintendents and employees can provide useful indicators for assessing potential contamination to surface waters. This can include visual checks during storm events to identify where water is running and what it looks like – does the runoff contain a color and/or sediment? Is there an oil or petroleum sheen around parking areas? Is there soil build-up and instability around catch basins? A simple tool for visual water quality inspection is the Imhoff Cone, a clear cone-shaped container that can be used for identifying color and measuring sand and settled matter in runoff. If the course superintendent identifies an abnormality in this visual inspection, it can inform further investigative sampling.



A visual inspection can indicate a contamination problem that is increasing turbidity of the water. © FBE

SPATIAL SELECTION OF SAMPLING SITES

The number of monitoring samples is highly variable and depends on the size, location, and number of water sources on or near the golf course. The entry and exit points of golf course water sources are logical sampling points. However, sampling and analysis of standing water sources (i.e. ponds), springs, and any other irrigation sources should also be included. It may also be wise to sample if a significant change has been made in course operation or design that could affect nearby water quality. When determining sampling locations, it is important to characterize the local hydrology to understand the impacts of upstream features on each sampling location, such as tributaries and wetlands.

FREQUENCY, DURATION, AND TIMING OF SAMPLING

Water sampling can be an effective way to evaluate changes in water quality in course ponds, or in downstream or upstream waters if evaluated under baseflow (dry weather) conditions. However, pollutant transport often occurs in storm event (wet weather) conditions when surface water runs over the land and enters surface waters. Over the duration of a storm event, the concentration of pollutants (e.g. mg/L) and the mass of pollutants (e.g. kg/time) will vary dramatically, often rising rapidly during or immediately after periods of intense rain and declining more gradually as the intensity drops. In addition to the variability presented by the occurrence of one storm event, pollutant concentrations can vary across storm events of different intensities or occurring in different seasons (i.e. antecedent soil moisture conditions).

As such, if wet weather sampling is included as part of the monitoring program, it is critical to work with a professional to develop an appropriate sampling regime. This should include flow-weighted samples (to extrapolate the results of one grab sample to the flow conditions at the time of sampling). This should also include a careful analysis of the storm intensity, duration, and antecedent conditions.

BIOLOGICAL MONITORING

To overcome the aforementioned variability in analyzing individual pollutants in surface waters, scientists often use biological indicators to identify any critical changes to the water quality. Stream biomonitoring is a method to evaluate the condition of a stream or river using biological surveys of the living organisms that inhabit the waters. It is a way of inferring the water quality based on what organisms are present and can be an excellent way of viewing changes in biota of a stream that represent the net effect of all potential impacts on water quality and habitat suitability. Sampling of stream benthic macroinvertebrates (benthic macroinvertebrates are relatively large organisms that inhabit bottom substrates of streams and lakes for at least part of their life cycles) is a useful addition to a monitoring program, as the composition and diversity of these species can be used as a relative assessment tool for stream health. In summary, scientists can collect benthic macroinvertebrates through a variety of methods (e.g. rock baskets, kick-netting) and can send them to a qualified taxonomist to identify the species present. If water quality conditions are poor, sensitive species will likely not be present (or present in limited numbers). Benthic macroinvertebrate data is particularly useful if coupled with continuous monitoring of temperature, dissolved oxygen, and conductivity. These field parameters can help to identify the specific stressor on the macroinvertebrate community. Benthic macroinvertebrate sampling can be an effective and efficient alternative to the expensive and difficult challenge of wet weather sampling. Often, such sampling can often be undertaken by university students in fulfillment of course work.

Water Quality Analysis

Testing protocols can be simplified to test only those parameters that are directly influenced by course management, including organic and inorganic levels of nitrogen and phosphorus and a pesticide screen for selected pesticides used on the course. Additional analytes can include watershed basin-specific parameters of concern, such as sediments, suspended solids, and heavy metals. During measurements of dissolved oxygen, pH and alkalinity can also be sampled.

Samples should be analyzed by a certified laboratory, and all quality assurance/quality control (QA/QC)

procedures must be followed. The purpose of QA/QC is to ensure that chemical, physical, biological, microbiological, and toxicological data are appropriate and reliable. If a golf course should ever need to produce data for an agency or go to court to defend the facility, the data must meet QA/QC standards to be defensible as evidence. Additionally, it is important that samplers consider the detection limit for each parameter sampled to ensure that the laboratory equipment can detect levels low enough to monitor results against recommended thresholds of contamination. This is especially important for phosphorus, the limiting nutrient in freshwater systems, which can cause excess plant and algae growth at concentrations lower than many laboratory detection limits¹.

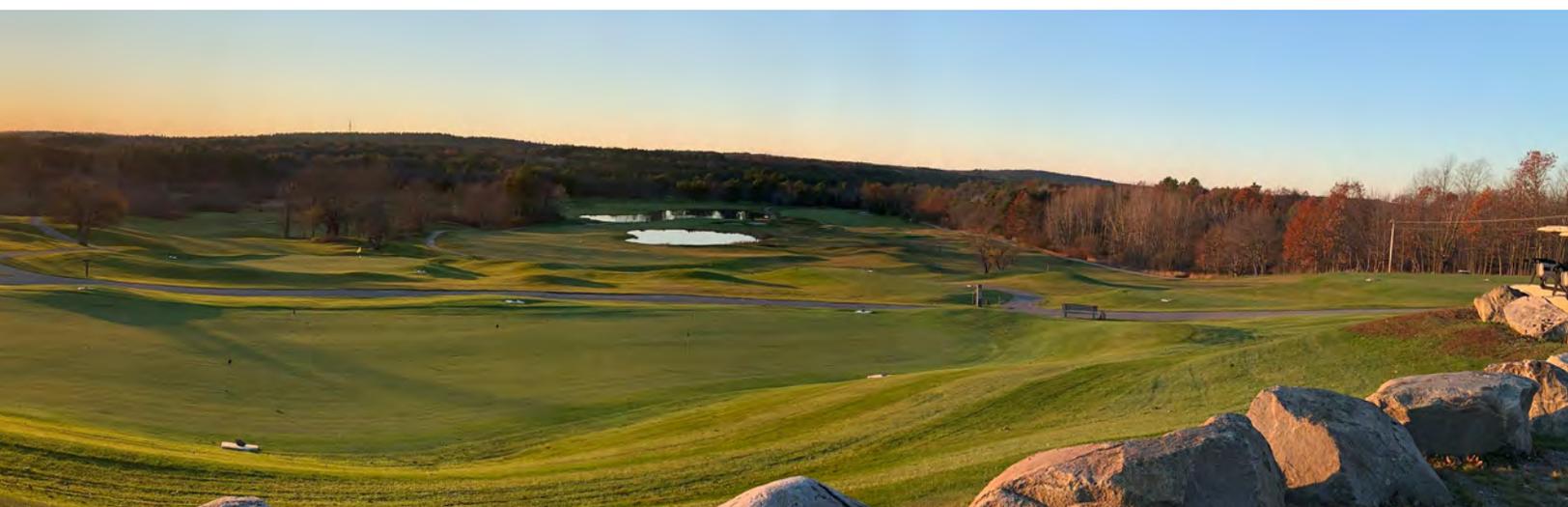
INTERPRETING WATER QUALITY RESULTS

Water quality can be analyzed by private companies or by university laboratories. Interpretation and use of water quality monitoring data depends to a large extent on the goal of the monitoring program. For example, the results may be analyzed to compare:

- Values over time.
- Values following implementation of BMPs, such as IPM measures.
- Monitoring points entering the site and leaving the site.

Results should also be interpreted and compared with the state's water quality standards, if standards have been established for the parameter being evaluated. Data analysis can also be used to identify issues that may need corrective action, based on findings such as a spike in nutrient levels. For example, operator error in nutrient applications, an extreme weather event, or some combination of factors may be responsible. Water quality problems can often be addressed by simple changes to a course's existing nutrient management program. A well-designed monitoring program is critical to interpreting water quality monitoring results. If a sampling design does not consider the factors discussed above (spatial, temporal, and event-based variability), it could lead to a misinterpretation of the cause of poor results. This could produce both false positives (identifying an issue when one does not exist) or false negatives (not identifying an issue that does exist).

Spring Meadows Golf Club, Gray, ME. © Tim O'Wril



¹ The Maine Health and Environmental testing Laboratory (HETL), located in Augusta, offers low total phosphorus detection limits of 2 ppb using method L-10-115-01-1-F.

Best Management Practices for Water Quality Monitoring

- Review existing sources of groundwater and surface water quality information. The Maine Environmental and Geographic Analysis Database (EGAD) stores physical, chemical, biological, and spatial data related to water quality in Maine. This includes groundwater and surface water sampling sites. A data request must be made to the database managers to receive data on specific waterbodies or geographic areas. This information is available [here](#), or at the link in the footnote².
- Work with an environmental professional and the Maine DEP to develop a water quality monitoring program. Ensure that this monitoring program considers applicable quality control procedures and accounts for seasonal, temporal, and event-based variability.
- If possible, establish a baseline quality monitoring program prior to construction of a new course.
- Identify appropriate sampling locations and sample at the same locations in the future.
- Visually monitor/assess any specific changes of surface waterbodies.
- Follow recommended sample collection and analytical procedures.
- Partner with other groups or volunteer water quality monitoring programs if possible, to share data and monitoring costs.
- Compare water quality monitoring results to benchmark quality standards.

Field water quality testing supplies, Val Halla Golf Club, Cumberland, ME. © Toby Young



2 <https://www.maine.gov/dep/maps-data/egad/>



CHAPTER 5: GOLF TURF FERTILIZATION AND NUTRIENT MANAGEMENT

Belgrade Lakes Golf Club, Belgrade Lakes, ME. © Tim O'Wril

PREFACE

Fertilization is the key cultural practice that supplies essential nutrients to turfgrass plants. A sound fertilization program ensures turfgrass persistence, performance, and quality. Under many instances, turfgrasses require supplemental fertilizers to provide nutrients not supplied in adequate amounts by the soil.

Proper fertilization, combined with appropriate mowing, irrigation, and pest control, produces healthy and attractive turf that withstands the wear and tear of intended use. Proper fertilization of turfgrass is extremely important because of the high demands for turfgrass performance and quality, the wide use and establishment of turfgrasses under less than favorable conditions, and the great demands placed on turfgrass for high recuperative capacity and wear tolerance. This requires a high level of cultural expertise including knowledge of plant nutrition, soils, and fertilizer technology and application.

Variables to consider when developing a fertilizer program include turfgrass species, mowing height and frequency, soil type and structure, irrigation, intended use of the turfgrass, temperature, environmental factors such as shade or sun, and whether clippings are returned.

Essential Elements for Turfgrass and Plant-Available Ionic Forms

At least 17 elements are considered essential for turfgrass growth. Macronutrients, the nutrients required in relatively large amounts, include carbon (C), hydrogen (H), oxygen (O), nitrogen (N), phosphorus (P), potassium (K), calcium (Ca), magnesium (Mg), and sulfur (S). Micronutrients, nutrients required in relatively smaller amounts, include iron (Fe), manganese (Mn), zinc (Zn), boron (B), molybdenum (Mo), copper (Cu), chlorine (Cl), and nickel (Ni)¹. Turfgrasses obtain C, H, and O from the atmosphere and water; the remainder are obtained primarily by roots from the soil. (Under certain special conditions, foliar uptake of nutrients happens through directed foliar nutrient sprays.) For turfgrasses to take up an essential element from the soil solution, it must be present in a plant-available ionic form, and water must be moving into the plant from the soil.

The plant-available ionic forms vary with each element (Table 5-1).

Table 5-1. Essential elements and their plant-available forms.

Macronutrients		Micronutrients	
Element	Plant-available ion	Element	Plant-available ion
Nitrogen (N)	NH_4^+ , NO_3^-	Iron (Fe)	Fe^{2+} , Fe^{3+}
Phosphorus (P)	H_2PO_4^- , HPO_4^{2-}	Manganese (Mn)	Mn^{2+}
Potassium (K)	K^+	Boron (B)	H_2BO_3^-
Sulfur (S)	SO_4^{2-}	Copper (Cu)	Cu^{2+}
Calcium (Ca)	Ca^{2+}	Zinc (Zn)	Zn^{2+}
Magnesium (Mg)	Mg^{2+}	Molybdenum (Mo)	MoO_4^{2-}
		Chlorine (Cl)	Cl^-
		Nickel (Ni)	Ni^{2+}

Though present in large quantities in most soils, the availability of macronutrients in the soil may be low if they are not in the plant-available forms. When availability is below optimum, N, P, and K are most often supplied by applying fertilizers. Calcium and Mg are routinely supplied from liming materials. Historically, sulfur fertilization of turfgrasses was generally not needed in New England because S requirements were met through atmospheric deposition of S-containing air pollutants and S mineralized from organic matter. However, improved air quality and reduced S-containing air pollutants is resulting in more S deficiencies than previously documented. Sulfur is also a component of some N- and P-containing fertilizers and certain pesticides.

Micronutrients are rarely deficient in New England soils, and therefore applying them on a routine basis is generally not necessary. However, on sand-based rootzones, micronutrient applications are warranted based on soil test results and can improve turfgrass performance. A commonly applied micronutrient on both soil and sand rootzones is Fe. This is not necessarily based on a deficiency of Fe, but because application of Fe, particularly as a foliar spray, has been shown to increase the dark green color in cool-season turfgrasses resulting in higher turf color quality.

¹ Sufficiency levels for Mo, Cl, and Ni are so low they are seldom actively monitored or managed.

Soil pH affects nutrient availability to plants (Figure 5-1) and has the single greatest effect on micronutrient availability, especially for Fe. Maintaining soil pH to slightly less than neutral (e.g., 6.8), results in optimal availability of all the nutrients. Acidic (<6.0) or alkaline (>7.0) soils may result in deficiencies of both macro- and micronutrients due to pH-dependent changes in ionic forms or complexing with other elements that precipitate in the soil solution, thus becoming unavailable.

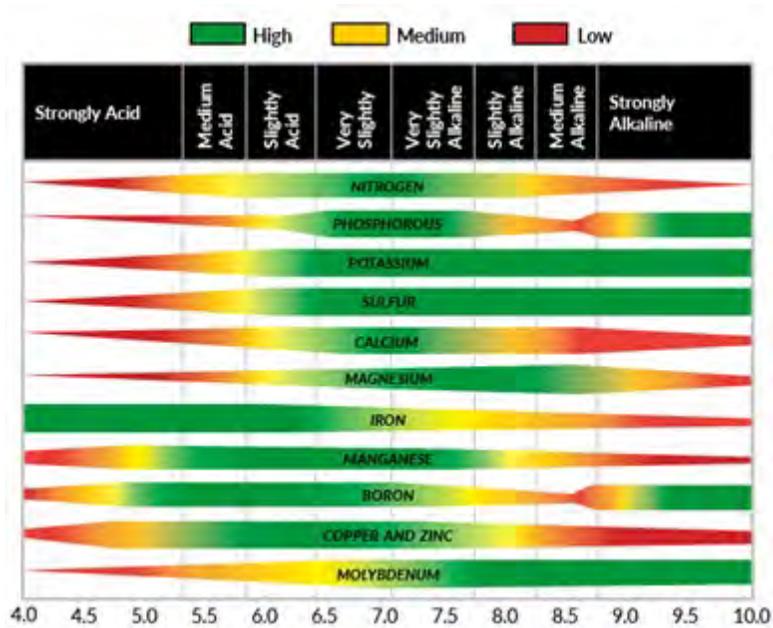


Figure 5-1. Relationship of soil pH and nutrient availability.

Basis for Fertilization

Fertilization of turfgrasses should be based on site-specific nutrient inputs and outputs and the needs of a particular turfgrass species. Nutrient inputs include fertilizers, organic matter decomposition, residue additions (i.e., return of clippings), atmospheric deposition, and nutrients contained in the irrigation water (Petrovic, 1990). Outputs of nutrients from a turf system include the removal of clippings, gaseous losses (denitrification and volatilization), losses with percolating water (leaching) and runoff, and immobilization in the soil organic matter or mineral fractions (Petrovic, 1990). When turfgrass nutrient requirements exceed the supplying capacity of the system, fertilization is needed to make up for this deficiency.

The capacity of any particular turfgrass plant to obtain available nutrients from the soil influences the fertilization requirements. Factors to consider include:

- Root extent and depth. Roots can only absorb nutrients where they are growing. Therefore, conditions that favor deep and extensive root development improve nutrient uptake capabilities. Compacted and/or wet soils have restricted rooting depths that may prevent adequate nutrient uptake. Fertilizer salt damage can also impact root viability and function.

A thatch layer is a dense layer of organic material located at the base of the turf grasses or vegetation. The thatch layer includes both dead and living organic materials and can restrict fertilizer movement to the soil.

- Thatch layer. If a large quantity of roots are located in the thatch layer, nutrients in the underlying soil may not be utilized and soil tests may incorrectly report the nutrient availability in the soil profile where roots should be concentrated. Furthermore, a thick thatch layer may restrict fertilizer movement so that nutrients may not reach the roots in the soil in sufficient amounts to support demand.
- Organic matter decomposition. The amount and rates of mineralization of organic matter affect the nutrient availability and existing fertility of a soil. Soils with high organic matter content may not need as frequent fertilizer applications as soils with low organic matter content if they are decomposing at a reasonable rate and releasing available forms of essential elements.
- Soil pH. The solubility of all essential elements and soil microbial activity is affected by pH. Maintaining a soil pH between 6 and 7 for turfgrasses is typically recommended, although turfgrasses can grow reasonably well over a wide range of pH values.
- Cation Exchange Capacity (CEC). Finer textured soils and soils with higher organic matter content have a higher CEC and a higher nutrient-supplying capacity than coarse textured soils and sand rootzones. Therefore, lighter and more frequent fertilizer applications may be required on coarse- textured soils and constructed sand rootzones than on fine-textured soils to supply necessary nutrition and minimize nutrient leaching.
- Losses. Gaseous losses of nitrogen via denitrification and volatilization and nutrient losses via leaching and runoff decrease the amount of nutrients available for turf growth. Applying fertilizers at appropriate times and rates and avoiding overwatering of turf limit these losses.

DETERMINING THE NEED FOR SPECIFIC NUTRIENTS

Soil test results should be the foundation of a turfgrass fertilization program. Soil tests are used routinely to determine the availability of essential nutrients. Past fertilization practices and responses are useful guides for N fertilization. Tissue tests are becoming more widely utilized but are expensive compared with soil tests. Reflectance meters and digital image analysis may prove to be useful in a turfgrass N management program in the near future (Karcher and Richardson, 2013; Bell et al., 2013; Guillard et al., 2016; Inguagiato and Guillard, 2016).

Other conditions and factors affect the needs for specific nutrients. The inherent nutrient supplying capacity of the soil and organic matter mineralization potentials may decrease or increase the need for specific nutrients based on turfgrass performance and quality goals. Incidence and severity of diseases, insects, and other pest problems may demand changes in current fertilization practices, as may abiotic environmental stresses such as drought, heat, or cold conditions. Increased traffic directly and indirectly affects nutrient needs and uptake. Clipping management (returned vs. removed) greatly affects the availability and nutrient status of a turfgrass soil. When clippings are returned to the turf, fertilization rates may be decreased by as much as 50% without loss in quality (Kopp and Guillard, 2002). Contrary to popular belief, grass clippings decompose rapidly under normal mowing practices and do not contribute to thatch (Kopp and Guillard, 2004). Therefore, whenever possible, clippings should be

THE FOUR “R’s” OF NUTRIENT MANAGEMENT

- 1. Right source**
- 2. Right rate**
- 3. Right time**
- 4. Right place**

returned as part of a sustainable nutrient management program.

Common sense should provide a guiding influence on nutrient management programs for turf. When the potential for water quality impacts exist, turfgrass fertilizer applications need to be closely managed. For example, fertilization (especially excess nutrient application) near open waterbodies or on sites with high leaching potential may contribute to contamination of receiving waters. In these cases, the solubility and release rates of N formulations, timing of application, and application rate need to be carefully considered. Fertilizers should only be applied during active growth periods - application of fertilizer during periods of drought induced dormancy leads to drought stress, root damage, and excess nutrient runoff when rains resume. Fertilizer applications should not be made to frozen turf.

IMPORTANCE OF SPECIFIC NUTRIENTS TO TURFGRASS GROWTH AND DEVELOPMENT

Although all essential elements are required for adequate turfgrass growth and development, some elements are more frequently associated with specific growth responses in turf (Table 5-2).

Table 5-2. Selected nutrients and general effects (+ positive and - negative) on turfgrass growth and quality.

Turfgrass Growth Response	N	P	K
Rooting	+/-	+	
Shoot Growth (leaves, tillers, rhizomes, stolons)	+		
Color	+		
Establishment (germination and seedlings)		+	
Environmental Stress Resistance and Tolerance			
Drought			+
Heat			+
Cold			+
Disease Susceptibility	+/-		+
Wear Tolerance	+		+
Recuperative Potential	+		
Composition of turf community	+/-	+/-	

Soil Testing

Soil testing plays an important role in a turfgrass nutrient management program. Soil tests measure the soil pH (how acidic or alkaline the soil), amounts of available macro- and micronutrients, and other chemical or physical properties such as soil texture, bulk density, porosity, CEC, soluble salts, and organic matter. A turf manager needs to know about the various soil properties to apply the proper amount of fertilizer and lime. Too little fertilizer and lime may result in reduced turf quality, vigor, stand persistence, performance, and tolerance to biotic and abiotic stresses. Too much fertilizer may increase problems with diseases and insects, reduce environmental stress tolerances, deplete plant storage carbohydrates (i.e., food energy), increase the potential for nutrient losses off-site, and increase economic losses due to unneeded nutrients.

Soil tests for the plant available N forms of NH_4^+ and NO_3^- are available, but their utility for turf is limited because the rapid transformations that can occur for these various forms of N in the turf-soil system make the tests largely meaningless, unless they are conducted at two-week or shorter intervals during the growing season (Geng et al., 2014). This is logistically challenging on most golf courses. Consequently, N fertilization needs for turf have been based historically on various measures of turfgrass species growth and quality such as clipping yields, color, visual symptoms of N deficiency, shoot density, tillering, density, rooting characteristics, and recuperative capacity. New soil tests for labile (i.e., “active”) N and C that predict mineralization have shown promise for guiding N fertilization of cool-season turfgrasses (Moore et al., 2019a and 2019b).

SOIL SAMPLING PROCEDURE

The first step in a nutrient management program is to collect soil samples. Soil samples can be collected any time the ground is not frozen or excessively wet (which makes it too hard to get a representative sample), and not shortly after a lime or fertilizer application. Because temperature affects the mineralization and weathering of organic matter and parental minerals, it is best to take soil samples the same time each year so that seasonal differences do not confound the test results. When renovating or establishing turfgrass, soil samples should be taken two or more weeks prior to beginning work in the area.

Soil test recommendations are only as good as the procedures used to collect samples. Generally, many samples are taken from one uniform area, mixed thoroughly in a clean pail, and a subsample taken from the many mixed samples to represent the area. A “Z” or “W” sampling pattern can be used to collect the samples in an area, with the samples taken along the path of the pattern. As a general guideline, 10 to 15 samples should be taken across the specific sections (separate greens, tees, fairways, and roughs into separate sections). Areas that are fertilized, mowed, irrigated, managed differently, or are unusual should be sampled separately to avoid biasing the sample.

Soil samples should be collected to a depth of no more than 4 inches for established turfgrass and 6 inches where a new seedbed has been prepared. When limited thatch is present in the profile, it should not be included in the sample. On sites where the majority (e.g., $\geq 75\%$) of roots are restricted to the thatch/mat layer, it may be preferable to include this portion in the sample.

Remember that the decision to retain or discard thatch influences soil test results from year to year. Once collected, the subsample should be placed into a plastic bag and labeled with the corresponding

collection area. Avoid heating the sample once placed in the plastic bag (e.g., a car trunk on a hot day can become very warm) if plant available nitrogen is to be included in the soil test because some chemical changes may occur and erroneous conclusions may be reached regarding the need for fertilizers. Nutrients other than plant available nitrogen are minimally affected by incubation of moist soil.

SOIL TESTING FREQUENCY

Newly established turf areas should be tested annually for a few years until the nutrient status of the soil becomes stable. For established turf without problems, a soil sample every two to three years should be adequate. Problem turf areas should be sampled annually until problems are corrected. High-value turf areas probably should be sampled annually because the margin for error in these systems is so low.

Soil samples can be analyzed by commercial laboratories, land-grant university systems, or agricultural experiment stations such as:

- Connecticut – [University of Connecticut Soil Nutrient Analysis Laboratory](#) and the [Connecticut Agricultural Experiment Station](#).
- Maine – [University of Maine Agricultural and Forest Experiment Station's Analytical Laboratory and Maine Soil Testing Service](#).
- Massachusetts – [University of Massachusetts Soil & Plant Nutrient Testing Laboratory](#).
- New Hampshire – [University of New Hampshire Cooperative Extension Soil Testing Services](#).
- Rhode Island (soil pH only) – [University of Rhode Island Master Gardener's Program](#).
- Vermont – [University of Vermont Agricultural and Environmental Testing Lab](#).

EXPRESSION OF SOIL TEST RESULTS

The ways in which soil test results are expressed varies among soil testing laboratories. Some lab results are expressed on a qualitative basis: low, medium (or adequate), high; or below optimum, optimum, and above optimum. Other labs express results on a quantitative basis: pounds of available nutrient per acre or in units of parts per million (ppm). Soil tests in the United States are traditionally based on the "acre furrow slice," which is the volume of soil in the upper 6 to 7 inches of the soil profile. Historically, this relates to the standard depth of a plow, and on average this amount of soil is considered to weigh 2 million lbs. per acre. Using this as a base, it can be reasoned that if a nutrient was reported to be available at 2 lbs. per acre, then 2 lbs. of nutrient are available per 2 million lbs. of soil, or in its simplest form: 1 ppm. Therefore, pounds per acre are two times the ppm. It may be important when interpreting soil tests to understand whether the values are reported in pounds per acre or ppm. Soil test results also differ on how results are interpreted and how recommendations are formed.

SUFFICIENCY LEVEL OF AVAILABLE NUTRIENTS (SLAN)

The Sufficiency Level of Available Nutrients (SLAN) (Eckert, 1987) soil test interpretation and recommendation method is used by most university and governmental soil testing laboratories in the United States. This method relies on data collected under field conditions where response curves are generated for as many turf species and soil types as possible. The underlying principle of the method is that a beneficial turf response to added fertilizer is observed when the soil extractable-nutrient levels are below optimum, and the response to fertilization becomes increasingly small as the soil extractable-nutrient levels approach an optimum (Figure 5-2). There is a low probability of response to fertilizer additions when extractable-nutrient levels are at optimum or beyond; often the response will plateau.

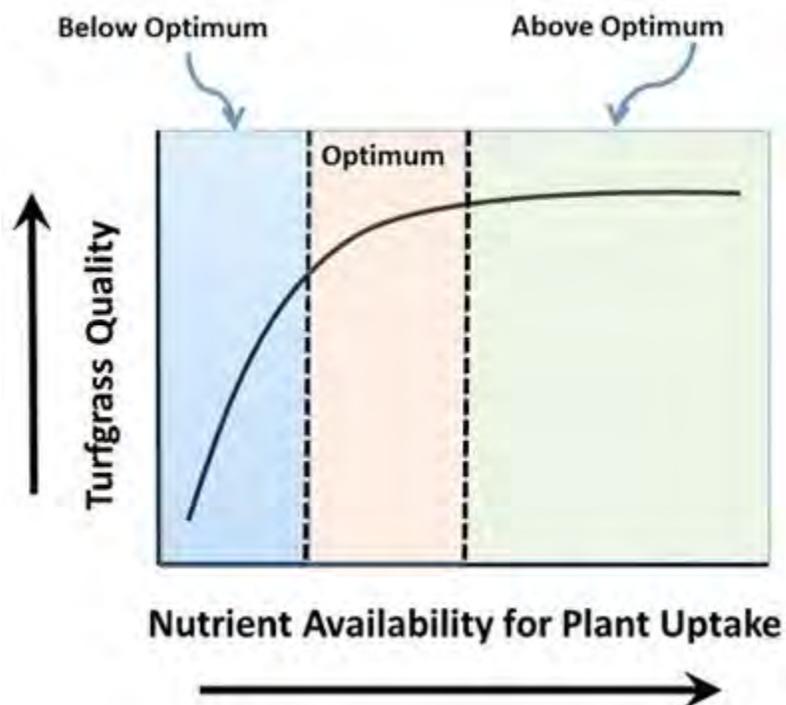


Figure 5-2. Theoretical turfgrass growth or quality response in relation to plant-available essential elements in the soil, according to the SLAN philosophy of nutrient management.

Categories and ranges for extractable nutrients used by the New England land-grant university soil testing laboratories for their SLAN turfgrass fertilizer recommendations are provided in Tables 5-3 to 5-7.

Table 5-3. University of Connecticut Soil Nutrient Analysis Laboratory categories and ranges for modified Morgan extractable nutrients, lbs./ac (or ppm) (Pettinelli, personal commun.).

	Below optimum	Optimum	Above optimum
Ca	0 – 1,799 (0 – 899.5)	1,800 – 2,399 (900 – 1,199.5)	>2,400 (>1,200)
Mg	0 – 174 (0 – 87)	175 – 249 (87.5 – 124.5)	>250 (>125)
P	0 – 5 (0 – 2.5)	6 – 20 (3 – 10)	>21 (>10.5)
K	0 – 99 (0 – 49.5)	100 – 349 (50 – 174)	>350 (>175)

Table 5-4. University of Maine Soil Testing Service optimum ranges or value for modified Morgan extractable nutrients, lbs./ac (or ppm) (Hoskins, 1997).

	All turf	New Seeding	Fairways	Greens/Tees
P	7 – 10 (3.5 – 5)	10 – 20 (5 – 10)	7 – 10 (3.5 – 5)	10 – 20 (5 – 10)
K	250 (125)	250 (125)	250 (125)	250 (125)

Table 5-5. University of Massachusetts Soil & Plant Nutrient Testing Laboratory categories and ranges for modified Morgan extractable nutrients, lbs./ac (or ppm) (Owen et al., 2016).

	Very Low	Low	Optimum	Above Optimum	Excessive
Ca	0 – 998 (0 – 499)	1,000 – 1998 (500 – 999)	2,000 – 3,000 (1,000 – 1,500)	>3,000 (>1,500)	-
Mg	0 – 48 (0 – 24)	50 – 98 (25 – 49)	100 – 240 (50 – 120)	>240 (>120)	-
P	0 – 3.8 (0 – 1.9)	4 – 7.8 (2 – 3.9)	8 – 28 (4 – 14)	28 – 80 (14 – 40)	>80 (>40)
K	0 – 98 (0 – 49)	100 – 198 (50 – 99)	200 – 320 (100 – 160)	>320 (>160)	-

Table 5-6. University of Vermont Agricultural and Environmental Testing Lab categories and ranges for modified Morgan extractable nutrients, lbs./ac (or ppm) (Bosworth, 2017).

	Low	Medium	Optimum	High or Excessive
P	<4 (<2)	4 – 7.8 (2 – 3.9)	8 – 19.8 (4 – 9.9)	≥20 (≥10)
K	<50 (<25)	50 – 198 (25 – 99)	200 – 318 (100 – 159)	≥320 (≥160)

Table 5-7 University of New Hampshire Cooperative Extension Soil Testing Services categories and ranges for Mehlich III extractable nutrients, lbs./ac (or ppm) (Saunders, 2018).

	Low	Optimum	High	Very High
Ca	0 – 1,600 (0 – 800)	1,600 – 2,400 (800 – 1,200)	2,400 – 4,000 (1,200 – 2,000)	>4,000 (>2,000)
Mg	0 – 120 (0 – 60)	120 – 240 (60 – 120)	240 – 320 (120 – 160)	>320 (>160)
P	0 – 60 (0 – 30)	60 – 100 (30 – 50)	100 – 150 (50 – 75)	≥150 (≥75)
K	0 – 340 (0 – 170)	340 – 560 (170 – 280)	560 – 860 (280– 430)	>860 (>430)

Martindale Country Club, Auburn, ME. © Scott Cybulski, CGCS



BASE CATION SATURATION RATIO (BCSR)

Soil test interpretation and recommendations may also be established on the base saturation percentage of the CEC. This method of testing is known as the Base Cation Saturation Ratio (BCSR) method (Bear et al, 1945) and is based on the concept that an ideal ratio of cations on the CEC sites produces the best plant response. Most private soil testing laboratories in United States use this method. According to this method, the proper ratio of nutrients in the soil exists when the percentage base saturation is approximately in the ranges shown in Table 5-8.

Table 5-8. Proposed optimum percentage base saturation for the BCSR method for soil test interpretation and recommendations (Albrecht, 1975) and University of Maine Agricultural and Forest Experiment Station’s Analytical Laboratory and Maine Soil Testing Service optimum ranges or value for modified Morgan extractable nutrients, % saturation (Hoskins, 1997).

Base cation	Proposed Optimum Base Saturation Ranges (%)				
	Albrecht, 1975	University of Maine			
	General crop	All turf	New seeding	Fairways	Greens/tees
Ca	60 – 75	60 – 80			
Mg	10 – 20	10 – 25			
K	2 – 5		2.1 – 3.0	2.8 – 4	2.8 – 4
H	10				
Other	5				

Interpretation of the BCSR often depends heavily these cation saturation ranges for each cation. Recommendations are modified to maintain minimum SLAN (Solvita Soil Labile Amino-Nitrogen) levels for very low CEC soils and maximum limits on application to very high CEC soils. Superintendents can also look at the ratio between cation saturations to interpret application rates (for example a Ca:Mg ratio of 6:5, Ca:K ratio of 13:1, and Mg:K ratio of 2:1). When the ratios vary from the proper ratio, fertilizer applications are recommended to restore the balance. Many turfgrass and soil scientists do not accept the BCSR method because no substantial scientific evidence exists to support the claim that maintaining an ideal ratio of soil cations maximizes crop response (Kopittke and Menzies, 2007; Chaganti and Culman, 2017). Under the BCSR method, it is possible that additional fertilizer is recommended even when the extractable-nutrient levels are high to very high. Regardless, turf practitioners indicate successful results following the BCSR method (see Simmons’ response in Schlosseberg and Simmons, 2012). In calcareous sand-based rootzones, the BCSR method could result in misleading ratios and cation saturation percentages (St. John and Christians, 2010) and should not be used as the sole factor in developing a turfgrass fertility program for sand-based rootzones that have either a low CEC or are calcareous (St. John and Christians, 2013).

MINIMUM LEVEL FOR SUSTAINABLE NUTRITION (MLSN)

The Minimum Level for Sustainable Nutrition (MLSN) is a newer soil test interpretation and fertilizer recommendation approach that identifies the minimum concentration of a soil element that supports “good” turf growth while maintaining desired turf quality and playability levels (Stowell and Woods, 2013). In the approach, nutrients are extracted from soils of “well performing turf” and concentrations are

fit to a log-logistic model that identifies the concentration where 10% of the soil samples fall below that point in the distribution. This concentration is then defined as the MLSN. (Woods et al., 2016). Figure 5-3 shows the relationship for P.

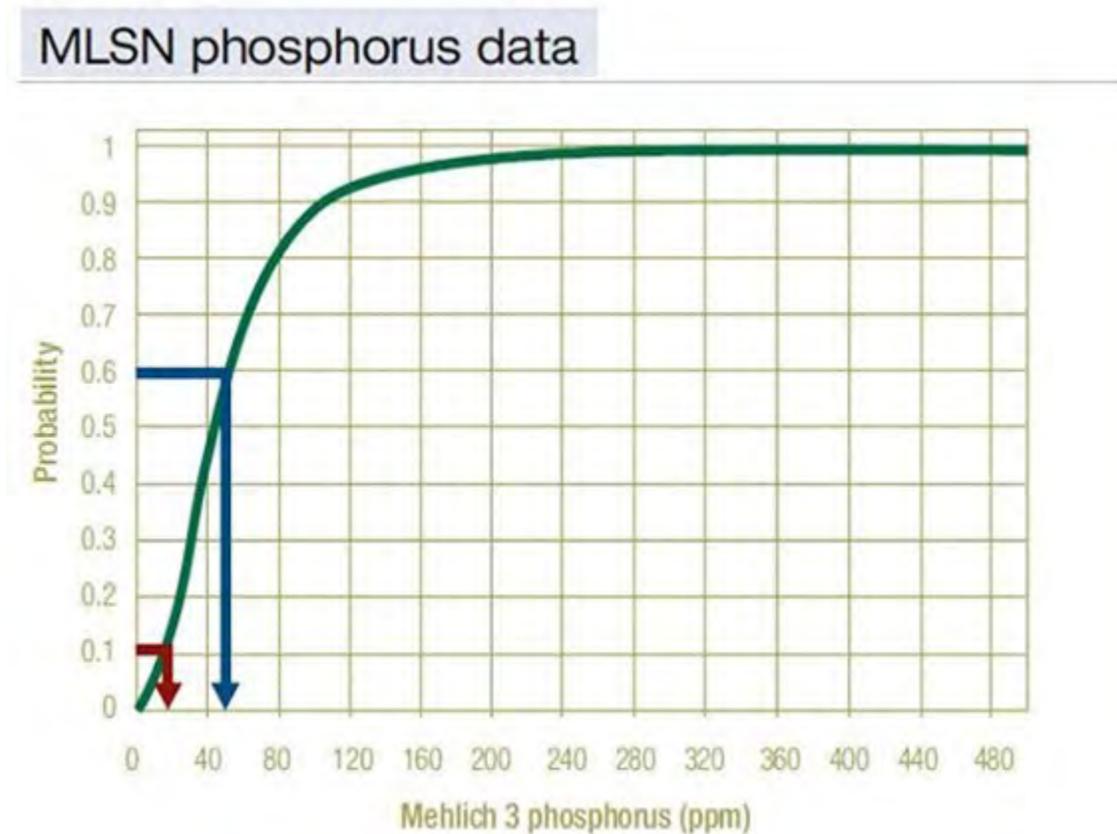


Figure 5-3. The cumulative distribution function for phosphorus to identify the MLSN guideline.

At the 0.1 probability level, 10% of the samples report phosphorus values lower than 18 ppm (red line). This is the phosphorus MLSN guideline. The blue line indicates the conventional guideline of 50 ppm for phosphorus. Fifty-nine percent (probability = 0.59 = sustainability index) of the samples report values lower than the conventional phosphorus guideline (Woods et al., 2014).

From the graph, one can plot out manually or use the predictive equation to estimate the MLSN at a Probability = 0.10 (i.e., concentration of the lowest 10% of well-performing samples). This is in contrast to the other methods that attempt to place soil test values into specific categories (e.g., optimum, low, high, etc.). The thinking in the MLSN method is that if the turf is performing well with soil nutrient concentrations below the optimum levels defined by the traditional SLAN and BCSR methods, then why add extra fertilizer? Development of the MLSN approach with more than 17,000 soil samples (and continuing) indicates that good quality turf can be maintained at concentrations well below that indicated as optimum with the SLAN or BCSR methods. Using the Mehlich III extractant, the MLSN guideline levels are 37, 21, 348, 47, and 7 mg kg⁻¹ for K, P, Ca, Mg, and S, respectively. These are much lower than optimum as indicated by the SLAN method (see Tables 5-4 to 5-7 above), and fertilizer additions would be recommended if following the SLAN interpretation. Since the MLSN levels are much lower than the conventional optimum levels, less fertilizer is recommended. This can result in substantial savings on most golf courses.

To date, the MLSN levels are based on the Mehlich III extractant. As indicated in the next section, most of New England soils are extracted with the Morgan or modified Morgan extractant. Therefore, unless soil test data comes from a lab utilizing the Mehlich III extractant, the MLSN values will not be directly translatable to soil test results based on other extractants.

EXTRACTANTS

Various extractants are used in soil testing laboratories. Often, the use of a particular extractant is regionally related or dependent on soil type or properties. In New England, soils are generally acidic, low in organic matter, high in Al and Fe, and have a relatively high sand content. For these soils, the Morgan (contains sodium acetate) or modified Morgan (contains ammonium acetate) extractants are regularly used for soil tests at the soil testing laboratories of the University of Connecticut, Connecticut Agricultural Experiment Station, University of Maine, University of Massachusetts, and University of Vermont. The Morgan extractants are weak acids and are appropriate for most types of soils in New England. Recently, the Mehlich III extractant (containing acetic and nitric acids along with ammonium fluoride, ammonium nitrate, and EDTA), which is the typical extractant for mid-Atlantic states, is being used to test soil samples submitted to the University of New Hampshire's soil testing services. The University of Rhode Island does not offer a soil testing service (other than pH through the Master Gardener's program). Some commercial soil testing services in New England will send the soil samples to analytical labs in the Midwest, where the Bray extractant (containing hydrochloric acid and ammonium fluoride) is used for soil testing.

It is important to understand the relationship between the various soil testing extractants and why they produce their results. Three common extractants used are the Bray method, the Mehlich method, the Morgan method, and the modified-Morgan method. Both the Bray and Mehlich extracts are acid-fluoride extracts and as such, extract almost identical P. The Morgan and modified-Morgan are designed to avoid aluminum-phosphates and as such, pull less P because they don't extract as much aluminum-bound P. The Morgan, modified Morgan, and Mehlich methods often extract similar amounts of Ca, Mg, and K, however the modified-Morgan uses ammonium acetate instead of sodium and therefore will extract more K from some (higher clay) soils than the original Morgan method. Therefore, P results should not be compared from different extractants. It is best to use the same extractant to monitor short- and long-term changes in soil test values over time.

© Bob Searle



Best Management Practices for Soil Testing

- Divide the course into logical sampling components such as greens, fairways, tees, roughs, etc., for each hole.
- Take 10 to 15 soil samples randomly from each respective section of the golf course and blend them together to provide a representative, uniform soil sample (separately for greens, tees, fairways, and roughs).
- Take each soil sample at the same depth.
- Use an extractant appropriate for the course's soils (historically, the Morgan and modified-Morgan extractants are best for New England soils). Most labs list the extractant used on their laboratory results, so confirm the method before interpretation.
- Ensure that the same extractant is used for each test in order to compare soil test results over time.
- Keep soil tests from prior years and review to observe changes over time. This practice can provide good evidence of the impact of the nutrient management plan.

Plant Tissue Analysis

Turfgrass fertilization recommendations can also be based on tissue testing. With tissue testing, the clippings are analyzed for nutrient concentration and this value is compared to a critical range indicating deficiency, sufficiency or excess (Table 5-9). Some believe that this method is more accurate than a soil test because it measures the concentrations of nutrients actually taken up by the grass rather than estimated from extractable soil values. This is especially true for nitrogen and micronutrients.

Table 5-9. Suggested sufficiency ranges for tissue nutrient concentrations of turfgrasses (Jones, 1980).

Macronutrients, %		Micronutrients, ppm	
N	2.8-3.5	Fe	35-100
P	0.3-0.6	Mn	25-150
K	1.0-2.5	B	10-60
Ca	0.5-1.3	Cu	5-20
Mg	0.2-0.6	Zn	20-55
S	0.2-0.5	Mo	not known
		Cl	not known
		Ni	not known

The major shortcomings of a tissue testing program include the expense as compared with soil testing; not all testing laboratories offer the test; results may not come back as quickly as soil test results; and the lack of actual calibration tests conducted with a wide range of turfgrasses under various conditions. Until more calibration tests are completed and verified, the above sufficiency ranges should be used as relative guidelines only.

Monitoring and recording turfgrass quality, clippings yield, and performance and then relating the observations to the tissue test concentrations will help guide future fertilization and nutrient management practices once a consistent relationship is established.

Best Management Practices for Plant Tissue Analysis

- Tissue samples may be collected during regular mowing.
- Do not collect tissue after any event that may alter the nutrient analysis such as fertilization, topdressing, and pesticide applications.
- Place tissue in paper bags, not plastic.
- If possible, allow tissue samples to air-dry before mailing them.
- Poor-quality turfgrass that is of concern should be sampled separately from higher-quality turfgrass.
- When turfgrass begins to show signs of nutrient stress, a sample should be collected immediately.
- More frequent tissue sampling allows a more accurate assessment of turfgrass nutrient status changes over time.
- The frequency of tissue analysis is dependent on individual course needs. Two to four tests per year are common on greens and one to two tests per year are common on tees and fairways.
- Keeping tissue test results from prior years allows for observation of changes over time.
- Tissue testing can provide good evidence of the impact of the nutrient management plan.

Fertilizers Used in Golf Course Management

Understanding the components of fertilizers, the fertilizer label, and the function of each element within the plant are all essential in the development of an efficient nutrient management program. By law, all fertilizers include a grade or analysis stating the percent by weight of nitrogen (N), phosphorous (as P₂O₅) and potassium (as K₂O) that is a guaranteed minimum in the fertilizer. A complete fertilizer contains N, P₂O₅, and K₂O. However, additional laws that govern fertilizer labeling vary greatly among states. Consult the land-grant university or the appropriate state agency (usually the Department of Agriculture) regarding state laws.

LABEL

The label is intended to inform the user about the contents of the fertilizer that, if understood and followed, will minimize environmental risk. The fertilizer label may contain the following information:

- Brand
- Size guide number (SGN)
- Manufacturer's name and address
- Guaranteed analysis/Grade
- "Derived from" statement
- Net weight

MACRONUTRIENTS

Macronutrients are required in the greatest quantities and include N, P, and K. Understanding the role of each macronutrient within the plant should provide you with a greater understanding of why these nutrients play such a key role in proper turfgrass management.

NITROGEN

Nitrogen is typically the nutrient required in the greatest quantity by turfgrasses aside from C, H, and O. Concentrations of N within tissues at sufficiency levels usually range from 2% to 6% N on a dry weight basis. When turf soils do not provide an adequate amount of N, persistence, performance, and quality of the turf suffers. This is usually expressed by reduced turf growth and development, reduced shoot and tiller density, reduced stolon or rhizome growth, increased weed infestations, and a yellowing of the leaf blades that reduces visual quality and resilience to abiotic and biotic stresses. It is critical that correct amounts be supplied at appropriate times in appropriate amounts. It is easy to see when turf is lacking N, but much harder to determine when N availability is beyond adequate and excessive.

It is commonly known that N fertilization results in a darker green leaf color, but consistent excessive N fertilization for a dark green turf color may not be beneficial in the long run. Under consistent and high N rates, turfgrass health may be compromised. Excessive N availability can result in the following:

- Poor root growth because shoot growth is stimulated at the expense of the roots.
- Poor rhizome and/or stolon development resulting in a weak sod.
- Higher incidence with hot- and cold-weather diseases.
- Reduced storage of food carbohydrates needed for regrowth following stress periods and overwintering; reduced recuperative ability.
- Poor tolerance to heat, cold, traffic, and drought stresses.
- Shifts of the turf community to species that are favored by high N (e.g., annual bluegrass).
- Higher succulence resulting in less wear tolerance and disease.
- More frequent mowing.
- Higher burn potential with certain types of N fertilizers.
- Environmental and economic losses of N.

Conversely, insufficient N fertility also can negatively affect performance of turfgrasses and the environment. Adequate N fertility enables turf to resist and/or recover from abiotic and biotic stresses. Maintaining sufficient N fertilization also increases turfdensity that can minimize weed infestation and the runoff of sediment, nutrients, and pesticides.

FATE AND TRANSFORMATION OF N

The goal of all applied nutrients is to maximize plant uptake while minimizing nutrient losses. Understanding each process increases the ability to make sound management decisions and ultimately leads to an increase in course profitability and a reduction in environmental risk. Nitrogen management is more complicated than other required essential elements because of the multiple transformations that can occur in the soil with N. Nitrogen processes are shown in Figure 5-4 and include:

- Mineralization, the microbial mediated conversion of organic N into plant- available NO_3^- and ammonium NH_4^+ .
- Nitrification, the microbial-mediated conversion of NH_4^+ to NO_3^- .
- Denitrification, the microbial mediated conversion of NO_3^- to N gas (NO , N_2O , or N_2); this primarily occurs in low-oxygen environments and is enhanced by high soil pH.
- Volatilization, the conversion of NH_4^+ to ammonia (NH_3) gas; this is enhanced by high soil pH and low oxygen environments.

- Leaching, the downward movement of an element through and below the rootzone.
- Runoff, the lateral movement of an element beyond the intended turfgrass location.

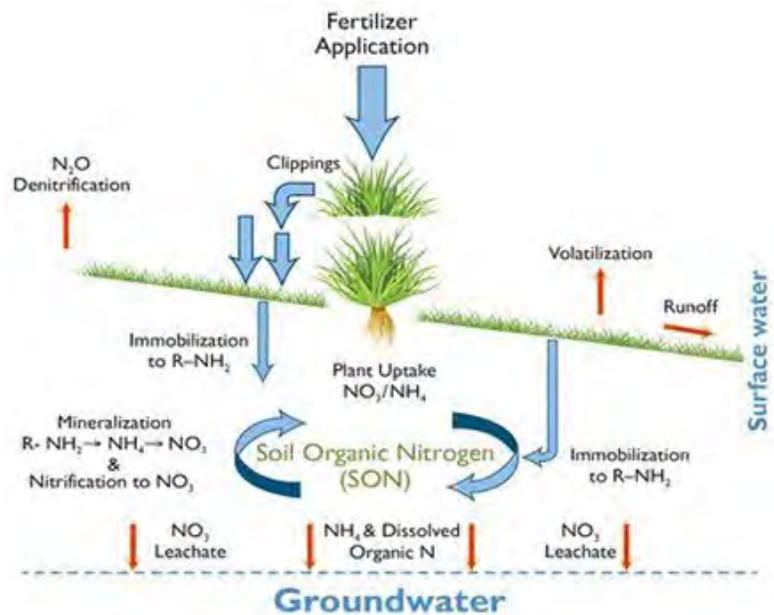


Figure 5-4. Fate and transport of nitrogen in fertilizers applied to turfgrass.

NITROGEN FORMULATIONS

Nitrogen can be found in various fertilizer formulations, either as a readily available form (highly soluble in the soil solution, or fast-release) or in a slowly available form (low solubility in the soil solution, or slow-release). The fast-release forms are composed of inorganic salts such as calcium nitrate (15.5% N) and potassium nitrate (13% N) or are synthesized by reacting ammonia with various compounds to form urea (45% to 46% N), ammonium nitrate (33% to 34% N), ammonium sulfate (21% N), or mono- and diammonium phosphates (11% to 20% N).

Understanding how certain N sources should be blended and applied is an essential component in an efficient nutrient management plan. Improper application of N sources increases the risk of negative environmental impacts and economic losses. Each N source (particularly slow-release forms) is unique and therefore should be managed accordingly. Applying a polymer-coated urea in the same manner one would apply a sulfur-coated urea greatly reduces the value of the polymer-coated urea. Similarly, applying 1 pound of N from ammonium sulfate may cause burning, while applying 1 pound of N from certain polymer-coated ureas may not provide the desired turfgrass response. Rate, application date, location, and turfgrass species all should be included in nutrient application decision-making.

SOLUBLE NITROGEN SOURCES

- Urea (46-0-0)
- Ammonium nitrate (34-0-0)

- Ammonium sulfate (21-0-0)
- Diammonium phosphate (18-46-0)
- Monoammonium phosphate (11-52-0)
- Calcium nitrate (15.5-0-0)
- Potassium nitrate (13-0-44)

Table 5-10. Advantages and disadvantages of fast-release N formulations.

Fast-release N formulations	
Advantages	Disadvantages
High percentage of N by weight.	Provide only a short-term response. Effectiveness lasts only four weeks or less, which necessitates more frequent applications.
Provide an immediate response.	High salt index and a high foliar burn potential. It needs to be watered in immediately after application.
Minimal temperature dependency that provides good response under cold temperatures in spring and fall.	Higher leaching potential because of solubility.
Relatively inexpensive per unit of N.	Higher volatilization potential especially with the ammonium-containing forms.
	Higher denitrification potential especially with the nitrate-containing forms.
	Ammonium and urea-form nitrogen fertilizers have an acidifying effect on the soil and nitrate forms tend to raise soil pH.

Because of the fast-release properties, turf response from soluble N sources is often characterized by short bursts of growth after application followed by periods of slow growth as the N is rapidly depleted or lost from the soil. The process is repeated over again with each subsequent N application. The peaks and valleys in growth not only result in a greater mowing frequency shortly after application but can also quickly deplete food carbohydrates in the grass. This has a highly negative effect on root growth and gradually may cause the turf to thin out.

Slow-Release Nitrogen Sources

A slow-release N source is any N-containing fertilizer in which the release of N into the soil is delayed either by requiring microbial degradation of the N source, by coating the N substrate to delay the dissolution of N, or by reducing the water solubility of the N source. These sources include:

- Sulfur-coated urea

- Polymer/resin-coated
- Isobutylidene diurea
- Urea-formaldehyde and methylene urea (urea and formaldehyde reaction products)
- Natural organics
- Urease and nitrification inhibitors
 - Urease inhibitors reduce the activity of the urease enzyme, which slows the conversion of urea to ammonium and slows down mineralization. This results in a reduction of volatilization and an increase in plant-available N.
 - Nitrification inhibitors reduce the activity of Nitrosomonas bacteria, which are responsible for the conversion of NH_4^+ to NO_3^- . This reduced activity results in a reduction of N lost via denitrification and an increase in plant- available N.
 - Although these products improve N availability and efficiency through reduced losses, they do not necessarily reduce the rate of N needed for desired goals.

The slow-release forms of turfgrass N fertilizers are derived either by reacting urea with various organic or inorganic compounds to form urea-formaldehyde and methylene ureas (up to 38% N), IBDU (31% N), and sulfur coated urea (22% to 38% N), or are derived from natural organic materials or residues such as manure, compost, bloodmeal, food industry by-products, and biosolids.

Table 5-11. Advantages and disadvantages of slow-release N formulations.

Slow-release N formulations	
Advantages	Disadvantages
Provide more uniform turfgrass growth during the growing season and do not produce peak and valley growth.	Higher cost per unit of N.
Lower salt index and a lower foliar burn potential in most situations.	May not supply sufficient N needed by the grass.
Have a long-term turfgrass response and can carry over from year to year.	Some are more dependent on temperature for release than the fast-release forms.
Lower potential for leaching, denitrification, and volatilization losses.	
Need to be applied less frequently; and with the natural organic forms, they often supply other nutrients.	

Because of advantages and disadvantages of fast- and slow-release forms (Tables 5-10 and 5-11), some turfgrass fertilizers are blends of both types of N carriers to combine the advantages and reduce the disadvantages associated with each.

SPECIFIC CHARACTERISTICS OF SLOW-RELEASE NITROGEN FERTILIZERS

Urea-formaldehyde (UF) and Methylene Urea (MU) are synthesized by combining urea with formaldehyde to form a compound containing units of methylene urea. Available N from UF and MU is dependent upon microbial hydrolysis of the carrier and is temperature dependent. Solubility of these materials is based on molecular weight (chain length) of the formulation. With UF and MU, the shorter the length of subunits, the greater the solubility. The Cold-Water Soluble N (CWSN) fraction of UF and MU represents the low molecular weight short chains of unreacted urea plus methylene urea that are generally more readily available. The Cold-Water Insoluble N (CWIN) fraction of UF and MU represents the larger chain lengths, which have higher molecular weights and are more slowly available. The Hot Water Insoluble N (HWIN) fraction of UF represents the very large chain lengths that have very high molecular weights and are very slowly available. At 77°F, the CWSN of UF and MU is readily absorbed by turfgrasses. Typically, there is little or no response to UF and MU when applied during cold temperatures because microbial activity is minimal. A UF and MU carrier should have an active ingredient (AI) of at least 40% to supply sufficient N. Another expression of UF-N or MU -N solubilization characteristics is the urea:formaldehyde ratio. Since UF and MU are synthesized by combining urea with formaldehyde (unreacted urea plus methylene ureas with varying chain lengths), some commercial formulations are manufactured with a urea:formaldehyde ratio of 1.3:1. This provides about 25% CWSN and 75% CWIN. A more soluble formulation can be obtained with a urea:formaldehyde ratio of 1.9:1, which provides 67% CWSN and 33% CWIN. The solubility of UF and MU can be controlled by the ratio. It is desirable to have some N immediately available and the remaining made available in small amounts with time.

Isobutylidene diurea (IBDU) is formed by combining urea with isobutyraldehyde. The solubility of IBDU is not influenced by microbial activity. Instead, the availability of N is dependent on chemical rather than microbial hydrolysis. With IBDU, N release is faster with smaller particle sizes (low mass:surface area), acidic soil pH, high moisture, and warmer temperatures. Below a pH of 5, the rate of N release for IBDU is very rapid. The rate of N release from IBDU is up to three times faster at 75 ° F than at 50 ° F. Usually, there is a slower response in the spring with IBDU, but N response is faster if applied the previous fall. This allows more time for breakdown over the winter. It is common to blend different particles sizes of IBDU so that N is released over a three- to four-month period. A product with particle sizes ranging between 71 and 238 SGN mesh is best for turfgrasses.

Sulfur-Coated Urea (SCU) and Polycoated Urea (PCU) is formed when urea granules are coated with sulfur and a thin coating of sealant or polymers (wax, resins). Water diffuses in and out of micropores in the coatings until sufficient pressure builds up to cause breakage of the coating. The same factors that affect the N release of UF and ME, also affect SCU (high moisture, higher temperatures). The initial turfgrass response to SCU is fairly rapid and is faster than UF, ME, and IBDU. With SCU, there is nonuniformity in the coating thickness that provides varying release rates. It is common to blend thin-, medium-, and thick-coated granules so that N is released over six- to eight-week period or longer. The N in SCU is not defined as a Water Insoluble N (WIN) and is often listed as a Controlled Release N (CRN) and characterized by a seven-day dissolution rate value. The seven-day dissolution rate value is the percent of N released from SCU in water at 95°F in one week under laboratory conditions. Most commercial SCU formulations have a seven-day dissolution rate of 20% to 30%. This means that 20% to 30% of the N is quickly released and 80% to 70% of the N is slowly released. A dissolution rate below 20% indicates a material with N that is probably too slowly available for most turf purposes and a dissolution rate above 30% indicates a material that would not be considered a slow release N source. Various polymer-coated

urea products are available. These materials control the release of N by diffusion through a polymer membrane that coats the urea. Release rates are dependent on moisture, temperature, and coating's composition and thickness.

Fertilizers Derived from Organic Materials are derived from natural organic sources. These materials are typically lower in N and usually more expensive per unit N than synthetically derived products, but they offer many advantages for use on turfgrass:

- Low potential for foliar burn.
- Low leaching and volatilization potential.
- Slower acidifying effect on soil pH.
- Wide range of both macro- and micro-essential elements.
- Potential improvement of the physical properties of turf soil.

Release of N from organic fertilizers is dependent on microbial activity. Depending on conditions, N release from organic fertilizers can be relatively rapid or may be immobilized within the soil organic matter and not released for months. Most plant and animal meal-based fertilizers above 5% N have a relatively rapid release rate.

NITROGEN FERTILIZER PROGRAMS

Frequency and intensity of fertilization depends on many factors:

- Species. Less fertility is required for the fine leaf fescues, common types of Kentucky bluegrass, tall fescue, and bentgrasses. Higher fertility is required for improved types of Kentucky bluegrass, perennial ryegrass, and annual bluegrass. Annual bluegrass populations can increase under higher N fertility at the expense of bentgrass (Dest and Guillard, 1987).
- Environmental conditions. Shaded turf requires less N than turf growing under full sun conditions. Reduce N rates by one-third to one-half in shaded areas. Higher precipitation and irrigation rates increase the amount of N lost through leaching and denitrification; therefore, more frequent fertilizer application may be required. If high rates of N are applied to cool-season turfgrass during high temperatures, shoot growth will be rapidly stimulated, reducing food carbohydrates and negatively affecting root growth. Disease susceptibility also increases. Turf decline may follow.
- Soil characteristics. Sandy soils and sand-based rootzones require lighter and more frequent fertilizer applications than soils with more clay and silt. Soils with higher organic matter content or more clay and silt require less fertilizer than sandy soils or soils with lower organic matter content.
- Maturity of turf stand. There is a greater fertility need for turf at establishment and less so as the stand matures. As turf matures, there is a greater potential for N losses because the storage potential of the soil organic matter is maximized. Therefore, less N is required on older mature stands of turf than younger, newly established stands of turf.

- Length of growing season. The longer the growing season, the more that nutrients are required to sustain turf growth and quality.
- Mowing height. As mowing height is lowered, individual shoots become smaller and turf density increases. Therefore, smaller amounts of N fertilizer should be applied because higher rates may burn the leaves due to collection of fertilizer on the denser leaf surface.
- Clipping management. Returning clippings to the turf markedly reduces fertilizer needs. On higher-cut turf, UConn studies indicate that N fertilizer rates can be reduced by 50% or more without a loss in turf growth and quality when clippings are returned (Kopp and Guillard, 2002). Contrary to popular belief, grass clippings do not contribute to thatch and are mostly decomposed (>80%) within eight weeks after being returned to the turf (Kopp and Guillard, 2004).
- Traffic and wear. Trafficked golf turf will require more frequent applications of N to recover from traffic and wear.

Because of the different variables, turfgrass fertilization programs are site specific. Therefore, it is important for the turf manager to evaluate each site and its particular combination of features before initiating a fertilization program.

NITROGEN FERTILIZATION FREQUENCY

An ideal turfgrass N fertilization program consists of frequent applications of very small amounts of nutrients during the growing season to meet growth requirements without overapplication. With such a program, the turf manager can meet nutrient needs quickly. The term “spoon feeding” is used for these types of fertilization programs.

Nutrients such as N are applied at rates of approximately 0.10 to 0.25 lb N/1,000 ft²/application or less during the season at seven- to 21-day intervals. For most golf courses, spoon feeding is applicable to the lower-cut turf. For higher-cut turf (roughs and other amenity areas), infrequent N applications (one to three times per year) at higher rates are appropriate.

NITROGEN FERTILIZATION RATE

The maximum amount of N that may be safely applied at any one time depends on the form and carrier of N, temperature, time of year, mowing height, species, and turf use. Here are examples of application rates:

- Under favorable conditions in New England, soluble N-containing fertilizers can be applied on short-cut turf at 0.10 to 0.25 lb N/1,000 ft²/application. For higher-cut turf, N should be applied to provide no more than 1 lb N/1,000 ft² in any one application. Apply soluble N-fertilizers only when turf is dry and when temperatures are below 80 °F. After application, apply water to wash the fertilizer off the foliage. Under hot temperatures, higher rates of soluble N may cause burn or excessive shoot growth.
- Under hot temperatures, N application should be avoided or limited to no more than 0.10 lb N/1,000 ft²/application on lower-cut turf, and 0.5 lb N/1,000 ft² in any one application for higher-

cut turf.

- With the natural organic materials, slightly higher rates may be used. However, no more than 1 lb N/1,000 ft² on lower-cut turf and no more than 3 lbs N/1,000 ft² on higher-cut turf should be applied at any one time. Reduce the number of applications by half when natural organic sources are applied at rates greater than 1 lb N/1,000 ft².
- In New England, an annual total of 2 to 3 lbs N/1,000 ft² is usually sufficient for good growth of short-cut turf. For higher-cut turf, up to 2 to 4 lbs of N/1,000 ft² per year is often applied. Reduce rates when clippings are returned.

NITROGEN FERTILIZATION TIMING

Fertilizer applications should be timed to maximize growth and vigor in the grass plant. Key times to apply fertilizer are spring, late spring, late summer, or early fall. Timing of fertilizer application should be based on environmental conditions and coincide with turfgrass needs. Therefore, time fertilizer application to provide nutrients at the beginning of periods when temperature and moisture conditions favor active turfgrass growth. Higher rates of N fertilization should be avoided when turf is stressed or with shaded turf. Summer application of fertilizer is generally not recommended unless recovery is required from traffic or wear, or where spoon-feeding programs are implemented. Excessive midsummer N fertilization promotes certain hot-weather diseases such as brown patch, summer patch, and Pythium. It also stimulates shoot growth at a low-growth period, which depletes the food carbohydrates in the grass. This reduction in carbohydrates is detrimental to root growth. A rapid reduction in root growth leads to decreased heat, drought, wear, and pest tolerance.

Late-season N fertilization for cool-season turfgrasses has been advocated anywhere from mid-September to mid-December to promote root growth, better color retention in the fall, and earlier green-up in the spring. Late-season N fertilization of cool-season turfgrasses has become a well-established management practice in the Northeast. The rationale behind this practice is based on the beneficial physiological and rhizome/rooting responses of turfgrasses during this period to the applied N.

The positive response of turfgrass to late-season fertilization is physiologically based and related to energy partitioning within the grass plant under decreasing photoperiods and temperatures.

- During the fall, air temperatures are not optimum for shoot growth but soil temperatures are ideal for root and rhizome growth. However, N from fall fertilization enables the turfgrasses to retain leaf color (chlorophyll content) into late fall and, as a result, higher levels of photosynthesis are sustained. Because air temperatures are not conducive for shoot growth at this time, the energy produced is used for root/rhizome growth.
- Higher photosynthetic rates and minimal shoot growth in the fall maintain higher storage carbohydrate reserves in the roots. High levels of carbohydrate reserves in the roots enhance winter survival and spring recovery. This enhanced carryover effect on spring green-up and growth substantially diminishes the need for early season N, which would stimulate top growth at the expense of root development.
- Other reported beneficial effects of late-season fertilization are turf that is more vigorous and less susceptible to diseases during spring re-growth, and turf that enters the summer heat

period in a more hardened, healthy state.

Although agronomic benefits of late-season N fertilization are reported, caution should be used with this practice with respect to N losses from runoff and leaching. Leaching losses of soluble forms of N are much higher during this time of the year than during the active growing season. Also, cool-season turfgrass species and cultivars differ substantially in their N uptake and use efficiencies. Fall or winter fertilizer application might further enhance the leaching potential of some grasses that inherently express poor N-use efficiency. Research conducted at the University of Connecticut indicates that a greater potential exists for nitrate leaching losses with soluble N-based fertilizers than with slow-release or natural organic fertilizers when applied after October 21 (Guillard and Kopp, 2004). Also, leaching losses of up to 66% for soluble N sources when applied to higher-cut turf have been observed in mid-October and later applications (Mangiafico and Guillard, 2006).

Heavy spring applications of N to turf are also not recommended. Heavy application of N in early to mid-spring on cool-season turf may encourage more severe incidence of spring and summer diseases and may reduce root development. During the winter, stored food carbohydrates (energy sources) are utilized by the grass for maintenance. Because environmental conditions do not favor rapid replenishment through photosynthesis during the winter, the grass plant enters a very vulnerable position by the time spring re-growth is required. Energy stores are at their lowest levels and excessive demands for energy by growth can severely weaken the grass. Heavy N applications in the spring stimulate very rapid shoot growth at the expense of the roots.

This results in a weakened turf that is more susceptible to stresses and pests and thins out during the summer.

Phosphorus

Turfgrasses take up P primarily as orthophosphate ($H_2PO_4^-$). Phosphorus forms high-energy compounds that are used to transfer energy within the plant. Although many mineral soils contain relatively large amounts of P, it occurs in forms not available to turfgrass plants. Phosphorus is readily fixed by Ca, Fe, or Al. At a pH below 5.5, Fe and Al form an insoluble complex with P that makes P less available to turfgrass. At a pH above 7.5, Ca complexes with P to form an insoluble complex that makes P less available to turfgrass. It is commonly believed that P is most available to turfgrasses when soil pH is between 6 and 7.

Because phosphorus is a major contributor to eutrophication of waterbodies, proper timing and rates of application should be used to reduce the risk of off-site movement of phosphorus.

The role of P in turfgrass culture is important in seed germination, seedling vigor, and rooting responses. Therefore, P is critical during turfgrass establishment. Fertilization of P should be based on soil tests or tissue tests. When establishing new turf, P should be applied when extractable P levels in a soil test indicate a need. The P-containing fertilizer should be incorporated into the soil before seeding or sodding to a depth of 4 to 6 inches to provide at least 2 lbs P_2O_5 /1,000 ft². On low P sites, the new seedlings or sod should be additionally topdressed to provide at least 1 lb P_2O_5 /1,000 ft² after emergence. With tissue tests, adequate P is available when leaf P concentrations are 0.2% or above on a dry weight basis. For mature turf, P should be applied as a maintenance fertilizer only when soil test extractable P levels read low to provide at least 1 lb P_2O_5 /1,000 ft².

A single application usually provides sufficient P for turfgrass growth for a growing season. There is little turfgrass response to P fertilizers when extractable P levels are medium or above. Continual fertilization of soils testing medium or above for extractable P will result in high soil test P values with no added beneficial responses for turf. High soil test P levels increase the potential for annual bluegrass infestations on short-cut golf turf, leading to a decrease in bentgrass populations. Soils testing high in extractable P also increase the potential for nutrient contamination of surface and ground water by P that leaches or runs off turf.

Fertilization with P is often dependent on N rates. Low maintenance turf with low N input will probably not require much, if any, P fertilization. Under higher N inputs with clippings removed, however, P fertilization may be required. If clippings are returned to the turf, there may be little need to fertilize with P. The residual supply from previous or initial P applications in addition to the recycling of P from the clippings may be adequate for turfgrass needs, provided that the soil pH ranges between 6 and 7. Phosphorus deficiency symptoms initially include reduced shoot growth and dark green color; later, lower leaves may turn reddish at the tips and then the color may progress down the blade.

Most P fertilizers are derived from rock phosphate ores treated with mineral acids. Superphosphate is derived from rock phosphate treated with sulfuric acid to form calcium phosphate and gypsum. Triple superphosphate is derived when rock phosphate is treated with phosphoric acid to form calcium phosphates. Ammonium phosphates are derived when ammonia is treated with phosphoric acid to form mono- and diammonium phosphates.

- P fertilizer sources
 - Superphosphate (SP) – $\text{Ca}(\text{H}_2\text{PO}_4)_2$; 20% P_2O_5
 - triple superphosphate (TSP) – $\text{Ca}(\text{H}_2\text{PO}_4)_2$; 46% P_2O_5
 - Monoammonium phosphate (MAP) – $(\text{NH}_4)(\text{H}_2\text{PO}_4)$; 52% P_2O_5
 - Diammonium phosphate (DAP) – $(\text{NH}_4)_2\text{HPO}_4$; 46% P_2O_5
 - Natural organics

Potassium

Turfgrasses take up large amounts of K as the cation K^+ . Turfgrass leaf concentrations of K can range from 2% to 5% of the dry matter. High K concentrations in turfgrass leaf tissue have been reported to improve tolerance of heat, drought, cold, disease, and wear. It is common to see high-K containing turf fertilizers, which are often called winterizers, promoted for late-season application. However, the effects of high K applications have been inconsistent and not observed in all cases.

Fertilization of K should be based on soil tests or tissues tests. Potassium should be applied only when the soil test extractable levels are low and at a rate to provide at least 1 lb K_2O /1,000 ft². Tissue concentrations below 1% are considered deficient (Jones, 1980) and an N:K ratio of 2:1 in the tissue is considered optimum. There is little turfgrass response to K fertilizers when soil test extractable K levels are medium or above that ratio.

Potassium is a constituent of many soil minerals and weathering of nonexchangeable forms has been

shown to provide a significant amount of K to turfgrasses in New England soils during the growing season, with variation based on soil type and rootzones. K fertilization has been shown to have little effect on bentgrass grown on high sand content rootzones across eight sources in southern New England (Dest and Guillard, 2001). Studies have also shown there has been little to no effect of K fertilization found in wear tolerance and traffic recovery (Hoffman et al., 2010). However, K fertilization beyond what is recommended for optimum shoot growth may increase the winter hardiness of annual bluegrass (Schmid et al., 2016) and perennial ryegrass (Webster and Ebdon, 2005).

If clippings are returned to the turf, probably little if any need to fertilize with K exists. In addition to clipping management, fertilization with K often depends on N rates. Low- maintenance turf with low N input will probably not require K fertilization. Release of nonexchangeable K from soil minerals will probably be sufficient to meet turfgrass needs under these conditions. However, under higher N inputs with clippings removed, K fertilization may be required if clippings are removed over many years. When K fertilization is needed under these situations, it is typical to use a N:K fertilizer ratio of 1:1.

Most K-containing fertilizers are derived from potassium salts such as muriate of potash (KCl, 60% K₂O). Combining KCl with sulfuric acid forms potassium sulfate (50% K₂O), whereas KCl combined with nitric acid forms potassium nitrate (44% K₂O). Muriate of potash has a higher burn potential (salt effect), but is fast acting and less expensive than most other K-containing fertilizers. Potassium sulfate is slower acting and more expensive than KCl and potassium is susceptible to leaching in sandy soils with low CEC. Therefore, overwatering of turf should be avoided to maintain adequate levels and reduce loss of K in the soil.

- K deficiency symptoms

Except under severe, documented deficiencies, K may not have an observable influence on turfgrass quality. Yellowing of older leaves followed by tip dieback and scorching of leaf margins have been reported.

- K fertilizer sources
 - Potassium sulfate – K₂SO₄; 54% K₂O
 - Potassium chloride – KCl; 60% K₂O
 - Potassium nitrate – KNO₃; 46% K₂O



Grass clippings being distributed while mowing instead of removed at Martindale Country Club, Auburn, ME. © Scott Cybulski, CGCS

Best Management Practices for Fertilization

- Apply nutrients when turfgrass is actively growing.
- Apply N at rates and intervals to maintain moderate turf growth and recuperative potential.
- Use light, frequent N applications (spoon-feeding) to provide turf consistent nutrition and minimize potential for leaching and runoff.
- Apply slow-release N fertilizer at the appropriate time of year to maximize the products' release characteristics.
- N application rates from slow-release materials should take into consideration the release rate of the chosen material.
- Select a N:K fertility ratio based on turf use, rootzone, and clippings management.
- Exercise caution when applying nutrient applications during turfgrass establishment as these applications are particularly susceptible to loss via leaching and runoff.
- Provide appropriate rates and products to minimize N loss without reducing turfgrass establishment.
- Be aware of the different types of spreaders and understand the advantages and disadvantages of each.
- Calibrate spreaders regularly to reduce environmental risk and increases profitability. FertAdvisor is a free smartphone application that assists in spreader calibration developed by the University of Connecticut and available for both iPhone and Android users.
- Reduce environmental risk by properly storing and loading fertilizer and cleaning up any spills.
- Avoid applying fertilizer to soils that are at, or near, field capacity or following rain events that leave the soils wet.
- Do not apply fertilizer when the National Weather Service has issued a flood, tropical storm, or hurricane watch or warning, or if heavy rains are likely.

Soil pH

Maintaining soil pH within certain tolerances plays an important role in turfgrass growth and quality. Nutrient availability and soil flora and fauna activities are closely associated with the pH of the soil. These activities are important for mineralization of soil organic matter, thatch, and grass clipping decomposition, severity and incidence of certain turfgrass pests, and influences on pesticide efficacy. Liming does not replace a sound fertilization program, but enhances one. Therefore, the turfgrass manager must understand and appreciate how pH influences the persistence, growth, and quality of turf.

Soil pH is the result of the chemical reactions that occur in the soil, and these reactions affect the degree of acidity or alkalinity of a soil solution. The pH scale is used to measure the effects of these soil reactions. This scale is related to the amount or concentration of hydrogen ions [H⁺] present in the soil solution, and then transformed into a value that is easily understood. Mathematically, the pH value is calculated as the negative logarithm (base 10) of the hydrogen ion concentration [H⁺], and ranges from 0 to 14.

Because the scale is measured using logarithms (base 10), it increases or decreases 10 times for each unit change of pH. For example, even though a pH of 5 does not seem that much lower than a pH of 6, the pH of 5 is 10 times more acidic than the pH of 6. A pH of 4 is only two units lower than 6, but 100 times more acidic.

Soil pH varies throughout New England, but most turfgrass soils are in the pH range of 6 to 7.0. The soil pH is usually a function of precipitation in which greater precipitation or irrigation induces more leaching of Ca²⁺, Mg²⁺, and K⁺, which are replaced by acidic H⁺ or Al³⁺ ions. Native soils of New England, which are located in a humid region of the country and receive 40 to 50 or more inches of precipitation yearly, generally have acidic soils.

Other factors also influence soil pH. The underlying parental material from which soil is formed will affect pH. Most soils of New England are formed from gneiss, schist, or granite that are naturally acidic. Soils high in organic matter also tend to have an acidic pH because organic acids are released from the organic matter as it decomposes.

Fertilization practices may also affect soil pH. Nitrogen applications generally have an acidifying effect (especially the urea and NH₄-based formulations) because H⁺ ions are released when NH₄ is reduced to nitrate (NO₃) in the soil solution.

OPTIMUM RANGES OF SOIL PH FOR TURFGRASSES

Although most turfgrasses can tolerate a wide range of soil pH values, a pH range of 6 to 7 is generally recommended for New England turf. Kentucky bluegrass, a popular turf species, does best when soil pH is between 6.5 and 7.2. Ryegrasses and bentgrasses are somewhat more tolerant of lower soil pH values than Kentucky bluegrass, but they also perform best under a neutral or slightly alkaline pH. The fine fescues and turf-type tall fescues can tolerate fairly acidic soil conditions, but their growth is also better under a neutral or slightly alkaline pH.

At extreme pH values, certain essential elements become less available and others become more available (excessive). Many of the micronutrients remain available at acidic pH, which can create problems with Al and/or Mn toxicities. At acidic pH, there is also a decrease in microbial, earthworm, and other soil flora and fauna activities. This results in a decrease in mineralization and decomposition of organic matter, a potential loss of favorable soil structure, and excessive thatch buildup.

LIMING MATERIALS

When soil pH is acidic, the turf manager needs to neutralize the soil acidity by adding liming materials. The most common liming materials are:

- Calcitic limestone (calcite) - calcium carbonate (CaCO_3), which is 40% Ca and called agricultural grade limestone.
- Dolomitic limestone (dolomite) - $\text{CaMg}(\text{CO}_3)_2$, which is 27% Ca and 13 - 15% Mg and also called agricultural grade limestone.
- Pelletized Lime - small limestone particles in pellet form (calcitic or dolomitic). This reduces dust and makes spreading easier. It dissolves rapidly into powder when it comes into contact with water. This is generally the most common form of liming materials used on golf courses.
- Burned lime or Quicklime - CaO or MgO , which has the fastest reaction in the soil but can burn the turf if applied incorrectly. It is caustic to handle and more expensive than ground limestone. Typically, not recommended for turfgrass.
- Hydrated lime or Slaked lime — $\text{Ca}(\text{OH})_2$ or $\text{Mg}(\text{OH})_2$ which is fast acting but also caustic to handle. It is more expensive than ground limestone and can react with ammonia-containing fertilizers to form ammonia gas that is toxic to grass. Therefore, this material should not be applied with fertilizers at same time. Wait at least two weeks between applications of fertilizer and hydrated lime. Typically not recommended for turfgrass.
- Calcium sulfate (gypsum, CaSO_4) or magnesium sulfate (Epsom salts, MgSO_4) has no effect on pH, but can supply Ca, Mg, or S.

RATES OF LIME APPLICATION

Soil tests are the only way to determine if the turf soil requires lime. The rate for liming materials is partly determined by soil texture. Soils with more clay and silt require more lime to neutralize acidity than sandier soils. Soils with higher organic matter may require more lime than the same soil type with lower organic matter content.

Because most New England soils contain an appreciable amount of sand, it is best to limit each application of ground limestone to 50 lbs/1,000 ft² when applied to established turf. Higher rates may result in excessive alkalinity near the soil surface before the lime eventually moves downward. This is especially the case with turf containing a thick thatch layer. If more than 50 lbs limestone/1,000 ft² are recommended based on a soil test, the applications should be split at least a few months apart. When establishing new turf, the total limestone requirement may be applied in a single application provided that it is thoroughly mixed into a 4- to 6-inch depth before seeding or sodding.

LIME MOVES SLOWLY THROUGH THE SOIL

Lime moves slowly downward in the soil profile at a rate of about 0.5 to 1 inch per year, but not laterally. Surface applications can take two or more years to increase the pH of the rootzone to proper levels. Therefore, the pH must not be allowed to drop too low before adding lime.

Soil tests for pH are recommended every two years. Lime should be added if the pH drops to 6 or below. Lime is generally best applied in the fall so that it can be worked into the soil by freezing and thawing

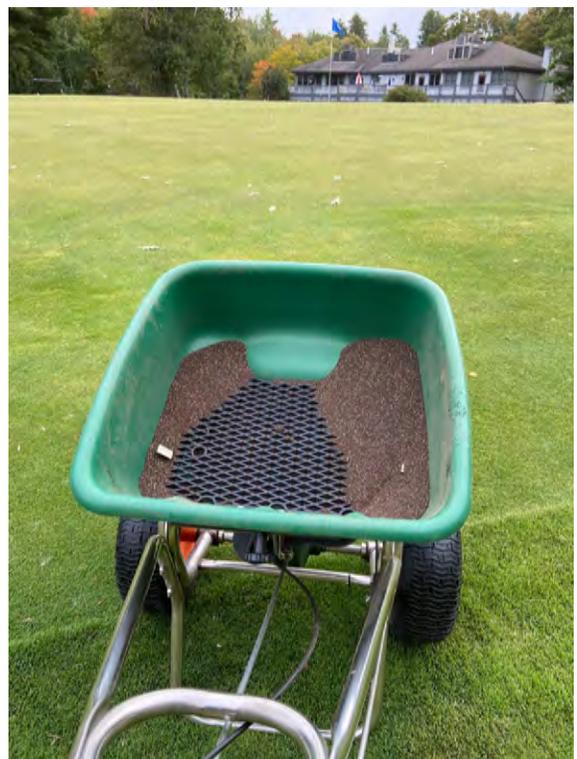
action and has sufficient time for reaction before active regrowth of turf in the spring. Applying lime after core aeration helps move lime more quickly into the rootzone.

CORRECTING SOIL ALKALINITY

If a soil test shows the soil pH above 8 in a turfgrass system, then it must be lowered. Often, this is a result of over liming of sandy soils. The excess sodium (Na^+), Ca^{+2} , Mg^{+2} ions form insoluble complexes with the micronutrients that may induce micronutrient deficiencies. In these situations, the application of sulfur at rates of 3 to 5 lbs/1,000 ft² should decrease pH to more favorable levels. To decrease pH to favorable levels, sulfur can be applied as elemental S, ammonium sulfate or iron sulfate. Calcium sulfate (gypsum), magnesium sulfate (Epsom salts), and potassium sulfate have little effect on soil pH and should be avoided for purposes of lowering soil pH.

Best Management Practices for Soil pH

- Maintain pH near 6.8 to optimize nutrient availability and reduce fertilization requirements.
- To increase soil pH, apply a liming material (calcium carbonate, calcium oxide, dolomitic limestone) that contains Ca^{2+} or $\text{Ca}^{2+}/\text{Mg}^{2+}$ and neutralizes acidity.
- To lower soil pH, products containing elemental sulfur should be applied.
- In some cases, utilizing injection pumps into irrigation water to address pH can be beneficial. Injecting acid (at proper rates) into irrigation water will neutralize alkalinity.



Turf fertilization at Martindale Country Club, Auburn, ME. © Scott Cybulski, CGCS



CHAPTER 6: CULTURAL PRACTICES

© Bob Searle

PREFACE

Cultural practices play a large role in turfgrass quality. In addition to selecting appropriate turfgrass species or cultivar, proper cultural management can help produce a dense, healthy playing surface. These practices are used on all areas of a golf course, including putting greens, fairways, tee boxes, and roughs and include a variety of methods, such as mowing, cultivation, cultivar selection, and rolling. These practices typically manage the top 3 to 4 inches of soil and improve nutrient and water uptake and the overall health of the plant.

Turfgrass Selection

Selection of turfgrass species or cultivar is one of the most important decisions a manager can make to ensure a healthy turfgrass stand. Selecting the wrong species can lead to turfgrass failure, resulting in poor density, poor playability, increased water use, and increased likelihood of pesticide application. Turfgrass managers should select grass species and cultivars based on the existing site conditions and the intended use of the turf. Criteria include the selection of:

- Drought-tolerant species and cultivars where water is limited or not available.
- Wear- and compaction-tolerant species and cultivars for heavy play and high traffic areas.
- Disease-tolerant and endophytic cultivars to reduce pest damage and pesticide use.
- Shade-tolerant species and cultivars for areas with limited or restricted light.

Breeding programs have made tremendous advances in the development of improved turfgrass cultivars. Within each turfgrass species, cultivars can now be selected for improved characteristics such as denser playing surfaces, tolerance of lower mowing heights, increased drought tolerance, improved

When planning new seeding or overseeding projects for the golf course, it is recommended that turfgrass mixtures (two or more species) or blends (two or more cultivars of the same species) be used to **improve genetic diversity** of the turfgrass stand.

Val Halla Golf Club, Cumberland, ME. © Toby Young



wear tolerance, improved pest and disease resistance, improved shade tolerance, and improved salinity tolerance. As a result of these improvements, managers can select cultivars or species appropriate for the site that require less water, pesticide, and fertilizer.

When planning new seeding or overseeding projects for the golf course, it is recommended that turfgrass mixtures (two or more species) or blends (two or more cultivars of the same species) be used to improve genetic diversity of the turfgrass stand. Consideration should be given to match turfgrass characteristics such as color, texture, growth rate, and mowing height requirement of grasses in the mix or blend.

Examples of typical grass species and blend or mixture recommendations for cool- season grasses in New England can be found in Table 6-1.

Table 6-1. Recommended turfgrass species for New England Golf Courses

Area of Golf Course	Species	Recommended Mixture or Blend
Putting Greens	Creeping bentgrass	Single Cultivar ¹ or Blend
	Velvet bentgrass	Single Cultivar ¹ or Blend
Tees and Fairways	Creeping bentgrass	Blend
	Perennial Ryegrass	Blend
	Compact Kentucky bluegrass	Blend
	Compact Kentucky Bluegrass/ Perennial ryegrass	Mixture
	Compact Kentucky Bluegrass/Fine fescue	Mixture
	Fine leaf fescue/colonial bentgrass ²	Mixture ² Minimally irrigated areas
Roughs ³	Kentucky bluegrass	Blend
	Kentucky bluegrass/ Perennial Ryegrass/fine fescue	Mixture
	Turf-type Tall fescue ⁴ (minimally irrigated roughs)	Blend
<p>¹ Most putting green complexes are seeded with a single cultivar. However, newer cultivars have been developed that have similar growth characteristics. Therefore, it is possible to use two or more cultivars in one blend.</p> <p>² Low input (fertility and water requirements).</p> <p>³ Rough areas are expansive and there are multiple microclimates throughout the golf course. Different species, blends, and mixtures may be utilized throughout the property.</p> <p>⁴ Less reliable performance in northern New England.</p>		

When considering new turfgrasses, managers can consult the [National Turfgrass Evaluation Program](#), the [Alliance for Low-input Sustainable Turf](#) (A-List Turf), or the [Turfgrass Water Conservation Alliance](#) to select cultivars that perform well in their region. Attending educational conferences, seminars, and turfgrass field days are other ways turfgrass managers can learn about new and improved cultivars (Figure 6-1).



Figure 6-1. National Turfgrass Evaluation Program fine fescues traffic trial for University of Connecticut. The study evaluated the use of low input (low water and fertility) for fine fescue species and cultivars maintained at golf course fairway mowing heights. Photo from New England Regional Manual.

Best Management Practices for Turfgrass Selection

- Identify site characteristics and use requirements that may impact turfgrass growth and performance.
- Select appropriate grass species and improved cultivars for the site and intended use with the following characteristics as needed:
 - Lower mowing height of cut
 - Increased density
 - Improved shade tolerance
 - Improved heat tolerance
 - Improved drought tolerance
 - Reduced disease susceptibility
 - Reduced insect susceptibility
 - Improved traffic (wear + compaction) tolerance

Mowing

Mowing is the most common cultural practice of managed turfgrass systems. Turfgrasses are unique in that they tolerate routine mowing at heights from 0.09 to 4 inches depending on the turfgrass species and/or cultivars. When considering mowing, five areas of interest affect turfgrass quality and playability: height of cut (HOC), mowing frequency, clipping management, mower selection, and mowing direction. Additionally, the type of roller (i.e. smooth or grooved) on the front of the mower can also affect mowing quality.

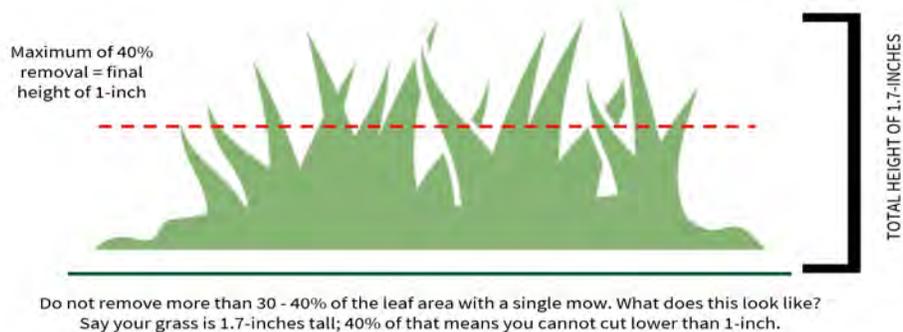
MOWING HEIGHT

Turfgrass HOC is determined by several factors, including, but not limited to, the following: species and cultivar, area of play (tee, green, fairway, etc.), budget, number of rounds, and tournament or non-tournament conditions. Examples of typical mowing heights and desirable ranges for New England golf turfgrass, are provided in Table 6-2. In general, no more than 30% to 40% of the leaf blade should be removed in a single mowing (Crider, 1955). On turf stands that have been previously maintained at a higher mowing height, or turf that has not been mowed for an extended length of time (i.e. too wet to mow), the HOC should be lowered gradually and in weekly intervals.

Table 6-2. Mowing heights* commonly utilized for golf course playing surfaces in New England

Species	Greens Regular Membership play	Greens Tournament conditions	Collars, Tees, and Approaches	Fairways	Rough (primary)
	<i>inches</i>				
Creeping bentgrass	0.1 - 0.14	0.09 - 0.125	0.25 - 0.4	0.35 - 0.5	-
Velvet bentgrass	0.1 - 0.14	0.09 - 0.125	0.25 - 0.4	-	-
Perennial ryegrass	-	-	0.4 - 0.5	0.4 - 0.5	1.5 - 4
Kentucky bluegrass **	-	-	0.5	0.5 - 0.14%	1.5 - 4
Fine Fescue	-	-	0.4 - 0.5	0.4 - 0.5	-
Tall fescue (turf-type)	-	-	-	-	1.5 - 4

Mowing heights may need to be adjusted based on weather conditions, time of year, turfgrass health, the presence of abiotic or biotic stresses, and the growing environment. For example, mowing HOC should be raised for turf grown under shade conditions. Doing so allows for increased leaf area that maximizes photosynthesis (Gardner and Goss, 2013).



Maintaining an optimal root-to-shoot ratio is critical for plant health. Mowing at an HOC less than the desirable range negatively affects the plant (Figure 6-2). Lower mowing heights result in decreased root growth (Liu and Huang, 2002) as well as decreasing rhizome production (Juska et al. 1955). When this occurs, drought tolerance is reduced, susceptibility to root-feeding insects and pathogens can potentially increase, and overall plant vigor decreases (Steinke and Ervin, 2013). Turfgrasses maintained below their optimum HOC can also result in increased weed pressure (Calhoun, et al. 2005). To maintain plant health when mowing turfgrasses below their range of adaptation, greater inputs of water, fertilizer and pesticide will be needed.



Figure 6-2. Operator error resulted in scalped turf. Photo from New England Regional Manual.

MOWING FREQUENCY

Mowing frequency is primarily a function of mowing height. Turf maintained at lower mower heights should be mowed more frequently to avoid removing too much leaf tissue in a single cutting. Frequent mowing increases shoot density and tillering, which can improve playability.

Mowing frequency has been shown to have an impact on root to shoot ratios. Krans and Beard (1985) reported that root:shoot ratios were greater for Merion Kentucky bluegrass plants that were clipped semiweekly compared with plants that were cut weekly or biweekly. Mowing strategies that can help maintain optimal root to shoot ratios are as follows:

- When possible, reduce mowing frequency when grasses are suffering from biotic or abiotic stresses, or from mechanical damage as a result of cultural practices.

- Do not remove more than 30% to 40% of the leaf area with a single mowing (Crider, 1955). In order to follow this rule, mowing frequency should be based on factors such as time of year, fertility level, water availability, and, ultimately, the rate of grass growth.

MOWING EQUIPMENT

Reel and rotary mowers are the two main types of mowers used on the golf course for maintaining areas of play (Figures 6-3 and 6-4). Quality of cut for both reel and rotary mowers depends upon sharp, well-adjusted blades. Dull blades increase the likelihood of wounding leaf tissue, increased water loss, and increased potential for infection or disease (Figure 6-5). Reel mowers provide the best quality of cut at mowing heights less than 1.5 inches. Generally, the lower the HOC the more blades there are on the reel cylinder. The combination of the number blades on the reel, the reel speed (rotational velocity), and the forward speed of the mower make up the clipping rate. It is critical that the clipping rate matches the HOC to provide the most uniform playing surface. Rotary mowers provide acceptable mowing quality when used at mowing heights equal to or greater than 1 inch. (Rotary mowers are more commonly used at heights greater than 2 inches.)



Figure 6-3. Reel mowers are used on greens, collars, tees, approaches, and fairways. Proper reel to bed knife adjustment is necessary to maintain plant health. Photos from New England Regional Manual.



Figure 6-4. Rotary mowers are used for rough and surrounds (bunker, green, and tee). Sharp rotary blades are necessary for a quality cut and plant health. Photos from New England Regional Manual.



Figure 6-5. Dull mower damage to turfgrass. Properly adjusted mowers minimize turf stress. Photos from New England Regional Manual.

CLIPPING MANAGEMENT

Turfgrass clippings are a source of nutrients. Research has shown that nitrogen fertilization rates can be cut by as much as one-half when clippings are returned (Kopp and Guillard 2002). Therefore, clippings should be returned to the site during the mowing process unless the presence of grass clippings will have a detrimental impact on play. Other times that clippings should be removed are during disease outbreaks, when practicing *Poa annua* control strategies (Gaussoin and Branham, 1989), or when weed seed production is high.

In areas where clippings cannot be returned, such as putting greens, they may be collected and either composted for future use or dispersed in areas such as roughs or practice ranges. On fairways and tees, clippings can be dragged back into the turf canopy with the use of a metal chain pulled between two golf carts or blown into the rough (Figure 6-6). Because nutrients contained in clippings can be a source of pollution, they should be handled properly and care taken to avoid depositing clippings into ditches, swales, wetlands, ponds, and streams or onto impervious surfaces like cart paths or parking lots.



Figure 6-6. To return nutrients back to the soil, collected clippings should be composted, blown, or spread into roughs and out of play areas on the golf course. Photos from New England Regional Manual.

ROLLING

Rolling putting greens has proven to be beneficial in maintenance programs (Figure 6- 7). It provides for a smoother (“truer”) putting surface and increases putting green speeds. Both benefit playability. Rolling allows managers to either reduce mowing frequency or raise mowing heights while maintaining acceptable putting green speeds. Rolling has also been shown to reduce incidence of dollar spot (Nikolai et al., 2001). To reduce the potential of compaction or spread of diseases, rolling should be avoided when soils are saturated or fungal diseases are active.

Brunswick Golf Club, Brunswick, ME. © Allan Greenleaf



Figure 6-7. Rolling putting green surfaces can allow managers to reduce mowing frequency or raise mowing heights while maintaining putting green speeds.

Best Management Practices for Mowing

- Maintain turfgrass mowing heights within the ranges of adaptation for the species and cultivars being grown.
- Avoid removing more than 30% to 40% of the total leaf area in a single mowing.
- Reduce mowing frequency when turf is suffering from biotic and abiotic stresses.
- Alternate between rolling and mowing when turf shows signs of stress.
- Increase mowing height and roll greens routinely (e.g., every other day) to maintain ball roll distance and turf health.
- Properly adjust mowers and sharpen blades to maintain mowing quality and reduce the possibility of disease or infection through wounded plant tissue.
- Return clippings to the turf when possible and account for the nutrients they contribute to the fertility program.
- Remove clippings during periods of weed seed production, to reduce disease spread, to eliminate potential smothering of turfgrass plants from excessive clipping volume, or when clippings interfere with functional use of the turf.
- Do not dispose of or compost clippings near ponds, streams, and waterways or on impervious surfaces.

Using a greens roller. © Bob Searle



Cultivation

Turfgrasses are unique in three ways: they tolerate frequent close mowing; they persist under traffic conditions; and they form a dense, contiguous community. These characteristics make turfgrasses ideal for functional outdoor spaces like golf courses.

However, high traffic areas such as fairways, tees, and putting greens can deteriorate with routine use.

The negative impacts of soil compaction and high wear will be evident in concentrated traffic areas. Thatch accumulation can be problematic in less trafficked areas. The surface of the soil profile (top 3 inches) needs to be actively managed to enhance turfgrass health by improving water movement, increasing atmospheric gas exchange, reducing root penetration resistance, and removing thatch accumulation. Accumulation of excessive thatch and organic matter will reduce root growth, reduce water infiltration, cause scalping, create an undesirable playing surface, and encourage disease and insect activity

Cultivation involves disturbing the thatch and/or soil through the use of various methods such as hollow-tine cultivation, solid-tine cultivation, slicing, spiking, water injection, air injection, verticutting, drill aerification, and deep-tine cultivation (Figure 6-8).

Depending on equipment used, goals, and turfgrass growth rate, cultivation techniques can result in disturbance of the playing surface that can require significant time for recovery. The level of disruption depends on the type of cultivation selected. Type and frequency of cultivation should be based on traffic intensity, growing conditions, degree and depth of soil compaction, and the amount of thatch accumulation.

© Bob Searle



Figure 6-8. Hollow-tine cultivation on a tee.

Best Management Practices for Cultivation

- Conduct more aggressive techniques such as hollow-tine cultivation only when grasses are actively growing to aid in quick recovery of surface density (typically during or just before periods of rapid root growth such as spring and fall).
- Design core cultivation programs to remove 15% to 20% of the surface area per year on sand-based putting greens. This typically will require two core cultivation treatments annually.
- Vary depth of aerification events every two to three years to prevent compacted subsurface layers in the soil profile. This can include varying the length of tines used, but ideally deep-tine equipment that can reach depths of 6 inches or more should be used.
- Use less aggressive types of cultivation such as small solid tines (needle tines), water injection, or air injection during more environmentally stressful periods to vent the surface (maintain gas exchange) and maintain infiltration rates (Murphy and Rieke, 1994; Carrow, 2003).
- Conduct shallow vertical mowing (blades do not penetrate the soil surface) monthly to prevent thatch accumulation and stimulate new growth to increase shoot density by cutting stolons.
- Only use more aggressive vertical mowing (blades reach the bottom of the thatch layer into the immediate soil surface) as a curative approach for thatch removal once the thatch layer reaches a 0.25-inch depth.
- Apply sand topdressing concurrently with more aggressive forms of cultivation such as hollow-tine cultivation (cores harvested, holes filled with topdressing) or aggressive vertical mowing to help maintain macroporosity at the surface, fill voids to smooth the playing surface, and reduce organic matter accumulation (Carrow, 2003).

Topdressing

The objectives of sand topdressing are to 1) dilute thatch accumulation, 2) smooth the playing surface, 3) maintain surface drainage, 4) increase infiltration, 5) increase soil macroporosity at the surface by increasing the sand content of the soil, and 6) increase surface firmness (Figure 6-9).

The goal of topdressing is to keep the crown of the turfgrass plant as close to the soil surface as possible by physically removing organic matter and thatch through cultivation and adding desirable rootzone material to the surface by sand topdressing. Obtaining this goal through proper management enables the turfgrass plant to maximize root development, minimize any disruption in water or air movement, and minimize pest pressure (disease/insect).

The particle size of topdressing material must be compatible with the existing rootzone material. Topdressing materials should have the same particle size distribution as the construction mix or be coarser in texture. Topdressing materials finer in texture than the original construction sand can negatively impact rootzone infiltration rates and result in excessive moisture retention in the topdressing layer. Soil modification with sand of the top 3 inches results in higher infiltration rates and reduced runoff.



Figure 6-9. Topdressing can help maintain good soil physical properties in high traffic areas.

Best Management Practices for Topdressing

- ❑ Apply higher rates of topdressing to putting greens in the spring and fall in conjunction with more aggressive forms of cultivation, harvest cores and fill holes with topdressing (Carrow, 2003). Apply lighter, more frequent sand applications (every seven to 14 days) throughout the growing season.
- ❑ Laboratory test prospective topdressing materials using [ASTM F1632](#), also known as the Standard Test Method for Particle Size Analysis and Sand Shape Grading of Golf Course Putting Green and Sport Field Rootzone Mixes. Compare the results to USGA guidelines for particle size distribution to determine the suitability as potential topdressing materials.
- ❑ Laboratory test prospective topdressing materials using [ASTM F1815](#), also known as the Standard Test Methods for Saturated Hydraulic Conductivity, Water Retention, Porosity, and Bulk Density of Putting Green and Sports Turf Rootzones, to ensure they meet USGA guidelines for hydraulic conductivity.
- ❑ Sample existing greens on the golf course (15 to 20 subsamples at 4 inch depth or to the current topdressing layer depth if previously topdressed) and laboratory test using [ASTM F1632](#), and compare with the results with prospective topdressing materials to ensure compatibility.

Plant Growth Regulators

Plant growth regulators (PGRs) are frequently used to reduce clipping yield, improve stress tolerance, and improve turfgrass quality and performance. An additional benefit of using PGRs is a reduction in the use of other inputs (e.g. fertilizers). Plant growth regulators require frequent reapplication during the growing season to maintain consistent growth suppression, but excessive PGR use can result in a number of undesirable side effects. These side effects can include mild discoloration, stressed turfgrass, and segregation of grasses. These effects can be confused with disease, can slow recovery, and can intensify damage from pests and traffic.

The best approach to planning PGR applications is to use growing degree day (GDD) thresholds instead of a calendar-based schedule. Tools are available online for assistance in using GDD information to schedule PGR applications, such as the web- based app [GreenKeeper](#) and Cornell University's [ForeCast](#) web site.

Best Management Practices for PGRs

- Use GDD to plan PGR use.
- Plant growth regulators should not be applied too early or too late in the growing season to avoid stressing turfgrass.

Val Halla Golf Course, Cumberland, ME. © Toby Young





CHAPTER 7: INTEGRATED PEST MANAGEMENT

© Bob Searle

PREFACE

When turfgrasses face stresses such as the heat, humidity, and occasional drought in some areas of New England in the summer, pests can become a problem. Pesticides alone will not control pests. A more effective approach is to develop an integrated pest management (IPM) program to reduce pest damage and reliance on pesticides. The United States Environmental Protection Agency (USEPA) defines IPM as an “effective and environmentally sensitive approach to pest management that relies on a combination of common-sense practices.”

The primary objective of an IPM program is to reduce the total pesticide load on the golf course by using a number of tactics to control or manage pests. This approach considers all strategies to reduce pest damage to acceptable levels in the most economical means, while simultaneously accounting for impacts on humans, property, and the environment.

IPM OVERVIEW

IPM is comprised of a range of pest control methods or tactics designed to prevent pests (insects, pathogens, nematodes, weeds, etc.) from reaching economically or aesthetically damaging levels while creating the least risk to the environment. IPM programs have basic components that provide the opportunity to make informed decisions on the control of pests at a golf course. Five basic steps for an effective IPM program for turf are as follows:

Step 1: Monitor pests and their damage and record information.

Step 2: Identify pests and understand their biology.

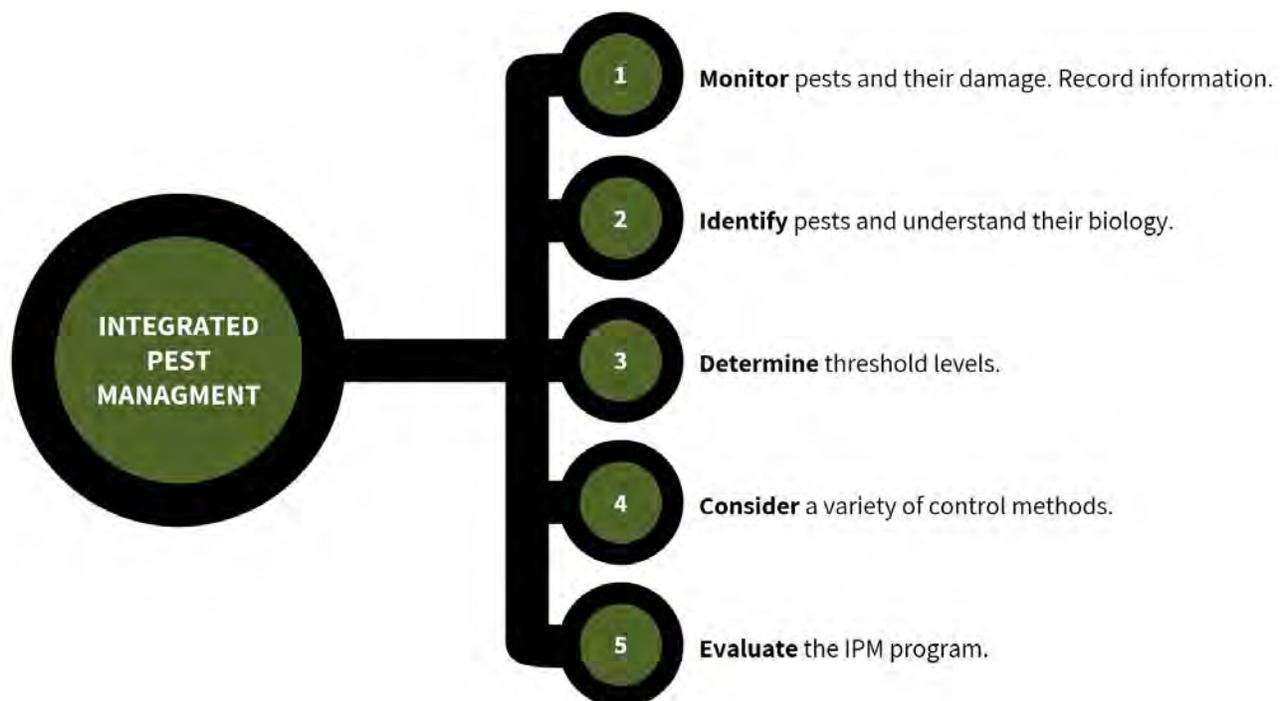
Step 3: Determine threshold levels.

Step 4: Consider a variety of control methods.

Step 5: Evaluate the IPM program.

IPM is flexible, and superintendents can usually balance course quality and environmental goals through its implementation. Growing healthy turf is the best and first line of defense against pests. For example, cultural conditions that predispose turfgrass to diseases include close mowing, inadequate or excessive nitrogen fertility, frequent or excessive irrigation, inadequate thatch management, poor drainage, and shade. Following cultural and nutrient BMPs can help alleviate these conditions. However, under the right conditions, pests can sometimes cause excessive damage to highly managed turfgrass.

A number of non-chemical and chemical control options are available. When chemicals are needed, selection of an appropriate pesticide should follow an evaluation process that considers potential impacts on beneficial organisms and the environment, as well as the potential for development of pesticide resistance. Pesticide products should be rotated, based on their resistance classification.



Best Management Practices for IPM

- Develop a facility-specific, written IPM plan. Available resources for writing an IPM plan include the Golf Course Superintendents Association of America's IPM information and online tools.
- Select turfgrass cultivars and species recommended for use in areas with similar climate and best suited for the intended use and environmental conditions of the specific site.
- Correct the soil's physical and chemical properties that may impact turfgrass health and its ability to resist pests.
- Evaluate the potential impact of the timing of cultural practices and nutrient applications on the incidence of pest problems.
- Use a defined pesticide selection process to select the most effective pesticide with the lowest toxicity and least potential for off-target movement.
- Document all IPM-related activities, including non-chemical control methods and pesticide usage.

Monitoring Pests and Recording Information

In an IPM plan, pest monitoring or “scouting” efforts should be described for all areas of the course such as putting greens, tees and fairways, roughs, and landscaped areas.

Scouting methods include visual inspection, soil sampling, soap flushes, and trapping for insects. Additional monitoring efforts can include weather tracking, which is especially helpful for predicting potential disease outbreaks. Here is one potential scouting schedule: daily on putting greens, at least weekly on tees and fairways, twice a month on roughs, and whenever the potential for pests increases due to weather. For example, warmer temperatures combined with high humidity favor the development of diseases such as dollar spot and brown patch.

When pests are discovered during monitoring, the pest pressure should be quantified with measurements such as:

- Number of insects per unit area.
- Disease patch sizes.
- Percent of area affected.

Documentation should include useful information such as photographs, delineation of pest boundaries on an area map, outbreak date, description of the prevailing weather conditions, and recent management practices. This information can be used to build a database for reference in future seasons and for updating the IPM plan.



As part of their integrated pest management (IPM) approach, Riverside Municipal Golf Course undertook a project to reduce pesticide application on their course in 2016. This involved grouping holes on a three-year rotation, with one year receiving pesticide treatment and two years of no treatment. Riverside hired FB Environmental Associates to conduct annual systematic monitoring of the location and density of undesirable plants and insect-caused turf damage within the project areas. An example of that monitoring is displayed on the map above and highlights the major pest-related damage observed. © FBE

Best Management Practices for IPM Monitoring

- Monitor prevailing environmental conditions for their potential impact on pest problems.
- Train personnel how to regularly monitor pests by scouting or trapping.
- Identify alternative hosts and overwintering sites for key pests.
- Assess pest damage when it occurs, noting particular problem areas, such as the edges of fairways, shady areas, or poorly drained areas.

Best Management Practices for IPM Monitoring (continued)

- Document when the damage occurred. Note the time of day, date, and flowering stages of nearby plants.
- Map pest outbreak locations to identify patterns and susceptible areas for future target applications.

Identifying and Understanding Pests

Once detected, pests must be properly identified. Understanding the biology of pest species and their vulnerable life stages assists in later control efforts. Just as important as identifying pests are recognizing and understanding beneficial organisms and their life cycles so their populations are not unduly affected while managing pests.

Superintendents and staff should continually hone their diagnostic skills by attending training seminars and field days, obtaining reference materials, and providing peer-to-peer training.

DISEASES

Cool-season turfgrasses are susceptible to a number of diseases. In many cases, diseases develop when conditions are favorable, regardless of management strategies. However, the severity of disease can often be greatly reduced by using cultural, biological, and genetic techniques. As a rule, healthy, well-managed turf better withstands disease outbreaks and recovers more rapidly than unhealthy turf.

In order to effectively treat turf diseases and implement an IPM program, it is important to know which disease is most likely to be active. Managers who do not understand disease pathology will risk treating the symptom, rather than the underlying disease.

Turf diseases are typically most common in the summertime for cool-season grasses and in the spring and fall for warm-season grasses. These diseases occur largely due to the shift in growth habits of the grasses from active growth to survival, giving a competitive advantage to disease pathogens.

Understanding the potential diseases for a given species or cultivar and the environmental conditions associated with them is essential. In situations where diseases develop, proper diagnosis assists with decisions on how best to proceed. Diagnostic services, which are often available from a state land grant university's plant disease clinic or from private laboratories, can help prevent choosing the wrong products or management tactics. Some of the more common golf turfgrass disease problems are described in Table 7-1.

Table 7-1. . Common Cool-Season Turfgrass Diseases

Conditions Favoring Disease Development	Disease (Common Names)
Low N	dollar spot anthracnose brown ring patch
High N	brown patch Pythium diseases snow molds (<i>Microdochium</i> patch and <i>Typhula</i> blight)
General	Fairy ring caused by various basidiomycete fungi (both cool- and warm-season grasses) leaf spots and melting out

WEEDS

Weeds are unwanted plants that are unsightly, disrupt playability, harbor pests, and competitively displace desirable turfgrass. Weeds exploit openings in the turfgrass canopy, where seedlings germinate and survive to become a persistent colony of perennials or seed producing annuals.

The potential for invasive weeds can be limited through implementation of the BMPs related to turfgrass selection, nutrient management programs, irrigation, and cultural practices. For example, sites that are over-irrigated may have higher densities of weeds, such as green kyllinga or yellow nutsedge. Cultural practices, such as mowing height and frequency can also impact turf weed populations. For example, mowing too low can open the canopy and provide a competitive advantage to germinating weeds. Because of the importance of soil quality in growing healthy turf, emphasis should be placed on soil testing for the maintenance of turf that can withstand pressure from weeds.

NEMATODES

In the Northeast, high populations of plant-parasitic nematodes are generally restricted to golf greens, which provide an ideal environment for nematodes. The pore spaces of sandy soils provide a favorable environment to support nematodes: adequate oxygen levels, room to accommodate nematode mobility, and water availability. Plant-parasitic nematodes adversely affect turfgrass health by debilitating the root system of susceptible species, thus decreasing the efficiency of plant water and nutrient uptake. Turf weakened by nematode infestations favors further pest infestation, especially weeds. Over time, turf in the affected areas thins out and, with severe infestations, may die. Turfgrass often begins showing signs of nematode injury during additional stresses such as drought, high or low temperatures, and wear.

INSECTS/ARTHROPODS

Annually recurring insect pest groups common to New England golf courses include numerous species, including billbugs, chinch bugs, and Lepidoptera order members such as armyworms, cutworms, and sod webworms.

Several species of grubs occur in New England, including the Japanese beetle, European chafer, Oriental beetle, annual bluegrass weevil, and black turfgrass ataenius. White grubs can destroy significant areas of turfgrass, with damage appearing in summer. Summer drought stress and insufficient irrigation may compound the damage to turf by grubs. Management of white grubs is most efficient when the specific population causing turf damage is identified. Because some insecticides are less effective against Oriental beetle or European chafer, species identification has become increasingly important for management decisions.

Best Management Practices for Identifying and Understanding Pests

- Identify key pests in the IPM plan.
- Determine the pest's life cycle and know which life stage to target (e.g., for insect pests, whether it is an egg, larva/nymph, pupa, or adult).
- Identify pests accurately. For diseases, correctly identifying the disease pathogen often involves sending samples to a diagnostic laboratory.

© Bob Searle



Determining Pest Threshold Levels

A key feature of IPM programs is the identification of tolerance thresholds. Thresholds are based on the pest population, the stage of the pest, and the life stage of the plant. Injury thresholds represent the pest level population that causes unacceptable injury. Treatment thresholds are less than the injury threshold and indicate the number of pests or level of damage that would justify treatment to prevent the pest population from causing unacceptable turf loss.

Best Management Practices for Identifying and Understanding Pests

- Establish injury and treatment thresholds levels for key pests.
- Document pest thresholds in an IPM plan.

Control Methods

Once a pest problem reaches the established treatment threshold, different methods can be used to control the problem, including cultural, mechanical, biological, and chemical. Selecting the most appropriate approach depends on a number of factors, including the site-specific location on the golf course, efficacy of non-chemical controls for the particular situation, economics, and pest populations.

CULTURAL CONTROLS

Cultural practices, especially irrigation, mowing, topdressing, core cultivation, and venting, greatly affect both short- and long-term plant health. Using and/or altering cultural practices, especially in times of stress, to keep plants and soil healthy helps turf to better withstand pest pressure (Table 7-2). It is important to recognize that turfgrass management practices such as core aeration and sand topdressing, while beneficial, can also stress turfgrass.

MECHANICAL OR PHYSICAL CONTROLS

Mechanical methods, such as vacuuming, or physical control methods, such as hand pulling weeds, exclude or remove pests, though these methods may be time consuming and work best when pest populations are low.

PRESCRIBED BURNS

As many golf courses convert maintained turfgrass areas to native grassed areas, many facilities use prescribed or controlled burns to reduce undesirable plants, including noxious weeds, and to encourage desirable species, enrich wildlife, and remove excessive plant debris. Prescribed burns are especially effective in suppressing non- native species and woody plant materials and can be used to create a links-style course that resembles a tallgrass prairie.

Any local notification requirements should be followed as required and all fire danger information should be reviewed before conducting a prescribed burn.

Table 7-2. Effect of Various Cultural Practices on Turfgrass Diseases.

Cultural Practice	Disease	Response	References
Fertilization	Anthracnose	Weekly applications of nitrogen at 0.1 lb./1,000ft ² reduced disease severity.	Inguagiato et al., 2008
		Maintaining a foliar N concentration equal or greater than 3.4% reduced anthracnose severity.	Inguagiato and Guillard, 2016
	Brown patch	Brown patch was more severe in plots treated with nitrogen, but nitrogen did not affect fungicide performance.	Fidanza and Dernoeden, 1996
	Dollar spot	Dollar spot was less severe in plots treated with nitrogen.	Williams et al., 1996
	Pythium blight	Pythium blight severity increased with nitrogen application.	Moore et al., 1963
	Red thread	Disease was more severe in nitrogen-deficient turf.	Cahill et al., 1983
	Summer patch	Patch severity was reduced with application of ammonium sulfate compared with calcium nitrate.	Thompson et al., 1995
Take-all patch	Acceptable levels of control were achieved in plots treated with ammonium chloride.	Dernoeden, 1987	
Mowing	Anthracnose	Anthracnose severity was reduced with increased mowing height.	Inguagiato et al., 2009
	Brown patch	Clipping removal had no effect on disease.	Settle et al., 2001
	Dollar spot	Morning mowing reduced disease.	Ellram et al., 2007,
		Clipping removal had no effect on disease.	Williams et al., 1996
	Pythium blight	Clipping removal had no effect on disease.	Williams et al., 1996
Irrigation	Anthracnose	Minimizing drought stress, while avoiding continuous high soil water content reduced disease severity.	Roberts et al., 2011
	Brown patch	Daily irrigation did not affect brown patch.	Settle et al., 2001
		Irrigation reduced brown patch severity.	Rowell, 1951
		Daily irrigation reduced brown patch severity on perennial ryegrass.	Jiang et al., 1998
Pythium blight	Daily irrigation aggravated Pythium blight in some cases.	Settle et al., 2001	

Cultural Practice	Disease	Response	References
Other	Anthracnose	Light-frequent or heavy-infrequent sand topdressing resulted in lower anthracnose severity.	Inguagiato et al., 2012
		Lightweight rolling every other day reduced anthracnose.	Roberts et al., 2012
		Shallow verticutting did not affect anthracnose severity.	Inguagiato et al., 2008
	Dollar spot	Rolling to remove dew reduced disease.	Williams et al., 1996
		Dew displacement reduced disease.	Ellram et al., 2007
		Dew displacement improved efficacy of chlorothalonil.	McDonald et al., 2006
		Lightweight rolling daily reduced dollar spot.	Giordano et al., 2012.

Table adapted from *A Practical Guide to Turfgrass Fungicides*. Latin, 2011. American Phytopathological Society. St. Paul, MN.

BIOLOGICAL CONTROLS

The biological component of IPM involves the release and/or conservation of natural predators, such as parasites and pathogens, and other beneficial organisms. Several organisms known to have some efficacy against turfgrass pests have been marketed as pest control products, such as such as *Bacillus licheniformis*. Natural enemies (e.g., ladybird beetles, green lacewings, and mantids) of some insect pests may be collected or purchased and released near pest infestations. Areas on the golf course can also be modified to better support natural predators and beneficial organisms, especially in landscaped areas.

PESTICIDES/CHEMICAL CONTROLS

Chemical control is an acceptable IPM practice when other methods will not alleviate the pest problem. In addition to traditional chemical control, reduced-risk pesticides and biopesticides provide a number of advantages over conventional pesticides and should be considered if applicable. The selection and use of any pesticide should follow a selection process and these criteria:

- Use a recommended product to treat a correctly identified pest.
- The pesticide should be effective in treating the pest problem.
- The timing of the pesticide application should be based on growing degree day (GDD) information for the pest to be controlled. [GDDTracker](#) is an example of a tool that can assist in timing applications; Cornell University's [ForeCast](#) website also has GDD information.
- Pesticide rotation, based on resistance classification, as classified by the [Fungicide Resistance Action Committee \(FRAC\)](#), [Herbicide Resistance Action Committee \(HRAC\)](#), and [Insecticide Resistance Action Committee \(IRAC\)](#).

- Costs should be considered.
- Environmental risk and potential for water quality impacts must be evaluated. Tools to evaluate risk are described in the Pesticide Risk Assessment Tools section of the “Pesticide Management” chapter of this document.
- Any restrictions on the pesticide label must be reviewed and rigorously followed.

The use of all pesticides must follow the label and adhere to state and federal regulations, as described in the “Pesticide Management” chapter.

Reduced-Risk Pesticides and Biopesticides

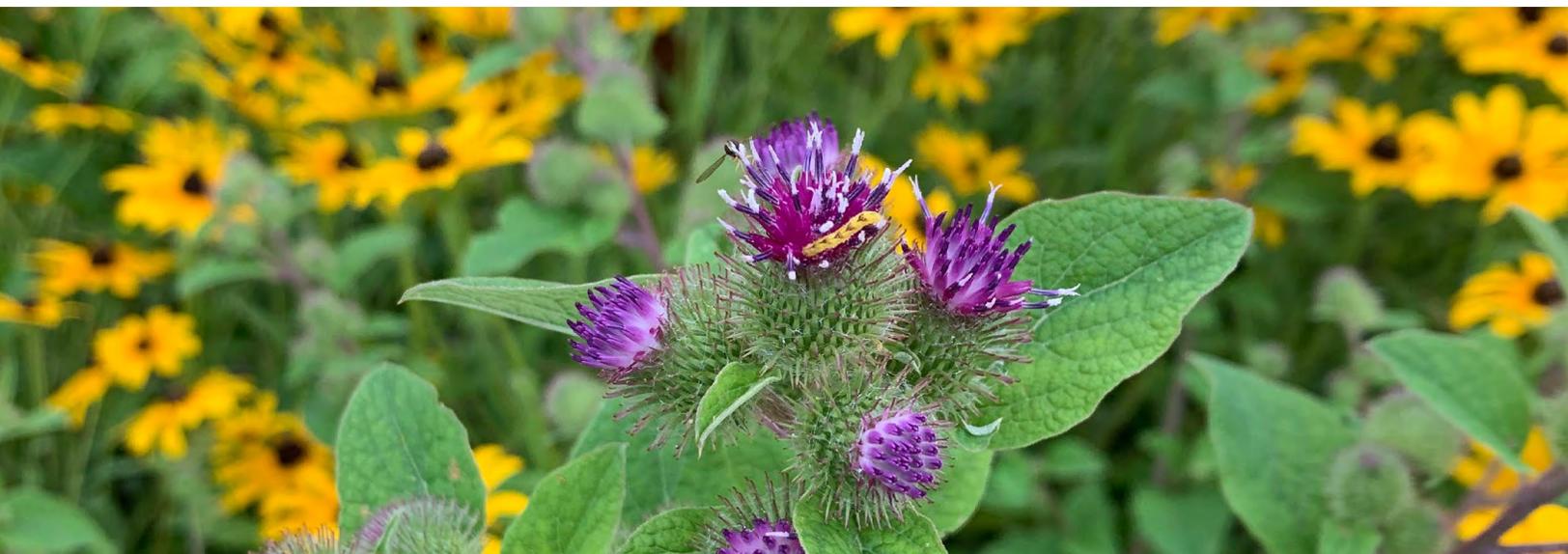
The USEPA registers reduced-risk pesticides through its Conventional Reduced Risk [Pesticide Program](#). These reduced-risk pesticides are commercially viable alternatives to conventional pesticides.

The USEPA characterizes the advantages of reduced-risk pesticides as follows:

- Low impact on human health.
- Lower toxicity to non-target organisms (birds, fish, and plants).
- Low potential for groundwater contamination.
- Low use rates.
- Low pest-resistance potential.
- Compatibility with IPM practices.

Biopesticides, which are derived from such natural materials as animals, plants, bacteria, and certain minerals, are classified separately by the USEPA. For more information on biopesticides, see the USEPA’s [Biopesticide Registration](#) page.

© Bob Searle



Best Management Practices for IPM Control Methods

- Implement proper cultural, irrigation, and turf management practices to reduce stress and pressure of pest establishment.
- Maintain a proper fertilization schedule to improve turf density and quality and reduce pest populations.
- Make sure materials, such as topdressing, are pest-free.
- Apply a preventative pesticide to susceptible turfgrass when unacceptable levels of disease are likely to occur.
- Regularly and systematically monitor for pests.
- Make pest management decisions based on monitoring results and action thresholds.
- Address damage from turfgrass pests such as diseases, insects, nematodes, and animals to prevent density/canopy loss to broadleaf weeds using appropriate combinations of biological, cultural, mechanical/physical and or chemical methods proven to be effective through research.
- When pesticides are used, apply the minimum amount needed, apply only to areas and at times when pests and susceptible life stages are present.
- Follow all label directions for mixing, storing, and applying pesticides. Ensure that applications are done legally and safely using only appropriate and effective products registered in Maine and labelled for the intended use.
- Divert traffic away from areas that are stressed by insects, nematodes, diseases, or weeds.
- When nematode activity is suspected, an assay of soil and turfgrass roots is recommended to determine the extent of the problem.
- If appropriate for the conditions and target species, release insect-parasitic nematodes to naturally suppress insect pests such as white grubs if needed.
- Identify areas on the golf course that can be modified to attract natural predators, provide habitat for them, and protect them from pesticide applications.
- Install flowering plants that can provide parasitoids with nectar or sucking insects (aphids, mealybugs, and soft scales) with a honeydew source.
- Avoid applying pesticides to roughs, driving ranges, or other low-use areas to provide a refuge for beneficial organisms.
- Follow guidelines and advice provided by the FRAC, HRAC, and IRAC as well as local experts such as state Cooperative Extension specialists and the state pesticide regulatory agency.
- Consider selection of reduced-risk pesticides and biopesticides to treat the problem if available and appropriate.

Evaluation and Record Keeping

It is essential to record the results of IPM-related efforts to develop historical information, document patterns of pest activity, and evaluate successes and failures. Records of pesticide use are required for restricted-use pesticides. For IPM purposes, records should be kept for all pesticide applications and should include additional information, such as pest monitoring records, weather records, cultural management logs, and pest response.

Best Management Practices for IPM Evaluation and Record Keeping

- After treatment, determine whether the corrective actions reduced or prevented pest populations, were economical, and minimized risks. Record and use this information when making similar decisions in the future.
- Observe and document turf conditions regularly, noting which pests are present, so that informed decisions can be made regarding the damage the pests are causing and what control strategies are necessary.

© Bob Searle





CHAPTER 8: PESTICIDE MANAGEMENT

Belgrade Lakes Golf Course, Belgrade, ME © Tim O'Writ

PREFACE

The proper use of pesticides is often a necessary part of a turfgrass integrated pest management (IPM) program, to minimize damage to golf course playing surfaces caused by disease, insects, and weeds throughout New England. The term pesticide is inclusive of fungicides, insecticides, growth regulators, and herbicides among others, and is defined by the

U.S. Environmental Protection Agency (USEPA) as any substance or mixture of substances intended to prevent, destroy, repel, or mitigate a pest; or any substance or mixture of substances intended for use as a plant regulator, defoliant, or desiccant.

The use of pesticides can pose a risk to human and environmental health. However, the relative risk of pesticides is largely mitigated through governmental regulation of pesticides and their use, and through responsible decision-making and actions of licensed or certified pesticide applicators. When an application is deemed necessary, pesticide selection should be based on effectiveness, toxicity to non-target species, site characteristics, solubility and persistence in the environment, and to a lesser degree,

cost. This chapter focuses on many of these factors, which golf course superintendents should consider to minimize human and environmental risk associated with pesticide use. These risks are best mitigated by understanding and following the pesticide label directions. Pesticide labels are legal documents that must be followed. If the superintendent does not fully understand the label language, he/she should consult with the local pesticide control agency or the Environmental Protection Agency as well as the pesticide manufacturer.

When an application is deemed necessary, pesticide selection should be based on effectiveness, toxicity to non-target species, site characteristics, solubility and persistence in the environment, and cost.

Human Health Risks

Pesticides belong to numerous chemical classes that vary greatly in their toxicity. Toxicity is a measure of a substance’s potential to cause injury, illness, or death. It is characterized through laboratory studies to determine the dose or concentration of a chemical that results in 50% mortality of an animal test population (i.e., lethal dose or LD50). All pesticide labels contain a “Signal Word” (Table 8-1) to characterize their relative risk to human health based on acute toxicity of six studies of oral, dermal, inhalation, and eye and skin irritation (USEPA, 2014).

Table 8-1. Pesticide signal words

Signal Word	Toxicity Level
Danger	Toxicity Category I
Warning	Toxicity Category II
Caution	Toxicity Category III
None required (or Caution as optional)	Toxicity Category IV

However, the risk to human health associated with pesticide use depends on both pesticide toxicity and the level of exposure. Exposure is related to how much an individual is exposed to a pesticide. Thus, the risk of a very highly toxic pesticide may actually be very low, if the exposure is sufficiently small. To minimize human health risks associated with golf course pesticide use, superintendents should select effective pesticides with lower toxicities, including reduced risk pesticides, and adopt practices that reduce exposure to applicators, staff, and their clientele.

REDUCED RISK PESTICIDES

Special designation is given by the USEPA to pesticides meeting the following criteria: low impact on human health, lower toxicity to non-target organisms (birds, fish, plants), low potential for groundwater contamination, low use rates, low pest resistance potential, and compatibility with IPM (USEPA, 2018). Several reduced risk pesticide active ingredients are labeled for use in turf (Table 8-2) but be sure to check with your state regulatory agency for any restrictions on specific pesticide products and/or changes to products and registration status. Some of these have been demonstrated to reduce hazard to golfers due to the reduced toxicity and use rates (Doherty, 2017).

Table 8-2. Examples of active ingredients found in some reduced risk pesticides labeled for use on golf course turf. Check with your state pesticide regulatory agency for any restrictions on specific pesticide products and/or changes to products and registration status.

Fungicides	Herbicides	Insecticides
boscalid	penoxsulam	clothianidin
penthiopyrad	carfentrazone-ethyl	chlorantraniliprole
trifloxystrobin	mesotrione	cyantraniliprole
fludioxonil	bispyribac-sodium	fipronil
azoxystrobin		spinosad

List as of June 2018. <https://www.epa.gov/pesticide-registration/reduced-risk-and-organophosphate-alternative-decisions-conventional>.

NONAGRICULTURAL USE REQUIREMENTS

The “re-entry” interval (REI) a period of time in which entry into a pesticide-treated area is restricted. Several pesticides used in production agriculture have specifically stated “re-entry intervals” on the label, preventing workers from entering a treated field for the stated period of time after application unless they use the required personal protective equipment (PPE). The purpose of this restriction is to protect farm workers from exposure to residues of pesticides while they work in the fields. Golf courses must follow the Nonagricultural Use Requirements for re-entry into a pesticide-treated area following application. For Nonagricultural Use Requirements, the applicator should follow the individual label instructions for each pesticide used to identify the appropriate re-entry interval following application. Noncertified and licensed applicators must receive training on these standards and procedures prior to use. Often (but not always), a one-hour re-entry period following golf course pesticide applications is appropriate to reduce worker and golfer pesticide exposure (Putnam et al., 2008; Doherty, 2017).

This is based on research conducted at the University of Massachusetts that has demonstrated that pesticide residue transfer from treated turf is greatest within the first hour after application (Figure 8-1 and 8-2) (Putnam et al., 2008; Doherty, 2017). Thus, pesticide exposure to golfers and staff can be reduced by limiting entrance into treated areas for a minimum of one hour following an application. However, applying pesticides at night to provide a re- entry period before the following day does not effectively reduce pesticide transfer, and can actually increase pesticide exposure compared with daylight applications (Putnam et al., 2008).

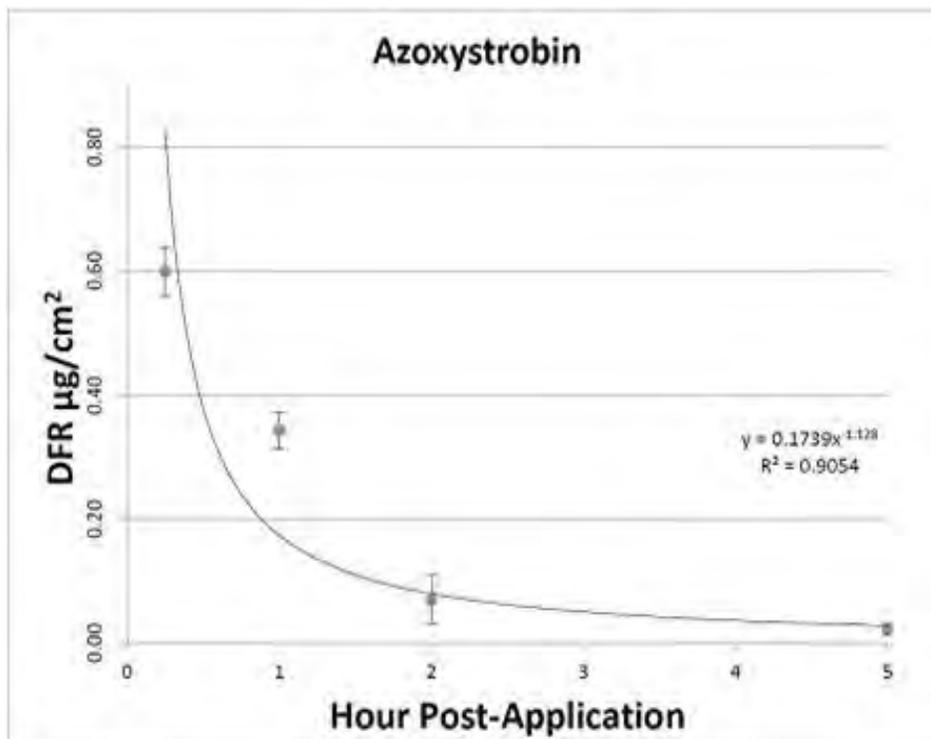


Figure 8-1. Azoxystrobin dislodgeable foliar residues were measured over the first 5 hours following application. Residues of azoxystrobin declined rapidly within the first two hours post application with a 43% reduction of residues 1 hour after application. From Doherty, 2017.

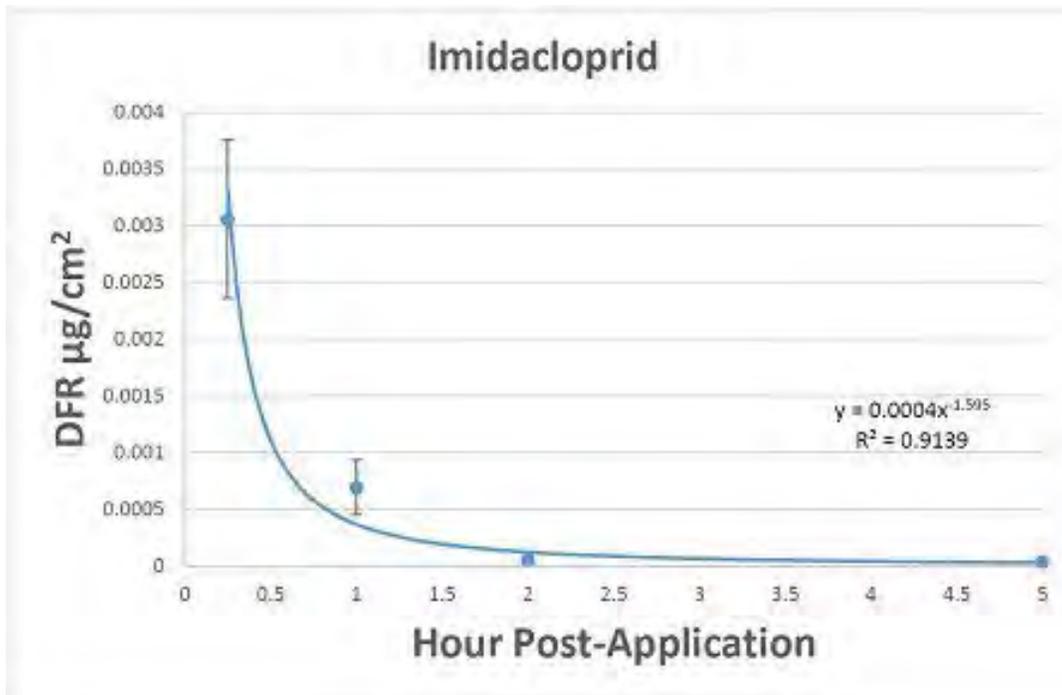


Figure 8-2. Imidacloprid dislodgeable foliar residues were measured over the first 5 hours following application. Residues of imidacloprid declined very rapidly with a 4.3- fold reduction within the first hour after application. From Doherty, 2017.

POST-APPLICATION IRRIGATION

Irrigating pesticide-treated turf immediately following application can move some pesticide residues from the foliage into the lower canopy, or to the thatch and soil. This practice can reduce dislodgeable foliar pesticide residues 9 to 30-fold compared with turf that is not irrigated after pesticide application (Doherty, 2017). Post-application irrigation may be an effective way to reduce pesticide exposure and may help target some turf pests (e.g., grubs, root pathogens, pre-emergent weeds). It can also delay volatilization of chemicals. However, irrigating treated turf can reduce efficacy of some pesticides, particularly contact materials. Always follow all label directions regarding irrigation before or after treatment.

Best Management Practices for Reduce Human Health Risks

- Select the least toxic pesticide with the lowest exposure potential.
- Use Reduced Risk Pesticides when appropriate.
- Restrict staff and golfer entry to pesticide treated areas until spray has dried or dust has settled or for at least one hour following application.
- Irrigate pesticides targeting soilborne pests following application to reduce exposure to foliar residues.
- Know the emergency response procedure and follow all label instructions in case excessive exposure occurs.

Environmental Fate and Transport

Pesticides applied to any environment have the potential to interact with wildlife or migrate into surface and subsurface waters. Environmental implications of a pesticide can often be determined by the environmental hazards statement found on pesticide product labels. The “Environmental Hazards” statement, found under the general heading “Precautionary Statements,” provides language advising the user of the potential hazards to the environment from the use of the product. The environmental hazards generally fall into three categories: general environmental hazards, non-target toxicity, pollinator hazards (look for the bee icon on the label – see right) and endangered species protection.

While pesticides can pose risks to the environment, it is important to recognize that turfgrass systems are particularly well suited to capturing and degrading pesticides due to their high plant density and sub-surface thatch layer (Branham, 2006). In fact, BMPs for minimizing pesticide and nutrient runoff from agricultural fields recommend use of turfgrass between fields and waterways as vegetative filter strips to minimize runoff (Krutz et al., 2005). Pesticides applied to turfgrass systems are often strongly adsorbed



to thatch and soil, limiting their movement (Dell et al., 1994; Lickfeldt and Branham, 1995). Moreover, naturally occurring processes including chemical and microbial degradation and photodegradation dissipate pesticides in the environment (Figure 8-3). Environmental fate and transport of pesticides is largely dependent on their physical and chemical properties (Table 8-3). Consideration of these properties can help guide golf course superintendents' selection of pesticides to reduce environmental impacts.

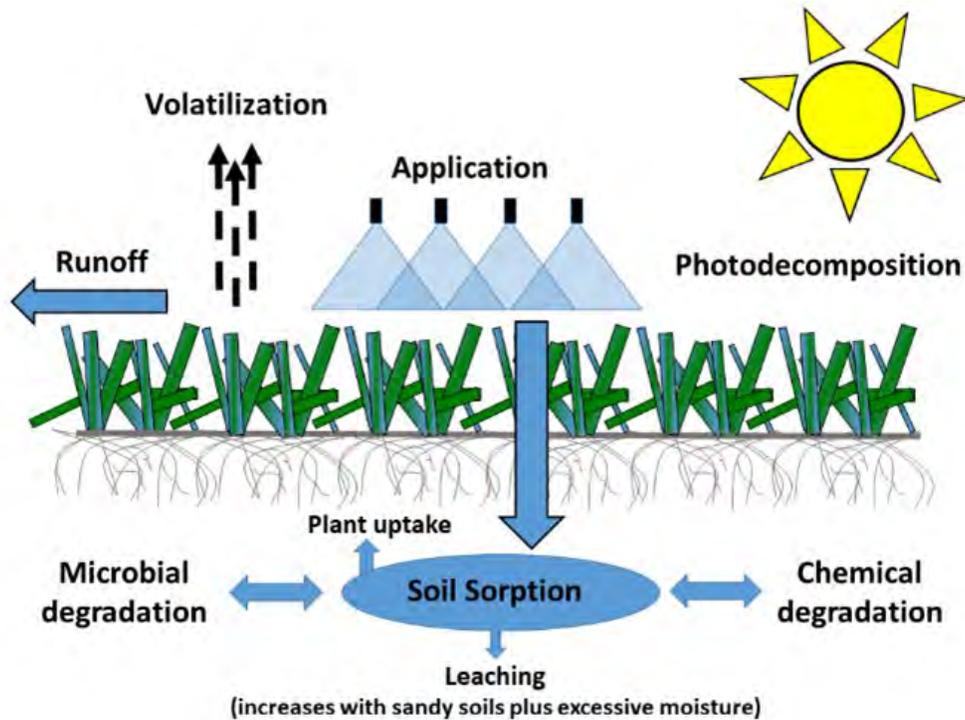


Figure 8-3. Sources of pesticide degradation and loss in the environment. Reproduced from Fundamentals of Turfgrass and Agricultural Chemistry. McCarty et al., 2003.

Table 8-3. Pesticide characteristics associated with increased risk of groundwater and surface water contamination.

Chemical Characteristic	Range for Potential Contamination
Water solubility	greater than 30 ppm (= mg/L)
K_d	less than 5, usually less than 1
K_{oc} (mL/g)	less than 300 to 500
Henry's law constant	less than 10^{-2} atm per m^{-3} mol
Hydrolysis half-life	more than 175 days
Photolysis half-life	more than 7 days
Field dissipation half-life	more than 21 days

PERSISTENCE (HALF-LIFE)

Most pesticide active ingredients are carbon-based molecules that degrade over time. The persistence of any pesticide is characterized by its half-life (DT50 or $t_{1/2}$), or the amount of time required for an initial amount to be reduced by half. The half-life of most pesticides is less than 120 days (McCarty et al., 2003).

Half-life is independent of the amount of pesticide applied, although the rate applied does affect how many half-life intervals are necessary for a pesticide to be completely degraded. Using pesticides with a shorter half-life can reduce their overall persistence in the environment. Additionally, selecting pesticides with lower use rates, or applying products at lower rates, can also reduce persistence, since less pesticide is initially applied.

SOLUBILITY

A chemical's ability to dissolve in water is known as its solubility and is reported as parts per million (ppm), milligrams per liter (mg/L), or grams per liter (g/L). A high solubility denotes a highly soluble chemical. More soluble pesticides have a greater potential of moving in surface or soil water and can be associated with greater risk of runoff or leaching. Pesticides with solubility greater than 30 ppm or mg/L are at increased risk for groundwater and surface water contamination unless the pesticide tightly binds to soils or organic matter (thatch).

ORGANIC CARBON SORPTION COEFFICIENT (KOC)

The affinity for various pesticides to adsorb to organic matter is commonly expressed as Koc (mL/g). Mature turfgrass stands typically contain a layer of organic matter, referred to as thatch, underlying the canopy. Pesticides with a small Koc value do not strongly adhere to thatch and soil organic matter and are therefore more likely to leach through the soil and reach groundwater. Conversely, pesticides with large Koc values tend to remain near the soil surface, reducing the likelihood of leaching, but can be carried to surface water via runoff or soil erosion.

Pesticide Risk Assessment Tools

USEPA is charged with providing risk assessments of all federally registered pesticides on an ongoing basis. When used according to label directions, registered pesticides are expected to have a reasonable certainty of no harm. Despite this, golf courses that may want to further understand and characterize the pesticides they use can consider one or a combination of several available pesticide risk assessment indices.

These tools exist to assess the potential risk of pesticide transport and impact on human and environmental health. They synthesize several characteristics described in the preceding sections to assign a single value to pesticides for comparison.

Superintendents may use these values, in addition to efficacy and cost, to select pesticides that further minimize risks. In all cases, these tools are estimates for how a pesticide may respond in the environment.

GROUNDWATER UBIQUITY SCORE (GUS)

The groundwater ubiquity score is a value used to describe the potential for a pesticide to leach to groundwater based on half-life and Koc plotted on a log-scale (Gustafson, 1989). Pesticides with GUS values greater than 2.8 are considered "leachers," while those with GUS values less than 1.8 are "non-leachers." GUS is a convenient tool to assess and compare pesticide leachability. However, in some

cases it may overstate (e.g., soils with high organic matter) or understate (e.g., coarse soils) the leaching potential since it does not consider site conditions.

WINDOWS PESTICIDE SCREENING TOOL (WIN-PST)

WIN-PST is a free software-based tool developed by the U.S. Department of Agriculture’s Natural Resources Conservation Service that can be used to evaluate the potential movement of pesticides in water or eroded soil/organic matter and to estimate the risk to non-target organisms. It uses soil survey databases with information such as soil type, organic matter content, and water table depth, along with pesticide characteristics, to provide site-specific risk estimates. Users can specify application method, relative application rate (i.e., standard, low, ultra low), and rainfall probability.

Long-term human and fish toxicity data and ratings are also included in WIN-PST. Toxicity ratings can be combined with the off-site movement potential ratings to provide an overall estimate of the potential risks from pesticide movement below the root zone and past the target area.

ENVIRONMENTAL IMPACT QUOTIENT (EIQ)

Environmental impact quotient is a comprehensive pesticide assessment tool that synthesizes risk factors affecting applicator/golf course workers, indirect human hazards, groundwater, and ecological toxicities into a single EIQ value (Kovach et al., 1992). An advantage of EIQ is the ability to compare the potential environmental impact of pesticides based on their formulation and application rates by calculating a “Field Use EIQ” (FUEIQ) (Table 8-4). FUEIQ values can be calculated using the formula below, or determined online using the “[Calculator for Field Use EIQ](#).” New York State golf course BMPs (2014) recommend a desirable single application FUEIQ to be less than 25. A value greater than 100 is considered an increased risk.

$$\text{FUEIQ} = \text{EIQ} \times \text{Rate (lbs/acre)} \times \% \text{ Active Ingredient}$$

Using FUEIQ, it is also possible to quantify the cumulative risk of pesticide management programs by totaling all single application FUEIQs over the course of the season.

FUEIQ can also be used to characterize pesticide risk of a treated area (e.g., fairway in an environmentally sensitive area) by multiplying the FUEIQ of an application by the total treated acreage. One disadvantage of EIQ compared to WIN-PST is that it does not use site-specific data in its risk assessment.

Table 8-4. Comparison of EIQ and FUEIQ of three contact fungicides applied for dollar spot control in fairway turf.

Active ingredient	% Active ingredient	Application rate	EIQ	FUEIQ
chlorothalonil	54.0%	2.0 fl.oz. / 1,000 ft ²	37.4	110.0
iprodione	23.3%	2.0 fl.oz. / 1,000 ft ²	24.2	61.5
fluazinam	40.0%	0.5 fl.oz. / 1,000 ft ²	23.3	12.7

Best Management Practices for Pesticide Risk Assessment

- Know what and where the target pest is and select an efficacious pesticide and application method.
- Understand pesticide sorption principles so that appropriate decisions can be made.
- Avoid using highly water-soluble pesticides when possible.
- Select pesticides that have a low runoff and leaching potential.
- Before applying a pesticide, evaluate the impact of site-specific characteristics (e.g., proximity to surface water, water table, and well-heads; soil type; prevailing wind) and pesticide-specific characteristics (e.g., half-lives and Koc).
- Identify label restrictions that may pertain to your facility (e.g., additional restrictions for sites with sandy soils).
- Select pesticides with reduced impact on pollinators (see “Pollinator Protection” chapter).
- Select pesticides that, when applied according to the label, have no known effect on endangered species present on the facility.

Pesticide Storage

Storage and handling of pesticides in their concentrated form poses the greatest potential risk to ground or surface waters. For this reason, facilities that store and handle these products must be properly sited, designed, constructed, and operated. Storage facilities should facilitate the secure, dry storage of pesticides; provide safe working conditions for personnel with easy access to PPE; and provide secondary containment of incidental spills due to normal mixing/loading practices and secondary containment of large accidental spills. The following suggestions in this document are offered for consideration, as well more detailed information in [Storing, Mixing and Loading of Pesticides](#) from the Massachusetts Department of Agricultural Resources (MDAR).

STORAGE LOCATION

Storage areas should be located to minimize risk to human health and the environment associated with potential spills, contaminated runoff, or fire. The location should be easily accessible to service vehicles in case of an emergency. Pesticide storage facilities should be at least 400 feet downhill from drinking water supplies and 200 feet from surface water. They should not be placed within a 100-year floodplain, and storm runoff should be diverted around them. It should be a detached structure separate from other spaces where employees or clientele are allowed.

ENGINEERING CONTROLS

Walls and doors: Storage buildings should be built to contain and resist potential fire. Fire rating of walls influences suggested building setbacks (Table 8-5.) Interior walls should be impervious to pesticides (e.g., painted steel, aluminum, fiberglass). Doors should be lockable, steel (solid core), and set in a steel frame that opens to the outside.

Floors and concrete specifications: Concrete floors with impervious sealant or comparable surface should be used for pesticide storage facilities. Type I or Type II cement is suggested. Epoxy, urethane, polyester, vinyl, chlorosulfonated polyethylene, and polyurea coatings prevent corrosion of floors due to fertilizers and pesticides. Coating efficacy varies and should be selected based on types of products stored in the facility. A continuous sill should surround the floor to contain 125% of the volume of the largest container in storage.

Lights and ventilation: Storage facilities should include enough light to clearly read pesticide labels. A ventilation system should be installed to dissipate potential chemical vapor and ensure a safe workspace. Fans should be wired to turn on with lights and displace six air changes per hour. All electronics should be explosion proof, especially if any flammables or volatile materials are stored there.

Table 8-5. Fire rating and suggested building setback for various wall fireproofing materials. Adapted from: Pesticide Storage Mixing and Loading Guidelines for Applicators. MDAR.

Fire Rating	Gypsum Wallboard [†]	Hollow Masonry	Solid Masonry	Solid Concrete	Building Setback
1-hour wall	1 layer	3 inch	4 inch	3 inch	50 feet
2-hour wall	2 layer	4 inch	6 inch	4 inch	25 feet
4-hour wall		6 inch	10 inch	6 inch	none

†Gypsum wallboard of 5/8 inch thickness on both sides of wall constitutes a 1-hour rated firewall. Two wallboards on both sides of wall are required for a 2-hour rated firewall.

STORAGE CONDITIONS

Pesticides should be stored in their original container with the label clearly visible. Pesticides within the storage facility should not be exposed to direct sunlight, freezing temperatures, or extreme heat. Flammable materials should be stored in fireproof containment. Separate the fungicides, insecticides, and herbicides within the storage area to prevent unintended usage. Dry pesticides should be stored separately from liquid formulations to prevent contamination in case of leakage. Place pesticide containers within chemical-resistant bins or on shelves with a raised lip to contain leaks. Food, feed, potable water, seed, and personal protective equipment should not be stored within pesticide storage areas.

Near the storage area there should be easy access to PPE, eyewash station, first aid kit, with antidotes if appropriate, and a spill kit.

Best Management Practices for Pesticide Storage

- Store, mix, and load pesticides away from sites that directly link to surface water or groundwater.
- Whenever possible, store pesticides in a lockable concrete or metal building that is separate from other buildings.
- Locate pesticide storage facilities away from other types of structure to allow fire department access.
- Storage facility floors should be impervious and sealed with a chemical-resistant paint.
- Floors should have a continuous sill to retain spilled materials and no drains, although a sump may be included.
- Sloped ramps should be provided at the entrance to allow the use of wheeled handcarts for moving material in and out of the storage area safely.
- Shelving should be made of sturdy plastic or reinforced metal.
- Metal shelving should be kept painted to avoid corrosion. Wood shelving should never be used because it may absorb spilled pesticides.
- Automatic exhaust fans and an emergency wash area should be provided.
- Light and fan switches should be located outside the building, so that both can be turned on before employees enter the building and can be turned off after they leave the building.
- Avoid temperatures less than 40°F or greater than 100°F inside the pesticide storage facility.
- Personal protective equipment should be easily accessible and stored immediately outside the pesticide storage area.
- Place a spill containment kit in the storage area, in the mix/load area, and on the spray rig.

Willowdale Golf Club, Scarborough, ME © Tim O'Wrl



Pesticide Inventory

Pesticides degrade over time. Do not store large quantities of pesticides for long periods. Adopt the “first in–first out” principle, using the oldest products first to ensure that the product shelf life does not expire. Avoid storing pesticides more than two years old and make sure temperatures do not exceed 100°F or drop below 40°F at any time. Utilize computer software systems to record inventory and use. Safety Data Sheets (SDS) and specimen labels for all pesticides on hand should be kept in an easily identifiable location, outside the pesticide storage facility.

Best Management Practices for Pesticide Inventories

- Avoid purchasing large quantities of pesticides that require storage for greater than six months.
- Adopt the “first in–first out” principle, using the oldest products first to ensure that the product shelf life does not expire.
- Ensure original labels are on every package and container.
- Consult inventory when planning and before making purchases.
- Control temperature to avoid extreme heat or cold.

Pesticide Mixing/Washing

Pesticide leaks or spills, if contained, will not percolate down through the soil into groundwater or run off the surface to contaminate streams, ditches, ponds, and other waterbodies. One of the best containment methods is the use of a properly designed and constructed chemical mixing center (CMC). These chemically impervious areas prevent seepage of pesticides into soil and facilitate easier clean-up and containment of spills or overflows. Suggestions for CMC building specifications are noted below, and are available in [Storing, Mixing and Loading of Pesticides](#) published by MDAR.

MIXING PAD

Chemical mixing centers should be located at similar distances from drinking water sources and surface waters as pesticide storage facilities, and preferably adjacent to them. An impervious, sealed concrete pad should be constructed to accommodate the sprayer size/weight and tolerate winter freeze/thaw cycles. Edges of the pad should be curbed to contain spills and discharges.

CONTAINMENT VOLUME

The CMC should be built large enough to contain a volume 1.25 times the size of the largest spray tank loaded on the pad. Preferably, the area would have a roof to protect the containment area from precipitation. A greenhouse frame covered with a three-year co-polymer film can be a low cost alternative to a roof. In the absence of a roof, the containment volume should be increased to

accommodate a two-year, 24-hour storm event (2.9 to 3.6 inches of rain).

SUMP DESIGN

The pad should slope to a sump to collect all spills, rinsate, and discharges. Chemically impervious materials should be used for the sump. Minimize dirt, clippings, rocks, and other debris from entering the containment pad and sump. Keep the sump area clean of solids. A pump should be installed to transfer the liquid collected in the sump to a holding tank for use in subsequent tank filling or applied as a pesticide to appropriate turf areas. (Consult with Maine DEP to ensure legality of transfer method.) Must make sure any pesticide material is applied at or below the maximum label rate allowed on the site treated with the rinsate.

PESTICIDE MIXING AND HANDLING

Handling open pesticide containers, measuring pesticide materials, or working with pesticide application equipment presents an exposure risk to the handlers and the environment. Applicators and handlers should put on label-required PPE prior to opening pesticide packages. All pesticide handling must be done by a trained employee and restricted to the appropriate storage area or CMC.

Best Management Practices for Pesticide Mixing/Washing Stations

- Loading pesticides and mixing them with water or oil diluents should be done over an impermeable surface, so that spills can be collected and managed.
- The mixing station surface should offer easy cleaning and the recovery of spilled materials.
- Pump the sump dry and clean it at the end of each day.
- Liquids and sediments should be removed from the sump and the pad whenever pesticide materials are changed to an incompatible product (i.e., one that cannot be legally applied to the same site).
- Apply liquids and sediments from the sump as you would a pesticide, strictly following label instructions.
- Absorbents such as cat litter or sand may be used to clean up small spills and then applied as a topdressing in accordance with the label rates or disposed of as a hazardous waste.
- Sweep up solid materials and use as intended.
- Collect wash water (from both inside and outside the application equipment) and use it as a pesticide in accordance with the label instructions.
- The rinsate may be applied as a pesticide (preferred) or stored for use as makeup water for the next compatible application.

Personal Protective Equipment

Based on exposure, pesticide handlers and applicators are at the greatest risk for potential adverse health effects. Exposure to pesticides can be mitigated by practicing good work habits and adopting modern pesticide mix/load equipment (e.g., closed- loading) that reduce potential exposure. PPE statements on pesticide labels provide the applicator with important information about protection. Fit testing is required for many respirators.

Best Management Practices for Personal Protective Equipment

- Provide all required PPE to all employees who work with pesticides (including equipment technicians who service pesticide application equipment).
- Have a written policy that employees must wear all required PPE during mixing/loading and application or when working on contaminated equipment.
- Ensure that PPE is sized appropriately for each person using it.
- Make certain that PPE is appropriate for the chemicals used.
- Ensure that PPE meets rigorous testing standards and is not just the least expensive.
- Store PPE where it is easily accessible but not in the pesticide storage area.
- Forbid employees who apply pesticides from wearing facility uniforms home where they may come into contact with children.
- Provide laundering facilities or uniform service for employee uniforms.
- The federal Occupational Safety and Health Administration (OSHA) requires employers to fit-test workers annually who must wear many respirators.
- Meet requirements for [OSHA 1910.134 Respiratory Protection Program](#).

Pesticide Container Management

The containers of some commonly used pesticides are classified as hazardous waste if not properly rinsed, and as such, are subject to the many rules and regulations governing hazardous waste. The improper disposal of a hazardous waste can result in high fines and/or criminal penalties. However, pesticide containers that have been properly rinsed can be handled and disposed of as nonhazardous solid waste. Federal law (Federal Insecticide, Fungicide, and Rodenticide Act, or FIFRA) and some state laws require pesticide applicators to rinse all empty pesticide containers before taking other container disposal steps. Under federal law (Resource Conservation and Recovery Act, or RCRA), a pesticide container is not empty until it has been properly rinsed.

Best Management Practices for Pesticide Container Management

- Rinse pesticide containers immediately in order to remove the most residue.
- Rinse containers during the mixing and loading process and add rinsate water to the finished spray mix.
- Rinse emptied pesticide containers by either triple rinsing or pressure rinsing.
- Puncture empty, rinsed pesticide containers and dispose of them according to the label.

Emergency Preparedness and Spill Response

Accidents can happen. Advance preparation on what to do when an accident occurs is essential to mitigate the human health effects and the impact on the environment. A spill containment kit containing absorbent materials (e.g., reusable gelling agents, cat litter, clay, soil, or sand), garbage can, and a shovel should be available for small spills.

Hydrated lime or bleach can be used to neutralize and clean surfaces where spills occur. Spill containment kits should be easily accessible within the pesticide storage area. Ensure that PPE, a first-aid kit, and eye-wash stations or eye-wash bottles are accessible outside the pesticide storage and mixing area.

An emergency response plan containing actions to take and personnel to contact in the event of a spill or accident should be in place. The plan should include the following information:

- Names and quantities of pesticides in inventory.
- Location of property, including a map and directions (to relay over phone in emergency).
- Names, addresses, and phone numbers of the designated spokesperson, superintendent, and key employees.
- Plan of facility showing pesticide storage locations, flammable materials, electrical service, water supply, fuel storage tanks, fertilizer storage, fire hydrants, storm drains, and nearby wetlands, ponds, or streams.
- Location of emergency equipment supplies.
- Contact information for fire, police, hospital, pesticide regulatory agency (DEP, DEC, DEM, etc.), spill clean-up firm, board of health, and facility owner.

Ensure that copies of the plan are located near the pesticide storage facility and the office and distributed to local police and fire departments. Maintain copies in English and any other language commonly used by employees. Be sure to update the information regularly for local police and fire departments.

Best Management Practices for Emergency Preparedness and Spill Response

- Develop a golf course facility emergency response plan that includes procedures to control, contain, collect, and store spilled materials.
- An inventory of the pesticides kept in the storage building and the SDS and specimen labels for the chemicals used in the operation should be accessible on the premises, but not kept in the pesticide storage room itself.
- Prominently post “Important Telephone Numbers” including CHEMTREC, for emergency information on hazards or actions to take in the event of a spill.
- Ensure an adequately sized spill containment kit is readily available.
- Designate a spokesperson who will speak on behalf of the facility should an emergency occur.
- Host a tour for local emergency response teams (e.g., firefighters) to show them the facilities and to discuss the emergency response plan. Seek advice on ways to improve the plan.

Sprayer Calibration

Properly calibrated application equipment is paramount to mitigating environmental and human health concerns and to effectively manage the pest. Sprayer output is dependent on several variables (e.g., speed, nozzle size, pressure). Spray coverage is often reduced at greater application speeds, regardless of nozzle size. To maximize efficacy of pesticide applications, applicators should consider optimization of spray coverage versus efficiency of labor when choosing spray speeds.

Best Management Practices for Sprayer Calibration

- Ensure that the spray technician is experienced, licensed (or certified), and properly trained.
- Minimize off-target movement of pesticides by using properly configured application equipment.
- Properly calibrate all application equipment at the beginning of each season (at a minimum) and after equipment modifications and document the calibration in your record book.
- Check equipment daily when in use.
- Use recommended spray volumes for the targeted pest to maximize efficacy.
- Calibration of walk-behind applicators should be conducted for each person making the application to take into consideration their walking speed and other variables.

Sprayers and Nozzles

Various types and sizes of application equipment are readily available. The size of the equipment (tank size, boom width, etc.) should match the scale of the target area.

Larger ride-on sprayers are more efficient for large areas, while small walk-behind boom sprayers are well suited for smaller areas. Smaller boom lengths may increase the accuracy of applications, minimizing overspray on non-target areas. Individual nozzle control on global positioning system (GPS) assisted boom sprayers can further minimize overspray of non-target areas and has resulted in 25% less pesticide applied at some golf courses (USGA Green Section, 2016).

Spray nozzle size and design affect the spray drop size. This can be an important factor influencing the potential for drift and off-target movement of pesticides. Smaller droplet sizes can improve the efficacy of some pesticides, although they are more susceptible to drift. Larger droplets are more resistant to drift, although may reduce the efficacy of some pesticides due to reduced coverage. Nozzles designed to encapsulate an air bubble within the droplet (e.g., air induction nozzles) provide a good compromise between drift reduction (larger droplet size) and coverage and efficacy (drop bursts into small drops on impact). Additionally, nozzles designed with a wider spray angle (i.e., 110° versus 80°) enable booms to be set lower to the ground where they are less susceptible to drift.

Best Management Practices for Sprayers and Nozzles

- Use an appropriately sized applicator for the size of area being treated.
- Equipment too large in size requires greater volumes to prime the system. This can result in significant waste that must be properly handled.
- Use wide-angle, air-induction, flat-fan nozzles to minimize spray drift to non-target areas.
- Wind speed should be between 2 – 10 miles per hour or as required by the label. The maximum allowable wind speed is 15 mph as required by the Maine Board of Pesticides Control.
- Understand how to recognize a temperature inversion and never apply under inversion conditions.

Pesticide Record Keeping

Maintaining accurate records of pesticide-related activities (for example, purchases, storage, inventory, applications, spills, etc.) is essential and required by most state pesticide regulatory agencies.

Best Management Practices for Pesticide Record Keeping

- Keep and maintain records of all pesticides used to meet legal (federal, state, and local) reporting requirements.
- Use records to monitor pest control efforts and to plan future management actions.
- Use electronic or hard-copy forms and software tools to properly track pesticide inventory and use.
- Keep a backup set of records in a safe but separate storage area.

The Ledges Golf Club, York, ME © Forrest Bell





CHAPTER 9: POLLINATORS

Riverside Golf Course, Portland, ME © FBE

PREFACE

Most flowering plants need pollination to reproduce and grow fruit. While some plants are pollinated by wind, many require assistance from insects and other animals. In the absence of pollinators, many plant species, including many of the fruits and vegetables we eat, would fail to survive. In fact, 35% of the fruits and vegetables that make up our diet require pollination by honey bees and other insect pollinators.

The western honey bee (*Apis mellifera*), a very important pollinator in the United States, is maintained in commercial and residential bee hives all around the country. In Maine, there are approximately 1,200 registered beekeepers who maintain just over 10,000 hives. Recent controversies have arisen regarding honey bee health. Some people believe honey bee hives are struggling and that improper pesticide applications are to blame for their decline. Others believe that honey bee colonies are performing at levels similar to the past 20 or 30 years, and that many factors are contributing to any decline in honey bee colony health. The truth is probably somewhere in the middle. Meanwhile, many other insect pollinators, including many bumble bee species, solitary bees, flower flies, and butterflies play a critical role in our food supply.

One thing that everyone agrees on is that honey bee colonies face many stresses, one of which is the use

of insecticides in pollinator foraging areas. Other factors that can impact honey bee health are:

Riverside Golf Course, Portland, ME © FBE

- Stress from a migratory lifestyle. Many hives are moved from one crop to another every five to eight weeks over the course of several months. Hives can have problems adapting to a new habitats and the limited nature of their diet can also cause problems.
- *Varroa destructor* mites, which invade colonies, introduce viruses into a colony and compromise the overall health of a hive.
- Presence of American foulbrood, a fatal disease caused by *Paenibacillus larvae*.
- Stress-related pathogens, including *Nosema species* and European Foulbrood (*Melissococcus plutonius*).
- Habitat loss, including both forage and nesting sites
- Climate change which disrupts bee/forage synchronicity, causes extreme weather events and reduces the nutritional content of available forage.
- Exposure to insecticides, especially when they are applied when bees are foraging.
- Winter mortality (e.g., drastic fluctuations in temperatures or desiccating winds).



For more on pollinator basics, see the University of Maryland webpage [Pollinator Basics](#).

Pesticides and Pollinators

The purpose of this section is to discuss methods to minimize any possible harmful impact of pesticides (especially insecticides) on honey bees and other insect pollinators. Pesticides are products designed to control pests (e.g., insects, diseases, weeds, or nematodes). Pesticides and other plant health products, including plant growth regulators, surfactants, and biostimulants, are often used in golf course management.

The unintended non-target effects of products used in golf course management are of increasing concern. Therefore, pesticide applicators – and the people making the decisions about pesticide

applications – must be mindful of the impact that pesticides can have on pollinator species and their habitats.

Pollinator-protection language is found on labels of all pesticides that can be harmful to pollinators. Some insecticides that are particularly toxic to honey bees have a bright yellow, diamond-shaped box (“bee box”) outlining restrictions on applications (Figure 9- 1). Several classes of insecticides, including neonicotinoids and pyrethroids, are known to be toxic to honey bees and other pollinators. Toxicity data for several turf insecticides commonly used in the Northeast are summarized in Table 9-1. The data for bees is given in micrograms per bee. Note that several insecticides (including all the neonicotinoids, most of the pyrethroids, chlorpyrifos, and spinosyn) have a honey bee LD50 of 0.01 microgram per bee or less. This means that each of these products has the potential to be harmful to honey bees, and therefore every possible step must be taken to minimize the likelihood of exposure of bees to the product. Scientific understanding of pesticide toxicity to honey bees is evolving quickly, particularly as declines in honey bee populations are observed. New research suggests that there are possible synergies between insecticides and other applied chemicals (particularly fungicides) that can be more harmful to honey bees when applied in tandem (Sgolastra et al. 2016, Thompson et al. 2014). These studies underscore the importance of minimizing the likelihood of pesticide exposure to pollinators.

THE NEW EPA BEE ADVISORY BOX
On EPA's new and strengthened pesticide label to protect pollinators

PROTECTION OF POLLINATORS

APPLICATION RESTRICTIONS EXIST FOR THIS PRODUCT BECAUSE OF RISK TO BEES AND OTHER INSECT POLLINATORS. FOLLOW APPLICATION RESTRICTIONS FOUND IN THE DIRECTIONS FOR USE TO PROTECT POLLINATORS.

Look for the bee hazard icon in the Directions for Use for each application site for specific use restrictions and instructions to protect bees and other insect pollinators.

This product can kill bees and other insect pollinators.
Bees and other insect pollinators will forage on plants when they flower, shed pollen, or produce nectar.

Bees and other insect pollinators can be exposed to this pesticide from:

- Direct contact during foliar applications, or contact with residues on plant surfaces after foliar applications.
- Ingestion of residues in nectar and pollen when the pesticide is applied as a stem treatment, soil, tree injection, as well as foliar applications.

When Using This Product Take Steps To:

- Minimize exposure of this product to bees and other insect pollinators when they are foraging on pollinator attractive plants and the application site.
- Minimize drift of this product on to beehives on to off-site pollinator attractive habitat. Drift of this product onto beehives can result in bee kills.

Information on protecting bees and other insect pollinators may be found at the Pesticide Environmental Stewardship website at:
<http://pesticideenvironmentalstewardship.org/pollinatorprotection/Pages/default.aspx>

Pesticide incidents (for example, bee kills) should immediately be reported to the state/local lead agency. For contact information for your state/tribe, go to www.epa.gov. Pesticide incidents can also be reported to the National Pesticide Information Center at: www.npi.orst.edu or directly to EPA at: beekill@epa.gov

Alerts users to separate restrictions on the label. These prohibit certain pesticide use when bees are present.

The new bee icon helps signal the pesticide's potential hazard to bees.

Makes clear that pesticide products can kill bees and pollinators.

Bees are often present and foraging when plants and trees flower. EPA's new label makes it clear that pesticides cannot be applied until all petals have fallen.

Warns users that direct contact and ingestion could harm pollinators. EPA is working with beekeepers, growers, pesticide companies, and others to advance pesticide management practices.

Highlights the importance of avoiding drift. Sometimes, wind can cause pesticides to drift to new areas and can cause bee kills.

The science says that there are many causes for a decline in pollinator health, including pesticide exposure. EPA's new label will help protect pollinators.

EPA

Read EPA's new and strengthened label requirements: <http://go.usa.gov/jHH4>

Figure 9-1. The USEPA Bee Advisory Box can be found on new insecticide labels that may pose a risk to pollinators.

Note that no correlation exists between mammalian toxicity and bee toxicity. While several insecticides have very low toxicity to mammals (e.g., dinotefuran, thiamethoxam, or spinosad), each of these products is highly toxic to honey bees. Also, note the considerable variation in honey bee toxicity within some chemical classes. For example, chlorantraniliprole (an anthranilic diamide) is virtually non-toxic to honey bees, while cyantraniliprole is moderately toxic to bees.

Recordkeeping is required by law in Maine (and most other New England states) in order to use most pesticides. IPM principles suggest that you keep records of all pest control activity so that you can refer to previous outbreaks to determine what worked or did not work, and to select the best course of action in the future.

Table 9-1. Ecotoxicology of several common turf insecticides in several different animal species (Richmond and Patton, 2014).

Insecticide class	Insecticide	Mammal LD₅₀ mg kg⁻¹	Bird LD₅₀ mg kg⁻¹	Fish LC₅₀ mg L⁻¹	bee LD₅₀ µg bee⁻¹
Neonicotinoid	chlothianidin	>500	430	104	0.004
	dinotefuran	>2,000	>2,000	>100	0.023
	imidacloprid	424	152	211	0.0037
	thiamethoxam	>1,563	576	>125	0.005
Pyrethroid	beta-cyfluthrin	77	>2,000	0.000068	0.001
	bifenthrin	54	1,800	0.00026	0.1
	lambda-cyhalothrin	56	>3,950	0.00021	0.038
Organophosphate	chlorpyrifos	64	13	0.0013	0.059
	trichlorfon	212	37	0.7	>0.4
Carbamate	carbaryl	614	>2,000	2.6	0.14
Avermectin	emamectin benzoate	24	23	0.174	0.0035
Spinosyn	Spinosad	>5,000	>2,250	2.69	0.024
Anthranilic diamide	chlorantraniliprole	>5,000	>2,250	>12	>4
	Cyantraniliprole	>5,000	>2,250	>12.6	>0.1

Pesticide Application Practices to Protect Pollinators

Pesticide applicators must make careful decisions about what materials to use and when to apply them to minimize the impacts of pesticides on bees, other pollinators, and beneficial arthropods. While integrated pest management BMPs address decision-making considerations for determining whether to use a pesticide to manage a pest (and which pesticide would be least disruptive to bees and other pollinators), countless other factors must also be considered when deciding on a course of action.

Superintendents must determine what is best for a specific facility based on local agronomics (e.g., turf species and cultivars, fertility practices), local geographic conditions (e.g., underlying soils, slopes, surface water, groundwater), cost of materials and labor, availability of application equipment, expectations of the membership, and budget.

Turf entomologists collaborated to create the national BMPs for protecting pollinators (Larson et al., 2017) listed below that can be implemented at golf facilities to protect pollinators. These BMPs provide guidance for decision-making, but the superintendent and golfers – must recognize that there often are challenging circumstances that limit a superintendent’s options.

© Bob Searle



Best Management Practices for Pesticide Application Practices

- Before applying a pesticide, scout the area for pest and beneficial insect populations (including pollinators), and only apply a pesticide when a pest damage threshold has been reached. Pest thresholds vary from one part of a golf course to another; for example, little or no damage is typically acceptable on putting greens, while golfers and turf managers are more tolerant of pest activity in roughs.
- Use non-pesticide pest management approaches (e.g., lures, pheromones, cultural manipulations) to manage pest insect populations when possible.
- When pesticides must be used to manage a pest insect population, select one with a lower impact on pollinators.
- Several classes of insecticides are known to be toxic to honey bees, including neonicotinoids, pyrethroids, carbamates, and spinosyns.
- Chlorantraniliprole has no activity against bees, ants, or wasps.
- Granular formulations generally reduce pollinator exposure, compared with a sprayable formulation of the same active ingredient.
- Apply water after the application to move the residue off the surface and reduce exposure.
- Avoid applying pesticides when plants (including flowering weeds) are in bloom or bees are foraging.
- If there are blooming plants in an area that needs to be treated, mow the area immediately before application to remove blossoms from flowering weeds or use herbicides to reduce weed populations (Larson et al., 2013).
- Remember that systemic insecticides (e.g., neonicotinoids) may be absorbed through the roots of nearby ornamental plants and translocated to flowers, so be very careful to avoid applying pesticides in a way that results in translocation of the active ingredient to flowers of nearby ornamental trees and shrubs.
- Whenever possible, schedule pesticide applications early in the morning or late in the evening, when fewer pollinators are foraging.
- Avoid applications during unusually low temperatures or when dew is forecast.
- Use spray technology (e.g., drift-reduction nozzles, larger droplet sizes) to reduce off-site drift of a pesticide.
- Refer to Chapter 8 “Pesticide Management” for details on properly disposing of unused pesticides and pesticide containers.

Pollinator-Related Communication

In addition, several steps can be taken to protect pollinators by increasing communication and interaction with people in the community. The State of Maine Board of Pesticides Control has a formal pesticide notification rule that provides citizens (i.e. neighbors) with the right to know when a pesticide is being applied in their vicinity. Citizens can receive notifications through two avenues: (1) a full Request for Notification which requires the citizen to make a request for their neighbor (within 500 feet or 1,000-feet for aerial applications) to notify them and supply information regarding pesticide applications, and (2) participation in the Maine Pesticide Notification Registry, which is a more formal notification that requires an annual fee and assures notification through a formal process. More details can be found on the Maine Board of Pesticides Control's webpage, listed in the footnote of this page¹.

Recommendations include the following:

Bee boxes at Willowdale Golf Club, Scarborough, ME. © Rick Lewis

- Consider joining a local beekeeper association to become connected with local educational events and mentoring opportunities.
- Consider setting up a couple of hives within a natural area of the golf course, to be maintained by an interested staff member or possibly a nearby beekeeper seeking another suitable location.
- Check with state or local agricultural and natural resource agencies and obtain a list of registered beekeepers within a three-mile radius of the golf course. Let them know of pending applications that could affect honeybees. This advance notice enables beekeepers to temporarily close hive entrances, keeping bees inside until pesticides have dried on the foliage. In Maine, contact the State Apiarist found at the Maine State Beekeepers Association webpage².
- Attend workshops or online seminars to learn more about pollinators and other beneficial arthropods Organizations in Maine offering workshops and seminars related to pollinators include the Maine Board of Pesticides Control, the Maine Landscape & Nursery Association, and the Maine Beekeepers Association.
- Use signage, social media and local news media to educate golfers and the general public about the steps taken to minimize impact on pollinators.
- Invite local master gardeners, garden clubs, or student groups to visit the golf course or host workshops at local garden centers to demonstrate how you use the information on a pesticide label to minimize impact on pollinators.



¹ <https://www.maine.gov/dacf/php/pesticides/public/notification.shtml>

² <http://mainebeekeepers.org/beekeeping-resources/maine-bee-inspector/>

Enhancing Pollinator Habitat

One of the biggest challenges pollinators face is the loss of natural habitat, as suburban areas encroach into more rural settings. Habitat necessary to support pollinators includes both plants for foraging and nesting sites. One way to encourage pollinator activity on golf courses is to provide pollinator habitat in non-play areas. General considerations for pollinator-friendly plantings include the following:

- Utilize native plants or well-behaved non-native plants
- Maintain a minimum of three plants blooming, continuous throughout season
- Plant a diversity of plants to support a variety of pollinators. When choosing plants vary the following:
 - Flower color: Bees prefer blue, purple, white and yellow flowers while butterflies generally prefer red.
 - Flower shape: Bees prefer bell shaped flowers while beetle and fly pollinators prefer open plate shaped flowers.
 - Plant height: some pollinators will forage up at the tops of trees while others will forage less than a foot off the ground.

Creating pollinator-friendly gardens and native grass or tall meadow areas on the property can provide opportunities for superintendents to reach out to the community. Organizing field trips for primary school-aged children can provide a tremendous opportunity to educate the community about the ecological and environmental benefits of golf courses.

Some pesticide manufacturers have developed pollinator programs and provide flower seed blends that have been developed for different regions of the United States. These include [Operation Pollinator\(Syngenta\)](#), [Bee Care \(Bayer\)](#), [Living Acres Monarch Challenge](#), and [Monarchs in the Rough](#). The Natural Resources Conservation Service (NRCS), U.S. Fish and Wildlife Service, and Xerces Society have partnered to create the New England Pollinator Partnership. This organization is working together towards a 25-year conservation effort and agreement to provide financial and technical assistance towards conservation efforts to participating landowners in New England. Participation in this program covers the landowner if their actions "...inadvertently harm or kill a threatened or endangered bumble bee during farming operations.". Contact an NRCS service center for more details³. Many environmental organizations provide resources to identify good native plants to enhance pollinator health, as described in the "Landscaping" chapter. Other simple steps for providing nesting sites for native species can include leaving stems, coarse, woody debris, and exposed patches of sand or well- drained soil in out-of-play areas. Many of the solitary nesting bees and wasps' nest in sand hazard areas. These solitary nesters pose little risk to golfers and is a great opportunity for public education. Signage in the club house as well as near the spots where this occurs can alleviate any fears and also provide an opportunity to showcase pollinator friendly activities. In addition, nesting boxes or hollow bamboo sticks can be provided for solitary nesting species. For more information, visit the Maine State Apiary webpage (<https://www.maine.gov/dacf/php/apiary/index.shtml>) and the Maine Extension service fact sheet on bees and pesticides (<https://extension.umaine.edu/publications/7153e/>).

³ <https://www.nrcs.usda.gov/wps/portal/nrcs/detailfull/me/technical/ecoscience/threat/?cid=nrcseprd1575616>

Best Management Practices to Enhance Pollinator Habitat

- Plant a diversity of flowering pollinator-friendly plants when renovating out-of-play areas.
- Leave nesting materials and sites in out-of-play areas whenever possible. Mow natural or pasture areas just once per year – late in the season when plants are going dormant – to control woody plants or other growth at a time that minimizes effect on pollinators. If mowing during the season is necessary, mow high to protect bumble bee nesting sites which tend to be at the base of bunched grasses.
- Consider providing man-made nesting sites for solitary nesting species.

Val Halla Golf Club, Cumberland, ME © Toby Young



Making Way for Pollinators

A case study from Riverside Municipal Golf Course

Pollinators face challenges related to loss of natural habitat. On golf courses, one way to encourage pollinator activity is to provide pollinator habitat in non-play areas with a diversity of wildflower species for a variety of food sources. Riverside Golf Course has designated pollinator areas and pollinator gardens.

In addition, Riverside Golf Course superintendent partnered with the Honey Exchange in 2017 and keeps domestic bee colonies on the golf course.

Riverside Golf Course also practices reduced pesticide best management practices and integrated pest management to protect pollinators.



© Riverside Golf Course

Photos: Bee hive on the golf course (top), bees on sunflower (left), and designated natural area for pollinators (right).



© Toby Young



© Riverside Golf Course



© Riverside Golf Course

Riverside Golf Course has designated natural areas with bird nesting boxes and bee hives in out of play areas on the course.

Nest boxes provide a place for birds to build their nests and be protected from the elements and predators. Birds provide many benefits to golf courses, including pest control, flower pollination, and wildlife viewing opportunities.

In addition, Riverside Golf Course tracks bird species present on their course, egg numbers in their nest boxes, and successful fledglings.

Nest boxes at Riverside primarily attract Eastern Bluebirds, tree swallows, chickadees, and wrens. Photos below show nest box monitoring and bird banding to track behavior.

Improving Habitat for **Birds** on the Course



© Riverside Golf Course



© Al Hardy



© Riverside Golf Course

Photos: Birdhouses installed on the course (top), eggs inside nest box on the course (left), female tree swallow banding at Salmon Falls Country Club (middle), and nest box monitoring (right).





CHAPTER 10: MAINTENANCE OPERATIONS

Val Halla Golf Club, Cumberland, ME © Toby Young

PREFACE

Facilities related to the storage and handling of pesticides, fertilizers, fuel and petroleum, and other chemicals, especially in their concentrated form, pose the highest potential risk to water sources if accidentally released in quantity. Therefore, anyone storing, mixing, or loading potentially hazardous chemicals should treat all leaks, spills, and fires as emergencies and be prepared to respond to these emergencies promptly and correctly. For unintended releases of any chemicals, an emergency response plan, spill kit, and first-aid kit should be readily available.

The “Pesticide Management” chapter includes storage- and handling-related BMPs specifically for pesticides. This chapter provides additional guidance for maintenance operations and points out differences between managing fertilizer equipment and pesticide equipment.

Storage and Handling of Fertilizers

Storage facilities that are well designed and well maintained protect people from exposure, reduce the potential of environmental contamination, protect chemicals from extreme temperatures and excess moisture, and reduce liability concerns and potential environmental risks. The storage area should be secure and provide secondary containment features. Secondary containment refers to a means of surrounding primary storage containers with a containment barrier or bin that will control any spills.

Best Management Practices for Storage and Handling of Fertilizers

- Contact the Maine DEP to review groundwater sensitivity information with state hydrologists before constructing any fertilizer storage facilities or handling areas.
- Storage facilities should not be located in areas prone to flooding.
- Locate storage buildings and secondary containment away from wells, water supplies, or surface water runoff area.
- Construct storage buildings upon concrete slabs to prevent seepage or spillage of fertilizer.
- Unless stored in a totally enclosed building, all non-liquid fertilizer materials should be covered and stored within an appropriate secondary containment storage structure.
- Construct liquid fertilizer secondary containment capable of holding 125 percent of the volume of the largest container plus the volume displaced by all the other containers inside the liquid containment area.
- Construct secondary containment dry storage of sufficient thickness and strength to withstand loading conditions, such as a concrete slab.
- Design loading areas to prevent spillage onto unprotected areas by installing secondary containment.
- This can be achieved with perimeter curbing.
- Post warning signs on fertilizer storage buildings, especially near entry or exit areas.
- Storage facilities should be secured and allow access only to authorized staff.
- Replace worn or faulty valves, plugs, and threaded fittings in storage containers.
- Install a backflow prevention device on water supply lines used for fertilizer or pesticide mixing or equipment rinsing so as not to contaminate the line.
- Lock valves and shutoff devices while storage containers and facilities are not in use.
- Follow hazard safety rules, worker protection laws, and fire prevention rules while handling and storing fertilizer.

Best Management Practices for Storage and Handling of Fertilizers (continued)

- Apply appropriate sealant to seams and cracks in all storage facilities and load/wash/rinse pad areas.
- Use approved containers designed for and compatible with the fertilizer being stored.
- Shelves should be made of plastic or reinforced metal. Metal shelving should be coated with paint to avoid corrosion. Wood shelving should not be used due to its ability to absorb spilled chemicals.
- Exhaust fans and an emergency wash station should be provided.
- Light and fan switches should be located on the exterior of the storage facility. Any electrical inside the storage building should be explosion proof.
- Store liquid materials separate and below dry materials to prevent contamination from a leak.
- Train staff and other management on how to access and use the facility's SDS database.
- Maintain accurate inventory lists and Safety Data Sheets on-site, including information on whom to contact in the event of a spill or release to the environment.

Val Halla Golf Club, Cumberland, ME © Toby Young



Equipment Storage and Maintenance

Equipment storage and maintenance facilities should be designed to prevent the accidental discharge of chemicals, fuels, or contaminated wash water from reaching water sources. Properly storing and maintaining equipment also extends the useful life of machines and reduce repairs.

Best Management Practices for Equipment Storage and Maintenance

- Store and maintain equipment in a covered area complete with a sealed impervious surface to limit risk of fluid leaks contaminating the environment and to facilitate the early detection of small leaks before causing significant damage to the turf or the environment.
- Seal floor drains unless they are connected to a holding tank or sanitary sewer.
- Store pesticide and fertilizer application equipment in areas protected from rainfall. Rain can wash pesticide and fertilizer residues from the exterior of the equipment and possibly contaminate soil or water.
- Store solvents and degreasers in lockable metal cabinets away from ignition sources in a well-ventilated area. These products are generally toxic and highly flammable. Never store them with fertilizers or in areas where smoking is permitted.
- Keep an inventory of solvents and Safety Data Sheets (SDS) for those materials on-site but in a different location where they will be easily accessible in case of an emergency.
- Keep basins of solvent baths covered to reduce emissions of volatile organic compounds.
- When possible, replace solvent baths with recirculating aqueous washing units. Soap and water or other aqueous cleaners are often as effective as solvent-based products and present a lower risk to the environment.
- Always use appropriate personal protective equipment (PPE) when working with solvents.
- Never allow solvents or degreasers to drain onto pavement or soil, or to discharge into waterbodies, wetlands, storm drains, sewers, or septic systems.
- Collect used solvents and degreasers in containers clearly marked with contents and date. Schedule collection by a commercial service.
- Blow off all mowing equipment with compressed air to reduce damage to hydraulic seals.

Equipment Washing

Equipment washing should be conducted under controlled conditions in an appropriate area with secondary containment to prevent adverse wash water runoff impacts whenever to the environment. Equipment washing guidelines and restrictions should be established that reduce the potential for pollutants to reach surface water or groundwater.

Proper cleaning of equipment helps prevent residues from reaching surface waters, groundwater, drainage pipes, or storm drains. The residues from washing equipment include grass clippings, soil, soap, oil, fertilizer, and pesticide.

A primary concern when washing mowing equipment is the nitrogen and phosphorus nutrients in grass clippings. Using compressed air to blow clippings off mowers before washing can help reduce the amount of nutrients that enter drains via wash water. The best practice is to have a dedicated wash area to collect grass clippings. Clippings can be collected, then composted or removed to a designated debris area. When formal washing areas are not available, a “dog leash” system using a short, portable hose to wash off the grass at random locations, away from surface waters, wells, or storm drains, is an option.

For equipment with possible pesticide residue, BMPs should be followed to ensure that wash water does not become a pollution source. Captured wash water can be used as a dilute pesticide per label, or it may be pumped into a rinsate storage tank for use in the next application and used as a dilute pesticide per the label.

Washpads to blow debris of equipment (right) and wash equipment (left) with wash water draining into rain garden. © Bob Searle



Best Management Practices for Equipment Washing

- Brush or blow off accumulated grass clippings from equipment using compressed air.
- Wash equipment on a concrete pad or asphalt pad that collects the water. Do not wash equipment in the rain and/or ensure that washing occurs under a roofed area. After the collected material dries, collect and dispose of it properly.
- Washing areas for equipment not contaminated with pesticide residues should drain into oil/water separators before draining into sanitary sewers or holding tanks.
- Do not wash pesticide-application equipment on pads with oil/water separators. Do not wash near wells, surface water, or storm drains.
- Use spring-loaded spray nozzles to reduce water usage during washing.
- Minimize the use of detergents. Use only biodegradable, non-phosphate detergents.
- Do not reuse containment wash water for irrigation.
- Do not discharge wash water to surface water, groundwater, or susceptible/leachable soils either directly or indirectly through ditches, storm drains, or canals.
- Never discharge to a sanitary sewer system without written approval from the appropriate entity.
- Never discharge to a septic tank.
- Do not wash equipment on a pesticide mixing and loading pad. This keeps grass clippings and other debris from becoming contaminated with pesticides.
- Solvents and degreasers should be used over a collection basin or pad that collects all used material.

Fueling Facilities

Fueling areas should be properly sited, designed, constructed, maintained, and monitored to prevent petroleum products from being released into the environment through spills or leaks. Fueling facilities may be regulated either locally or through the state depending on size and material(s) stored. Check with local Code Enforcement to ensure proper installation and operation.

An aboveground storage tank (AST) is easier to monitor for leakage and is therefore the preferred storage method. Depending on the amount of petroleum product stored, a Spill Prevention Control and Countermeasure Plan may be needed. Because of the potential for groundwater contamination from a leaking underground storage tank (UST), leak detection monitoring is a critical aspect of UST compliance. Any leaks or spills must be contained and cleaned immediately and reported to the appropriate response authority.

Fueling areas should be sited on impervious surfaces, equipped with spill containment and recovery

facilities, and located away from surface waters and water wells. Spill clean-up equipment such as booms and absorbents should be stored on-site. Catch basins in fueling areas should be directed toward an oil/water separator or sump to prevent petroleum from moving outside any containment structure. Floor drains in fueling areas should be eliminated unless they drain to containment pits or storage tanks.

Best Management Practices for Fueling Facilities

- Locate fueling stations on concrete or impervious surfaces under roof structures whenever possible.
- Develop a record-keeping process to monitor and detect leakage in USTs and ASTs.
- Regularly inspect ASTs visually for leakage and structural integrity.
- Secure the fuel storage facilities and allow access only to authorized and properly trained staff.
- Ensure that fuel tanks and pumps are properly labeled.
- Post “No Smoking” signs near fueling facilities.
- Protect tanks and fueling areas with strategically placed concrete bollards or other devices.

Val Halla Golf Club, Cumberland, ME © Toby Young



Waste Handling

Facilities need to regularly review how they handle the disposal of unwanted, expired, or accumulated items, including chemicals, paints, pesticides, tires, batteries, used oils, solvents, paper products, plastic or glass containers, fluorescent light tubes, and aluminum cans. Developing recycling programs reduces waste and minimizes the quantity of waste reaching landfills. In some cases, recycling of some wastes may be required locally, and superintendents should be aware of these requirements.

All packaging from chemicals, their containers, and other wastes should be properly disposed of. Pesticide-specific waste handling requirements are identified on the pesticide label and are discussed in more detail in the “Pesticide Management” chapter.

Best Management Practices for Waste Handling

- Label containers for collecting used solvents, oils, and degreasers.
- Recycle lead-acid batteries. If not recycled, batteries are classified as hazardous waste.
- Store old batteries on impervious surfaces in areas protected from rainfall and run-on.
- Recycle used tires, paper products, plastic or glass containers, aluminum cans, and used solvents, oils, and degreasers.
- Provide a secure and specifically designated storage area for the collection of recyclable waste products.
- Recycle or properly dispose of light bulbs and fluorescent tubes.

© Samoset Golf Club, Rockport, ME.



Emergency Preparedness and Spill Response

As discussed in the “Pesticide Management” chapter, enough absorbent material must be available to control a spill of the largest container in storage areas. Sorbent materials include booms, socks or mini booms, pillows, pads and rolls, and loose sorbents. These sorbent materials may be universal or more specific (such as for petroleum products).

Having a readily accessible spill kit is a necessity at any facility where chemicals are used or stored. The spill kit should contain, at a minimum, the following:

- Proper clothing and Personal Protection Equipment (PPE).
- A supply of neutral absorbing materials that may include activated charcoal, clay, or vermiculite.
- Clean water source.
- Class B fire extinguisher for chemical fires.
- Detergent that can be used for deactivation of spill site.
- Salvage drum with lid for waste cleanup.
- Cleanup tools such as brooms, shovels, and dust pans.

Best Management Practices for Spill Response

- Develop a golf course facility emergency response plan that includes procedures to control, contain, collect, and store spilled materials.
- Prominently post “Important Telephone Numbers,” including the hotline number for emergency information on hazards and response actions in the event of a spill.
- Ensure an adequately sized spill containment kit with response equipment such as booms and sorbent materials are readily available.
- Designate a spokesperson who will speak on behalf of the facility should an emergency occur.



CHAPTER 11: SUSTAINABLE LANDSCAPING IN OUT OF PLAY AREAS

Sable Oaks Golf Course, South Portland, ME. © Forrest Bell

PREFACE

During play, golf competitors often become immersed in their landscaped surroundings. While care of tees, greens, and fairways will always be the highest priority for golf course superintendents, out-of-play areas are also an important component of superintendents' responsibilities. Landscaped and "non-play" areas help delineate in-play areas and contribute to the overall beauty of the golf course design. They enhance course aesthetics, provide wildlife habitat, and add a natural buffer that moderates external noise. Maintaining these aesthetically pleasing areas as sustainably as possible is economically advantageous and supports the biodiversity of pollinators and other wildlife. Developing or expanding naturalized areas may reduce dependence on water, chemical, and fuel inputs, while allowing more intensive maintenance to be reserved for areas dedicated to play (Lyman et al., 2007; Gross and Eckenrode, 2012).

The substantial acreage of golf course properties provides an ideal opportunity for environmental stewardship and conservation. Vegetated areas with a greater diversity of plant species support wildlife by providing forage and habitat for pollinators and other beneficial insects (Tallamy, 2009). Less intensively managed vegetation (e.g., tall grass and naturalized areas) directly correlates with a

higher biodiversity of plants and pollinators (Colding and Folke, 2009; Dobbs and Potter, 2013). Critical to this effort are native plants, which are important in the restoration of local ecosystems and form the foundation of the food web. Golf courses can contribute to plant and pollinator diversity by expanding natural habitat and restoring native plants throughout the course – in the rough and other out-of-play areas, as well as in high-visibility areas, such as the property surrounding the clubhouse and other outbuildings.

Benefits of Sustainable Areas on the Golf Course

An ecosystem with a healthy variety of plants fosters a robust biodiversity of animal and insect species. Plants provide a primary food source and habitat, yield nutrients, improve soil health, and produce oxygen. Golf courses can provide a critical link that connects wildlife corridors by increasing naturally vegetated habitat, including unmown grass and native wildflower out-of-play areas. Benefits of increasing the sustainability of out-of-play areas include:

- Attracting beneficial wildlife, supporting pollinator habitat, enhancing biodiversity, and creating aesthetic interest that provides year-round visual pleasure for golfers on the course (Figure 11-1).
- Providing an option for out-of-play areas that requires fewer non-renewable inputs (fertilizer, water, and gasoline) to maintain.
- Protecting soils, natural vegetative cover, water resources, and water quality.
- Greater carbon sequestration potential than high maintenance areas, due to increased plant biomass production (Wissman, 2016).

The University of Maine Cooperative Extension defines **native plants** (also called indigenous plants” as ‘... those that either originated here or arrived without human intervention.” Native plants play a critical role in the landscape, providing food and habitat for animals and functions such as shading for other plant species.

© Bob Searle.



Figure 11-1. Tall grass areas add to the beauty of golf course landscapes.

Sustainable Landscaping Concepts

What is sustainable landscaping? According to the American Society of Landscape Architects, “sustainable landscapes sequester carbon, clean the air and water, increase energy efficiency, restore habitats, and create value through significant economic, social, and environmental benefits.”

Dense, healthy turfgrass plays an important function in sequestering carbon. However, frequent maintenance practices contribute to carbon emission, which diminish the carbon sequestering benefits of turfgrasses. The leading cause of increased carbon dioxide (CO₂) emissions is the direct result of fossil fuel use (Gillette et al., 2011). By reducing frequently mown acreage and expanding sustainable areas that are managed with fewer inputs, golf courses can potentially be valuable net carbon sinks (natural systems that absorb and store carbon dioxide from the atmosphere). Naturalized areas can offset the higher carbon demands of intensively managed, priority areas.

When designing a sustainable landscape, plants are selected for much more than simple aesthetic value. Plants should be selected because they are already adapted to the existing soil conditions, available water, and the microclimate, so additional inputs of irrigation, fertilizer, and soil amendments can be reduced or eliminated. Native plants are prioritized because they have evolved in concert with native wildlife and pollinators, providing the foundation of local food webs that enable butterflies, birds, and other wildlife to survive. Most herbivorous insects and pollinators are specialists that cannot survive on introduced or exotic plant species.

The population of many beneficial insects and pollinators has declined due to a variety of factors, including loss of natural habitat, lack of forage opportunities, diseases, predatory insect infestations, stress, and exposure to pesticides (Mader et al. 2011).

Native plants in the landscape contribute to the restoration of local ecosystems and create conditions that support a wide variety of indigenous, beneficial animal and insect species. Therefore, in naturalized or less intensively managed areas of the landscape, where tolerance of potential pest damage is higher, native plants should be prioritized to support pollinators, food chains, and native ecosystems.

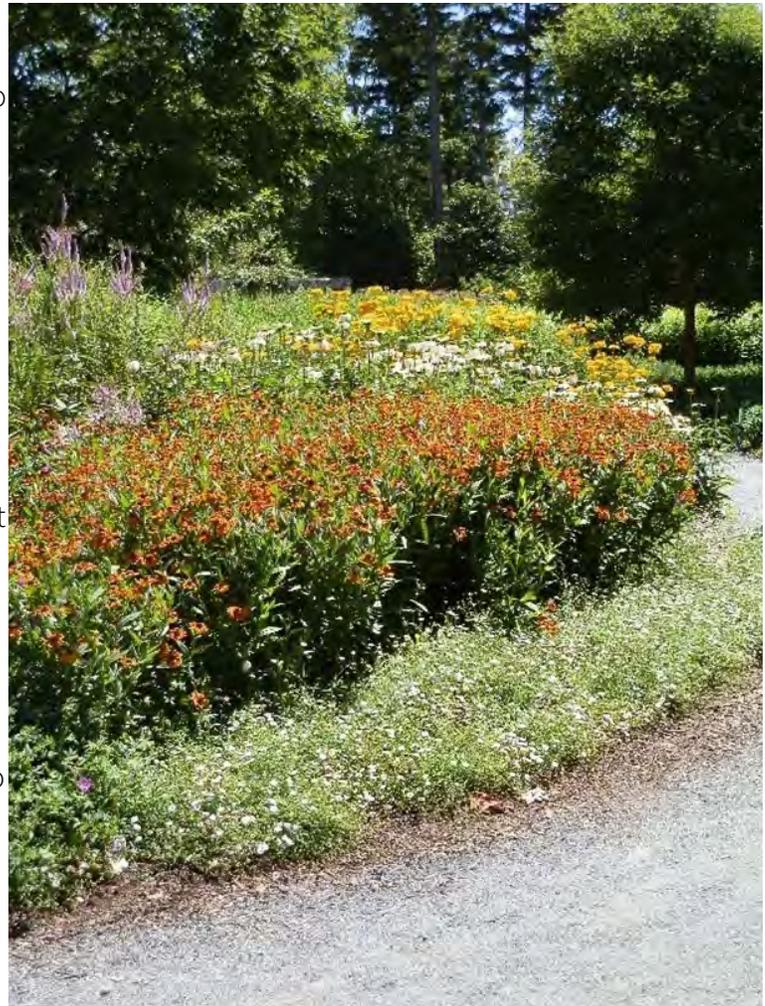


Figure 11-2. A traditional garden design that incorporates native plants. Photo taken from New England Regional Manual.

Two distinct approaches to sustainable landscape design are as follows:

Traditional design, which uses native plants as an alternative for introduced or exotic ornamental species in a formal garden, often including mulched landscape beds and lawn areas (Figure 11-2). Required maintenance is the same as any typical garden area, with possibly reduced irrigation if drought tolerant plants are used. This type of design is best suited for priority areas of high visibility around the clubhouse or other out-buildings that provide aesthetic focal points.

Naturalized design, which uses maturing and evolving native plant communities, such as tall grass, meadow, and forested areas. This style is a more viable and cost-effective option in the long term for large out-of-play areas. Required maintenance is consistent with meadows and periphery areas.

Facility managers seeking to conserve water and protect ecosystems on the course can employ a type of sustainable landscaping known as Green Infrastructure (GI). GI is effective and economical and improves the safety and quality of life (USEPA, 2017) through the intentional use of the ecosystem services provided by plants in the managed landscape. GI conserves, restores, or replicates the natural water cycle by reducing and treating stormwater runoff, thus turning a potential pollutant into an environmental and economic benefit. Green roofs, rain gardens, bioswales, cisterns, and permeable pavements are examples of GI. Learn more about how to incorporate GI into the golf course in the fact sheet “Sustainable Landscaping in Out-of-Play Areas on Golf Courses.”

© Bob Searle



Sustainable High Visibility Areas

Landscape design on the golf course should meet the needs of the membership, protect the course's environmental resources, and remain economically sustainable. It is important that high visibility areas of the golf course (around the clubhouse and other out-buildings) are aesthetically pleasing focal points that enhance the overall course aesthetics (Figure 11-3). The landscaping around buildings makes a lasting impression on club members and guests, which can influence club membership and bookings for club functions and golf outings, as well as support playability of the course. Sustainable landscaping concepts can be incorporated into these landscaped areas to fulfill both the course's environmental commitment and its aesthetic goals in high-priority areas.



Figure 11-3 . Native perennials used in a traditional, formal border planting. Photo taken from New England Regional Manual.

Sound design includes the selection of site-appropriate plant cultivars that permit reduced maintenance to remain healthy and attractive. National Turf Evaluation Program, Alliance for Low Input Sustainable Turf, and Turfgrass Water Conservation Alliance can provide information on improved cultivars of turfgrasses for non-golf areas with improved drought tolerance and pest resistance. Whenever new construction or renovation occurs, landscaped areas should be amended to include more native plant material. Native plants are best adapted to the local soils, site conditions, and provide native pest resistance while also supporting the foundation of the food web. For example, caterpillars and insects are critical for birds to raise their young, and native plants provide habitat for native caterpillars and

insects. Incorporating native plants supports a reduced maintenance program that requires less time and expense to maintain. Establishing strong, healthy plants is key for weed management in sustainable landscapes.

Any changes to the landscape design should blend in with existing site features. Changes should be made slowly and with significant investment in membership buy-in. Signage and educational messaging can help members understand how sustainable changes in management decisions supports a healthy ecosystem that in-turn, benefits the course. For example, many native plants grow more slowly than the introduced exotic plants common to contemporary landscapes and the presence of insects and caterpillars indicates a healthy food web foundation for birds and other wildlife. Once established, these communities make golf course landscapes more resilient to nuisance pests and animals. Therefore, communicating plans and progress to the membership will improve membership response to these changes.

Best Management Practices for Landscaping High-Visibility Areas

- Integrate low-maintenance turf and native plant species into landscape areas around the clubhouse, service buildings, and other out-of-play areas and design these areas for ease of maintenance.
- Consider the soil characteristics, climate, sun exposure, water conditions, and pest possibilities when selecting plants. Select plant material not regularly browsed by deer. (Refer to fact sheet “Sustainable Landscaping in Out-of-Play Areas on Golf Courses” for more information.)
- Group plants with similar watering, pH, and fertilizer requirements together to allow for the most efficient use of resources.
- Select each plant based on its unique contribution to the overall project, including blooming schedules, bark, fruit, texture, and habit.
- Utilize native plants wherever possible, integrating them into the landscape along with annuals to maintain season-long color and aesthetic interest.
- Use plants that will perform well over time. Ensure that the mature height and spread of each plant is considered in the landscape design to avoid the need for excessive pruning or regular replacement. For an immediately full, dense landscape, plant annuals or perennial species that can be relocated or divided as plants mature. Quick-growing, short-lived “filler” plants may also help to temporarily embellish a planting bed as it develops.
- Where feasible, design with drought-tolerant and low-water-use plants.
- Use native drought-tolerant plants around buildings, parking areas, or other appropriate places.
- Where irrigation is necessary, utilize high-efficiency irrigation systems (e.g., drip irrigation) in all landscaped areas for maximum efficiency. If possible, design recycling water features, such as collecting rainwater for graywater use.
- Perform a soil test and analysis when analyzing problems or when renovating landscapes.

Best Management Practices for Landscaping High-Visibility Areas (continued)

- Use organic amendments (e.g., compost, compost tea, or leaf mulch), calibrated as part of the overall nutrient management plan, to build healthy soils, establish beneficial soil organisms, and release nutrients over the long term.
- Where necessary, maintain 2-4 inches of organic mulch over the surface of soil, applied a few inches from the base of trees and plants, to keep soil moist and minimize weeds.
- Minimize the use of impervious surfaces and increase permeable features.
- Replenish groundwater by adding rain gardens, green roofs, bioswales, and other permeable surfaces. Install gravel pathways or borders that permit water infiltration, but have low evaporation potential.
- Where possible, recycle, reuse, or use locally sourced materials for plants and hardscapes.

Sustainable Naturalized Areas

Sustainable naturalized areas can help golf superintendents' meet their goals to improve both environmental protection and economic sustainability. While tees, greens, and fairways normally require sufficient irrigation for overall plant health (which contributes to visual impact and recovery from traffic), not all areas of a golf course must be maintained as intensely. In particular, facilities that experience increased seasonal water limitations may consider design alterations to reduce the expanse of maintained turfgrass areas.

Using alternative landscape features in selected areas that do not naturally impact the game of golf can result in substantive water, nutrient, labor, and other maintenance cost savings. They can also restore habitat and increase biodiversity. Conventional landscapes use less than 15 species in an average landscaped lot, while the average undisturbed forest or meadow can support 100 species in the same area. In addition, diverse, multi-storied plantings store more carbon than mown turf areas (Selhort, 2012).

A change in design must be carefully implemented in a manner that does not interfere with game play. While developing a plan to improve and expand wildlife habitat, existing native habitats should be protected and existing natural amenities expanded or enhanced. Retain or restore existing native vegetation, where possible. Where appropriate, existing vegetation should be enhanced through the supplemental planting of native species around tee complexes, out-of-play areas, and water sources.

Construct or modify any existing storage ponds with shallow margins (vegetated buffers) planted with native wetland vegetation utilized by many wildlife species. Nuisance, invasive, and exotic plants should be removed and replaced with native species adapted to the site.

Best Management Practices for Sustainable Naturalized Areas

- Manage natural areas to encourage wildlife diversity, by increasing habitat for locally threatened or endangered species, and to provide habitat connectivity by linking natural areas.
- Actively manage open-space areas to support native habitats and avoid introduction and establishment of invasive species.
- Utilize a diverse range of species in plant selection.
- Ensure that the areas selected for low-maintenance modification will not significantly impact the pace of the golf game or game satisfaction. Use of GPS data trackers will correctly identify placement of these landscapes by identifying and delineating areas that are not in play for golfers.
- Use unmown turf as a landscape element to help focus the design of formal areas.
- Allow beneficial “weeds” (e.g., milkweed, which supports the survival of monarch butterflies) to grow and mature in out of the way areas where they will not interfere with integral in-play areas.

Habitat Corridors

Golf courses can make a positive and significant impact on wildlife diversity by creating new habitat corridors, which provide a safe haven for many species while simultaneously enhancing the golfing experience. Corridors are areas of habitat that physically connect plant and animal populations that cannot maintain healthy, genetically diverse populations when highly fragmented by human activities or structures (UC-Davis, 2008).

© Bob Searle



Best Management Practices for Habitat Corridors

- Establish wildlife corridors that connect areas of habitat, enabling animals to travel and forage for food.
- Corridors should be made as wide as possible and located away from roads, trails, and cart paths, to minimize human interactions with wildlife.
- When establishing or renovating out-of-play areas, include a diversity of pollinator-friendly plants in areas where they will not interfere with routine play.
- Remove invasive exotic plants and replace them with native species adapted to the particular ecological conditions prevalent at the site.
- Protect existing ponds and streams by increasing surrounding vegetative cover height (including turfgrass).
- Incorporate well-adapted, drought-resistant plants, including low-growing ground covers, shrubs, and trees that require little, if any, supplemental irrigation once established.
- Identify and preserve the habitat requirements (food, water, cover, space) for local wildlife species. Preserve existing areas of critical habitat. Retain riparian buffers along waterways to protect water quality and provide food, nesting sites, and cover for wildlife.
- Preserve available nesting materials and sites in out-of-play areas when possible.
- Construct and place birdhouses, bat houses, nesting boxes, or bee houses in out-of-play areas; leave dead tree snags, coarse woody debris, and exposed patches of sand or well-drained soil for nesting and feeding sites, provided they pose no danger to people or property.
- Maintain clearance between the ground and the lowest portion of a fence or wall to allow wildlife to pass, except in areas where animals need to be excluded (BMP-Maryland, 2017).

Meadows/Tall Grass Areas

Replacing supplemental areas of turf with native vegetation, such as in meadows or tall grass areas, provides essential habitat for many species threatened by encroaching development. A meadow is an area of natural grasses and/or native wildflowers that, over time, becomes self-sustaining. Native meadow plants are resilient, accustomed to the regional climate and can survive adverse conditions. Meadow plants have adapted to the existing soil conditions, water availability, and microclimate challenges.

Remediated areas improved by human input, through changes to irrigation, fertilizer, and soil amendments, can successfully be reduced or eliminated, over time.

Meadows that are successfully incorporated into landscape management programs can reduce the burden of some property maintenance expenses. Meadows incorporated into golf environments must be maintained as low-growing and thinly vegetated to give players the ability to easily locate and retrieve

balls that have made their way out of the fairway.

Proper site selection, plant selection, site preparation, and maintenance are critical to designing, establishing, and sustaining a flourishing, beautiful meadow (Figure 11-4). Refer to fact sheet “Sustainable Landscaping in Out-of-Play Areas on Golf Courses” for a list of recommended meadow plants. Most meadow plants prefer full sun. A substantial portion (about 40%) of a meadow should be comprised of grasses (Zimmerman, 2010), in order to sufficiently proliferate between perennial forbs to prevent weed seed germination and development. Time spent on site preparation that eliminates competing vegetation leads to fewer weeds in the meadow in subsequent years. Soil surface disturbance during site preparation should be minimized whenever possible, to prevent unnecessary weed germination at the soil surface. Less disturbance to the site will also maintain soil structure and integrity.

As part of the overall meadow establishment protocol, an effective maintenance plan should be developed before planting and implemented at planting for the successful longevity of the meadow. The initial three years of meadow establishment require both patience and focused effort. During establishment, a nurse crop such as a quick-establishing, clump-forming grass can be used to reduce weed invasion, hold the seed or young plants in place, and protect the soil from erosion.

In the first growing season, perennial meadow plants grow slowly, with an average overall height of 2-6”, depending on the species. Annual weeds will proliferate and grow quickly if given the opportunity. Regular mowing and spot treating weeds can prevent weeds from growing too tall and outcompeting the desired perennials. In the second and subsequent years, the meadow should be mowed annually in late winter or early spring, before the next year’s growth begins.

Best Management Practices for Meadows/Tall Grass Areas

- Select an area for a meadow that receives no less than half a day of direct sunlight to ensure success with sun-loving plants.
- For multi-year health of the meadow, include both short-term species (nurse grasses, annuals, and biennials) and long-term perennial species that take multiple years to establish.
- Select grasses to comprise a substantial portion of the plants to populate the meadow.
- Prepare the site by removing competing vegetation, while avoiding unnecessary disturbance to the soil to maintain soil structure and integrity.
- Where meadows are begun by seed in bare soil areas, utilize an annual “nurse” crop in the first year to aid in establishment.
- Mow every 4 to 6 weeks to a height of 4-6” during the first growing season to control weeds, along with spot treating weeds as needed.
- Mow established meadows annually either in late winter or early spring before the next year’s growth begin.



Figure 11-4. A successfully established meadow with thriving grasses and perennials. Photo taken from New England Regional Manual.

Plant Selection

The fundamental principle for the environmentally sound management of landscapes is “right plant, right place.” Proper plant selection is the most important step in designing a sustainable landscape planting. Use a detailed, completed site analysis as described in fact sheet “Sustainable Landscaping in Out-of-Play Areas on Golf Courses ” to select appropriate plants for the site and ensure successful establishment based on sunlight requirements, soil conditions, and water availability. Native plants should be an integral component of the landscape design throughout the course and at key focal points.

Native plants are best adapted to the local soils, site conditions, and pests. The goal of species selection in a sustainable landscape is to maintain as close to a natural ecosystem as practical. Whenever possible, 50-70% of the non-play areas should remain in natural cover (Gross and Eckenrode, 2012). As much natural vegetation as possible should be retained and enhanced through the supplemental planting of native trees, shrubs, and herbaceous vegetation to provide active wildlife habitat. Over time, incorporating native plants not only enhances the ecosystem function of a landscape by supporting a wide variety of indigenous, beneficial animal and insect species, it can also reduce the time and expense spent on maintenance. Planting, or preserving, habitat with native plant species provides the greatest benefit to wildlife and increases biodiversity on the golf course property. Prioritize the planting of species that provide the greatest benefit to wildlife diversity. For example, native species such as oak and aster best sustain species of native butterfly and moth caterpillars (Table 11-1), while non-native species such as Ginkgo or Zelkova support none. Planting these species will be the most effective means to maintain and improve biological diversity of butterflies, moths, and the higher life forms that they sustain such as birds.

Table 11-1. Species that provide the greatest support to native butterfly and moth caterpillars (Tallamy, 2009).

Trees		Perennials	
Common Name (Botanical Name)	Butterfly/Moth Species Supported	Common Name (Botanical Name)	Butterfly/Moth Species Supported
Oak (<i>Quercus</i>)	534	Goldenrod (<i>Solidago</i>)	115
Black Cherry (<i>Prunus</i>)	456	Aster (<i>Symphyotrichum</i>)	112
Willow (<i>Salix</i>)	455	Sunflower (<i>Helianthus</i>)	73
Birch (<i>Betula</i>)	413	Joe Pye Weed (<i>Eutrochium</i>)	42
Poplar (<i>Populus</i>)	368	Blue Grass (<i>Poa</i>)	42
Crabapple (<i>Malus</i>)	311	Sedge (<i>Carex</i>)	36
Blueberry (<i>Vaccinium</i>)	288	Lupine (<i>Lupinus</i>)	33
Maple (<i>Acer</i>)	285	Rye, Blue Wild (<i>Elymus</i>)	31
Elm (<i>Ulmus</i>)	213	Violet (<i>Viola</i>)	30
Pine (<i>Pinus</i>)	203	Wild geranium (<i>Geranium</i>)	24

Cape Arundel Golf Club, Kennebunkport, ME. © Brendan Parkhurst



Best Management Practices for Plant Selection

- Whenever possible, retain 50-70% of the non-play areas in natural cover.
- Choose each plant based on the soil characteristics, climate, sun exposure, water conditions, and existing wildlife.
- Select more stress-tolerant species or cultivars to manage periodic dry/wet conditions.
- Use plants that will perform well over time. Ensure that the mature height and spread of each plant is accounted for to avoid the need for excessive pruning or regular replacement.
- Select plants that don't require excess care to maintain (deadheading, frequent pruning, etc.).
- Group plants with similar water, pH, and nutrient requirements together to allow for the most efficient use of resources.
- Choose plants that are known to occur together naturally to significantly increase the odds of survival and provide the most benefit to wildlife.
- Select each plant based on its unique contribution to the overall design, including flower blooming schedules, bark, fruit, texture, and habitat.
- Where feasible, leave in place the existing understory (brush and young trees) and native grasses and communicate the value of these natural ecosystems to membership.
- The design should look "natural." Replicate the natural system of layers to create harmony and provide year-round interest: a canopy of large shade trees, a medium understory of large shrubs and small flowering trees, a smaller shrub layer, and an herbaceous layer.
- Group plants in odd quantities (1, 3, 5, 7, 9...) for aesthetic appeal.



CHAPTER 12: ENERGY MANAGEMENT

The Ledges Golf Club, York, ME © Forrest Bell

General Energy Efficiency Considerations

Energy, in the form of electricity, natural gas, diesel, propane, heating oil, gasoline and other fuels, is a significant expense on the golf course. According to the Golf Course Superintendents Association of America (GCSAA), the average 18-hole golf course uses nearly 450,000 kilowatt-hours in electricity alone (Golf Course Environmental Profile, 2012). At an average New England electric rate of \$0.16/kWh, that's \$72,000 in annual electricity costs. Fortunately, golf course managers have many opportunities to reduce energy consumption in many areas of course operations. Making investments in energy efficiency saves money for the golf course and positions the course to be more resilient to future increases in energy prices or regulatory changes. An energy-efficient golf course is also a good steward of natural resources and can enhance its standing in the community by publicizing its efforts to reduce energy consumption and carbon emissions.

ENERGY AUDIT

Golf course managers should consider having an energy audit performed for the golf course as a preliminary step before making large capital investments. An energy audit analyzes the energy use of the facility to uncover areas of greatest potential energy savings, allowing a golf course manager to prioritize energy improvements based on return-on-investment. Qualified auditing firms include those holding the Certified Energy Manager (CEM), Certified Energy Auditor (CEA), or Certified Golf Irrigation auditor certifications. The contract with the auditing firm should be clear with respect to the costs and scope of the audit. Some auditors only look at electrical use, while others may only look at the stationary equipment and not the mowing or maintenance equipment.

An irrigation audit may be conducted separately by an irrigation specialist. The golf course's utility company or state energy office is a good source for names of reputable auditors, and many states have programs that provide free or reduced-cost audits or other technical assistance. An energy audit can also identify available incentives and financing to reduce the initial investment of energy efficient equipment.

ESTABLISH AN ENERGY MANAGEMENT PLAN

Once an energy audit has identified areas of greatest concern, the audit's recommendations can be utilized to create an energy management plan with specific goals for reducing energy consumption over time. The manager can prioritize the capital expenditures and make a plan to invest in energy efficient equipment as the budget allows. Priorities depend on an organization's overall goals. For instance, golf courses looking for cost savings may prioritize a different project than courses with a goal of carbon reductions. Managers should periodically review an energy management plan to ensure it remains in line with overall organizational goals. Management can also codify best practices for energy efficiency and provide staff training on energy efficient practices. To track energy consumption, managers can make use of online portals from the electric utility as well as monitoring tools supplied by electronic control systems (e.g. submeters).

ENERGY TRACKING

Organizations should seek to collect energy consumption and cost data from any fuel being utilized in stationary equipment and vehicles. Utilities typically provide online platforms for accessing use and cost. Some vendors may only provide invoices that must be recorded manually or through an automated process. Submetering allows managers to monitor consumption around campus without splitting the main utility service meter.

Data collection is an important step to understandings a golf course's operations. Data is critical for monitoring performance of new or recalibrated equipment. Organization's with a commitment to reducing greenhouse gas (GHG) emissions are typically required to submit energy consumption annually; regular data collection streamlines the preparation of the submission. Finally, understanding the structure of your energy billing is important to evaluating the cost efficacy of potential projects; energy is not always measured and billed volumetrically (e.g. kWh), a demand component may also be a factor (e.g. demand). Manager's should be careful evaluating energy projects using only a volumetric rate (e.g. \$/kWh) if a demand charge is present on the related utility account.

ENERGY PROCUREMENT

The state of Maine is a deregulated energy market for fuels purchased from a utility. This means electricity and natural gas supply may be purchased separate from the utility. Golf course managers may consider energy products with varying risk profiles to meet specific budgetary needs; operational characteristics, such as curtailment capabilities, may open more opportunities in energy contracting. Energy consultants licensed in the state of Maine can help golf course managers navigate the various options in energy products and suppliers.

EMPLOYEE TRAINING

A facilities' employees are on the front lines of energy conservation, as their daily choices can contribute to the facility's energy efficiency. Even simple activities such as turning off lights in empty rooms, following a maintenance checklist to keep equipment clean, and keeping equipment properly calibrated can all contribute to energy savings. Management can consider developing a checklist of employee-led maintenance activities and energy conservation behaviors. Staff training allows the employees to buy in to the energy conservation goals and take ownership of their role in energy management.

Buildings and Amenities

Lighting is used throughout the golf course in both for interior and exterior spaces. Lighting represents one of the easier cost-effective ways to save energy. In the past decade, light-emitting diode (LED) technology has rapidly advanced while costs for these products have decreased by approximately 90%. LED lighting is quickly making other types of lighting obsolete due to the sharp increase in efficiency, decrease in cost, and long life. LEDs can replace not only indoor lighting, but also older mercury vapor or metal halide exterior lighting.

CLUBHOUSE

In a clubhouse, energy efficiency and water conservation measures can be implemented to save energy, including those associated with kitchen equipment, swimming pools, heating, ventilation and air conditioning (HVAC) equipment, bathrooms, and offices.

Best Management Practices for Buildings & Amenities

- Pick LED lights that come with at least a three-year warranty and consult third-party listings like the [Design Lights Consortium](#) to evaluate options.
- Install timers or photocells on outdoor lighting and consider the use of motion/occupancy sensors where appropriate.
- Where motion/occupancy sensors are not workable, train staff to turn off lights when not in use.
- When considering a lighting retrofit, prioritize the oldest lights that also have the longest run time (hours in use per day).

Best Management Practices for Buildings & Amenities (continued)

- Utilize translucent wall panels to provide natural lighting in areas such as equipment maintenance/storage and irrigation pump houses.
- Consider solar energy and other renewable energy sources to reduce overall electric costs for lighting.
- Look for EnergyStar-certified kitchen equipment such as dishwashers, refrigerators, and walk-in coolers. EnergyStar is a joint program of the U.S. Department of Energy and U.S. Environmental Protection Agency that labels energy-efficient appliances and allows the consumer to easily compare the energy consumption of various equipment.
- Ensure kitchen equipment is clean and in good working order. Dirt and dust build-up can lead to wasted energy use and premature equipment failure, so staff should adhere to a maintenance/cleaning checklist.
- Turn off equipment such as burners and broilers when not in use.
- Ensure seals and gaskets around ovens, steamers, refrigerators, and freezers are aligned properly.
- Heat water to the proper setting (140 degrees Fahrenheit) and insulate hot water lines.
- User proper dishwasher setpoints and operation mode. Set rinse pressure to 15 to 25 pounds per square inch to avoid excess water use.
- Set the wash tank temperature to 160 degrees Fahrenheit and the booster heater setpoint to 180 degrees Fahrenheit in accordance with guidelines from NSF International, an organization that develops standards for public health and safety.
- Run the dishwasher only when full and do not run in manual mode.
- Upgrade to low-flow pre-rinse sprayers. Replace sprayers that take less than 30 seconds to fill a one-gallon pail.
- Use a high-efficiency pool heater and consider the use of a solar pool heating system.
- Ensure pool pumps and motors are properly sized and are the most energy-efficient model available.
- Maintain an appropriate water temperature when the pool is in use and turn down the pool heater when not in use.
- Use a pool cover to decrease evaporation when the pool is closed.
- Add windbreaks (trees, shrubs, fencing) around the pool to further reduce evaporation.
- Consider installing EnergyStar-certified commercial boilers with a thermal efficiency of 94% or greater and a turndown ratio of 5:1.

Best Management Practices for Buildings & Amenities (continued)

- Change HVAC filters on a regular basis, typically every one to three months.
- Find an HVAC technician to perform regular check-ups to ensure the HVAC equipment is working properly.
- Consider installing EnergyStar air conditioning equipment, especially if the air conditioner is over 10 years old.
- Use a programmable thermostat in conditioned spaces to reduce heating and cooling costs during periods of low use.
- Properly seal heating and cooling ducts and ensure the ducts are insulated.
- Ensure all new building construction meets current state and federal energy codes
- Ensure that buildings are properly insulated and that leaks are sealed.
- Consider adding advanced digital economizer controls to an existing rooftop HVAC unit. These controls bring in ventilation only when needed, reducing the overall energy consumption of the HVAC unit.
- Consider use of a commercial geothermal or water-source heat pump for heating and cooling. A qualified HVAC technician can inform course managers if this technology is applicable for the golf course's buildings.
- Efficient wood boilers can be an effective way to provide supplemental heat. Burning wood may also eliminate or reduce disposal problems on golf courses that generate wood debris.
- Install low-flow faucets and showerheads.
- Install dual-flush, low-flow toilets.
- Install water-free urinals.
- If the golf course does laundry on the premises, ensure washers and dryers are the most efficient models available. Consider ozone laundry systems as an alternative to large commercial washers. Ozone machines use cold water and ozone gas, instead of hot water and chemicals, to clean and disinfect laundry.
- Look for the EnergyStar label when purchasing office equipment such as computers and photocopiers; set the equipment to power down automatically after a period of inactivity.
- Install energy-efficient vending machines and retrofit older vending machines with a controller that reduces the machine's run time.

Course Management

PUMPING AND IRRIGATION

Irrigation is one of the largest energy users on a golf course, with irrigation pumping contributing about 30% of the electricity use in the average course (2015 Golf Course Environmental Profile). Fortunately, ample opportunities exist for energy savings by optimizing the design of the irrigation system, ensuring pumps and other system components are properly maintained, and utilizing automated sensors and controls.

Regular monitoring and maintenance, as described in the “Irrigation” chapter, is the key to uncovering leaks, waste, and problems that could lead to expensive repairs and wasted energy in the future. Golf course managers should implement a daily, weekly, and monthly maintenance plan with help from their irrigation designer.

CART CHARGING

Golf cart charging can use a lot of electricity. Some of that electric use can be saved by moving the charging time to off-peak hours to reduce demand charges from the utility, as many utilities charge their larger energy users a demand charge for energy used during times of highest electric demand. A golf course with a demand charge can save energy and money by switching certain activities from high-peak times to off-peak times. Typically, off-peak times are early morning and late at night.

MOWING

Conventional commercial lawnmowers use gasoline or diesel fuel. Many innovations in alternative-fuel turf equipment can lower emissions and provide other benefits such as extended life, decreased maintenance, and eliminating the risk of fuel theft or spillage. Alternative-fuel turf equipment has a wide range of price points and features. Golf course managers should thoroughly research the specifications of alternative fueled equipment to make the best decision.



Irrigation system upgrade at Willowdale Golf Club, Scarborough, ME © Rick Lewis

Best Management Practices for Efficient Course Management

- Consider switching golf cart charging to off-peak hours. If it is not possible to switch the entire fleet to off-peak times, see if charging can be staggered to minimize the number of carts being charged in the peak hours.
- New national standards for battery charger energy efficiency took effect in 2018. Consider replacing an older battery charger with a model manufactured according to the new standard to save the most energy.
- Consider solar-charged golf carts. These carts work by using a solar panel on the roof of the cart and can reduce electricity consumption of cart charging by 50% to 75%.
- Compressed natural gas requires less maintenance, extends equipment life, and does not spoil or clog the fuel system during storage.
- Propane has many of the same benefits of compressed natural gas. New propane-powered equipment can be purchased, or some gasoline equipment can be converted to propane using a conversion kit.
- Biodiesel, which can be blended with petroleum diesel without modifying the equipment, reduces emissions. Check with the equipment manufacturer to see if a biodiesel blend is approved for use.
- Commercial electric equipment is powered with rechargeable electric batteries. Recent innovations have improved battery life, enabling extended use of commercial equipment between charges. Solar-powered electric equipment is another recent addition to the market that golf courses can consider.
- Hybrid equipment, using a combination of alternative and traditional fuels, is also available for many fuels and offers improved fuel efficiency and emissions reductions.

Renewable Energy

Maximizing energy efficiency is the first step in reducing a facility's carbon footprint. To further reduce the carbon footprint and reduce energy costs, facilities can invest in renewable energy sources. Some types of renewable energy, such as solar and wind, are highly visible to a golf course's guests and the public and can enhance the golf course's environmental image.

SOLAR PV

Solar photovoltaic (PV) systems have decreased in price dramatically in recent years, making solar system installation an economically viable option for many golf courses, especially when coupled with federal and state incentives. Some considerations for solar include the size of the system, whether to install a ground or roof-mounted system, and whether to lease or own the system. With multiple companies able to install a solar project, golf courses should consider the services of a third-party consultant to help evaluate which solar company offers the best terms.

SOLAR HOT WATER

A solar hot water heater can be a cost-effective way to provide hot water for the golf course. A solar hot water heater can cut water heating expenses by 50% to 80% in some instances. Golf courses should check with a reputable installer to ensure the system makes sense for the needs of the course.

GEOHERMAL HEAT PUMP

A geothermal, or ground-source heat pump uses the Earth's heat in cold weather by drawing up the warmer air from below ground. In warm weather, the heat pump sends warm air back into the Earth to provide cooling. A geothermal heat pump can save 40% to 70% in heating and cooling costs, according to a geothermal industry trade group.

Golf courses should check with a qualified heat pump installer to understand whether a geothermal heat pump is a viable option.

WIND

Smaller-scale wind turbines are available for individual facilities to offset some of their electric use. Golf courses interested in wind power should get a professional evaluation of wind energy potential to make sure the wind speed is sufficient to make a turbine economically viable.

BIOMASS

Biomass is any renewable organic matter that can be used for fuel, with the most common fuel source being biogas created by anaerobic digesters. Another type of biomass heating source is a wood-pellet boiler, which can work in some commercial applications. A golf course that produces a large volume of biomass or organic wastes may find biomass energy generation to be a viable option. Check with a qualified consultant to better understand the costs and benefits of biomass energy.

Solar panels on a building roof at Riverside Golf Course, Portland, ME © FBE



OFF-SITE RENEWABLE ENERGY

Supporting renewable energy projects does not require an on-site installation. End users are able to purchase Renewable Energy Credits (RECs) from renewable energy generation; RECs allow end users to directly support renewable energy and allow energy use with emissions to be offset. Many types of RECs are available at the national and state level and vary across renewable technologies; the cost per REC is dependent on the various factors.

Remote net metering may also be a viable option for golf courses without optimal on-site opportunities for renewables. Net metering allows end users to receive credits on their own utility bills from a remote renewable energy site; contracts may also include RECs associated with the renewable project.

A qualified energy consultant should advise golf courses on off-site renewable energy due to the breadth of options and complex nature of related contracts.

Funding Resources

The U.S. Department of Agriculture administers the [Rural Energy for America Program](#) (REAP) through its Rural Development office. REAP offers grants of up to 25% of project cost and loan guarantees of up to 75% of the project cost to rural businesses that install energy-efficient or renewable-energy projects. To qualify as rural, a golf course must be located in an area with less than 50,000 inhabitants. Grants range from

\$1,500 to \$2,500 for energy efficiency and \$2,500 to \$500,000 for renewable energy. Loans range from \$5,000 to \$25 million.

The Federal Business Energy Investment Tax Credit and Modified Accelerated Cost Recovery System offer [tax benefits](#) to businesses that install renewable energy projects. The 30% tax credit for renewable energy significantly lowers the cost of many renewable energy projects.

The [Section 179D Commercial Buildings Energy Efficiency Tax Deduction](#) provides a deduction of \$1.80 per square foot for installations of lighting, building envelope, and HVAC or hot water systems that reduce the building's total energy use by 50% or more. Facilities can receive a deduction of \$0.60 per square foot for meeting a partial qualification. Tax deductions can be retroactively applied to projects installed since January 1, 2006. Golf course managers should check with their accountants to ensure compliance and can work with a qualified energy consultant to develop the documentation for the tax deduction.

New England states may also have state-based energy efficiency resources and funding opportunities.

REFERENCES

GENERAL REFERENCES

- Aerts, M.O., N. Nesheim, and F. M. Fishel. April 1998; revised September 2015. Pesticide recordkeeping. PI-20. Gainesville, Florida: Institute of Food and Agricultural Sciences, University of Florida. Available: <http://edis.ifas.ufl.edu/PI012>
- Aquatic Ecosystem Restoration Foundation. 2014. Biology and control of aquatic plants: A best management practices handbook: 3rd Ed. Gettys, L.A., W. T. Haller, and D. G. Petty, editors. <http://aquatics.org/bmpchapters/BMP4ed.pdf>
- ASCE, January 2005. The ASCE standardized reference evapotranspiration equation. Final report of the Task Committee on Standardization of Reference Evapotranspiration, Environmental and Water Resources Institute of the American Society of Civil Engineers. 1801 Alexander Bell Drive, Reston, VA 20191 Available: <https://ascelibrary.org/doi/book/10.1061/9780784408056>
- Bohmont, B. 1981. The new pesticide users guide. Fort Collins, Colorado: B & K Enterprises.
- Brecke, B.J., and J.B. Unruh. May 1991; revised February 25, 2003. Spray additives and pesticide formulations. Fact Sheet ENH-82. Gainesville, Florida: Institute of Food and Agricultural Sciences, University of Florida.
- Broder, M.F., and D.T. Nguyen. 1995. Coating concrete secondary containment structures exposed to agrichemicals. Circular Z-361. Muscle Shoals, Alabama: Tennessee Valley Authority, Environmental Research Center. Tel. (205) 386-2714.
- Broder, M.F., and T. Samples. 2002. Tennessee handbook for golf course environmental management. Tennessee Department of Agriculture.
- Buss, E.A. January 2002; revised July 2003. Insect pest management on golf courses. ENY-351. Gainesville, Florida: Institute of Food and Agricultural Sciences, University of Florida.
- Butler, T., W. Martinkovic, and O.N. Nesheim. June 1993; revised April 1998. Factors influencing pesticide movement to groundwater. PI2. Gainesville, Florida: Institute of Food and Agricultural Sciences, University of Florida.
- California Fertilizer Association. 1985. Western fertilizer handbook, 7th ed. Sacramento, California.
- Carrow, R.N., R. Duncan, and C. Waltz. 2007. Best management practices (BMPs) water-use efficiency/conservation plan for golf courses. Available: [https://www.gcsaa.org/uploadedfiles/Environment/Get-Started/BMPs/Water-use-efficiency-and-conservation-best-management-practices-\(Georgia\).pdf](https://www.gcsaa.org/uploadedfiles/Environment/Get-Started/BMPs/Water-use-efficiency-and-conservation-best-management-practices-(Georgia).pdf)
- Carrow, R.N., R.R. Duncan, and D. Wienecke. 2005. BMPs: Critical for the golf industry. Golf Course Management. 73(6):81-84.

- Center for Resource Management. 1996. Environmental principles for golf courses in the United States. 1104 East Ashton Avenue, Suite 210, Salt Lake City, Utah 84106. Tel: (801) 466-3600, Fax: (801) 466-3600.
- Clark, G.A. July 1994. Microirrigation in the landscape. Fact Sheet AE254. Gainesville, Florida: Institute of Food and Agricultural Sciences, University of Florida.
- Clark, Mark and Acomb, Glenn; Florida Field Guide to Low Impact Development: Stormwater Reuse. Univ. Florida 2008. http://buildgreen.ufl.edu/Fact_sheet_Stormwater_Reuse.pdf
- Colorado Nonpoint Source Task Force. 1996. Guidelines for Water Quality Enhancement at Golf Courses Through the Use of Best Management Practices.
- Connecticut Department of Environmental Protection. 2006. Best Management Practices for Golf Course Water Use.
- Cromwell, R.P. June 1993; reviewed December 2005. Agricultural chemical drift and its control. CIR1105. Gainesville, Florida: Institute of Food and Agricultural Sciences, University of Florida. Available: <http://edis.ifas.ufl.edu/AE043>.
- Crow, W.T. February 2001; revised November 2005. Nematode management for golf courses in Florida. ENY-008 (IN124). Gainesville, Florida: Institute of Food and Agricultural Sciences, University of Florida. Available: <http://edis.ifas.ufl.edu/IN124>.
- Daum, D.R., and T.F. Reed. n.d. Sprayer nozzles. Ithaca, New York: Cornell Cooperative Extension. Available <http://psep.cce.cornell.edu/facts-slides-self/facts/gen-peapp-spray-nozz.aspx>.
- Dean, T.W. February 2003. Pesticide applicator update: Choosing suitable personal protective equipment. PI-28. Gainesville, Florida: Institute of Food and Agricultural Sciences, University of Florida. Available: <http://edis.ifas.ufl.edu/PI061>.
- April 2004; revised November 2004. Secure pesticide storage: Facility size and location. Fact Sheet PI-29. Gainesville, Florida: Institute of Food and Agricultural Sciences, University of Florida.
- April 2004; revised November 2004. Secure pesticide storage: Essential structural features of a storage building. Fact Sheet PI-30. Gainesville, Florida: Institute of Food and Agricultural Sciences, University of Florida.
- Dean, T.W., O.N. Nesheim, and F. Fishel. Revised May 2005. Pesticide container rinsing. PI-3. Gainesville, Florida: Institute of Food and Agricultural Sciences, University of Florida. Available: <http://edis.ifas.ufl.edu/PI003>
- Delaware Nutrient Management Commission. 2006. Water quality best management practices: nutrients, irrigation and pesticides for golf course. Athletic Turf, Lawn Care and Landscape Industries.
- Dodson, R.G. 2000. Managing wildlife habitat on golf courses. Sleeping Bear Press. Chelsea, MI.
- Elliott, M.L., and G.W. Simone. July 1991; revised April 2001. Turfgrass disease management. SS-PLP-14. Gainesville, Florida: Institute of Food and Agricultural Sciences, University of Florida. Available: <http://edis.ifas.ufl.edu/LH040>.

- Fishel, F.M. March 2005. Interpreting pesticide label wording. Gainesville, Florida: Institute of Food and Agricultural Sciences. Available: <http://edis.ifas.ufl.edu/PI071>.
- Fishel, F.M., and Nesheim, O.N. November 2006. Pesticide safety. FS11. Gainesville, Florida: Institute of Food and Agricultural Sciences. Available: <http://edis.ifas.ufl.edu/pdf/CV/CV10800.pdf>.
- Florida Department of Agriculture and Consumer Services. n.d. Pesticide recordkeeping—benefits and requirements. Available: <https://www.fdacs.gov/content/download/2983/file/PRP%205-05.pdf>.
- Florida Department of Agriculture and Consumer Services. Division of Agricultural Environmental Services. Suggested pesticide recordkeeping form. Available: <https://www.freshfromflorida.com/content/download/2990/18861/Suggested%20Pesticide%20Recordkeeping%20Form.pdf>.
- Division of Agricultural Environmental Services. Suggested pesticide recordkeeping form for organo-auxin herbicides. Available: <http://forms.freshfromflorida.com/13328.pdf>.
- Florida Department of Agriculture and Consumer Services and Florida Department of Environmental Protection. 1998. Best management practices for agricultural chemical handling and farm equipment maintenance. Available: <http://www.dep.state.fl.us/water/nonpoint/docs/nonpoint/agbmp3p.pdf>
- Florida Department of Environmental Protection. 2008. Florida stormwater, erosion, and sedimentation control inspector's manual. Tallahassee, Florida: Nonpoint Source Management Section, MS 3570, 3900 Commonwealth Blvd., Tallahassee, Florida 32399-3000. Available: <http://www.dep.state.fl.us/water/nonpoint/docs/erosion/erosion-inspectors-manual.pdf>
- December 27, 2002. Environmental risks from use of organic arsenical herbicides at south Florida golf courses. FDEP white paper.
- April 2002. Florida water conservation initiative. Available: <https://fdep.maps.arcgis.com/apps/Cascade/index.html?appid=473b768b4af049bf91b2879b83ea961c>
- Florida Department of Environmental Protection. 2015. Florida-friendly Best Management Practices for Protection of Water Resources by the Green Industries. Third printing. Available: <https://fyn.ifas.ufl.edu/pdf/grn-ind-bmp-en-12-2008.pdf>
- Florida Department of Environmental Protection. 2007. Best Management Practices for The Enhancement of Environmental Quality on Florida Golf Courses. Florida Department of Environmental Protection. Third printing. Available: <http://www.dep.state.fl.us/water/nonpoint/docs/nonpoint/glfbmp07.pdf>.
- Revised August 2009. A guide on hazardous waste management for Florida's auto repair shops. Available: <https://docplayer.net/7520340-A-guide-on-hazardous-waste-management-for-florida-s-auto-repair-and-paint-and-body-shops.html>.
- October 2005. Checklist guide for 100% closed loop recycle systems at vehicle and other equipment wash facilities. Available: https://floridadep.gov/sites/default/files/ChecklistGuideClosed-LoopRecycleSystems_0.pdf.

- October 2005. Guide to best management practices for 100% closed-loop recycle systems at vehicle and other equipment wash facilities. Pollution Prevention Program and Industrial Wastewater Section. Available: <https://floridadep.gov/sites/default/files/GuideBMPClosed-LoopRecycleSystems.pdf>.
- Florida Department of Environmental Protection. 2006. State of Florida erosion and sediment control designer and reviewer manual. Nonpoint Source Management Section. Available: <https://floridadep.gov/dear/florida-stormwater-erosion>.
- Florida Department of Environmental Protection. 2016. Operation cleansweep for pesticides website. Available: <http://www.dep.state.fl.us/waste/categories/cleansweep-pesticides>.
- Florida Department of Environmental Protection. 2005. Standards and specifications for turf and landscape irrigation systems. Fifth Edition. Available: <http://ufdc.ufl.edu/UF00076845/00001>.
- Committee on Landscape Irrigation and Florida-Friendly Design Standards. 2006. Landscape Irrigation & Florida-Friendly Design Standards. Florida Department of Environmental Protection, Office of Water Policy, 3900 Commonwealth Blvd., MS 46, Tallahassee, FL 32399-3000. Available: https://www.swfwmd.state.fl.us/sites/default/files/medias/documents/40D-002.0912l_to_1d-DEP_LandscapeIrrigationFloridaFriendlyDesign_eff_042710.pdf
- Gilman, E. 2006. Pruning shade trees in landscapes. Available: <https://hort.ifas.ufl.edu/woody/pruning.shtml>.
- Golf Course Superintendents Association of America. 2012. Golf Course Environmental Profile; Volume IV; Energy Use and Energy Conservation Practices on U.S. Golf Courses. Available: <https://www.gcsaa.org/Uploadedfiles/Environment/Environmental-Profile/Energy/Golf-Course-Environmental-Profile--Energy-Use-and-Conservation-Report.pdf>
- LandStudies, Inc. and The Pennsylvania Environmental Council. Golf course water resources handbook of best management practices (Pennsylvania). 2009. Available: <http://pecpa.org/wp-content/uploads/Golf-Course-Water-Resources-Handbook-of-Best-Management-Practices.pdf>
- Havlin, J.L., S.L. Tisdale, J.D. Beaton, and W.L. Nelson. 2004. Soil fertility and fertilizers, 7th Ed. Prentice Hall.
- Haydu, J.J., and A.W. Hodges. 2002. Economic impacts of the Florida golf course industry. UF-IFAS Report EIR 02-4. Available: <https://fred.ifas.ufl.edu/pdf/economic-impact-analysis/EIR02-4r.pdf>.
- Helfrich, L.A., D.L. Weigmann, P. Hipkins, and E.R. Stinson. June 1996. Pesticides and aquatic animals: A guide to reducing impacts on aquatic systems. Virginia Cooperative Extension Service. Publication Number 420-013. Available: <https://www.pubs.ext.vt.edu/420/420-013/420-013.html>.
- Hornsby, A.G., T.M. Buttler, L.B. McCarty, D.E. Short, R.A. Dunn, G.W. Simone. Revised September 1995. Managing pesticides for sod production and water quality protection. Circular 1012. Gainesville, Florida: Institute of Food and Agricultural Sciences, University of Florida.
- Insecticide Resistance Action Committee Website. Available: <http://www.irac-online.org/>.

- King, K.W., and J.C. Balogh. 2001. Water quality impacts associated with converting farmland and forests to turfgrass. Transactions of the ASAE, Vol. 44(3): 569-576. Available: <https://www.ars.usda.gov/ARSEUserFiles/3013/King12.pdf>.
- Lehtola, C.J., C.M. Brown, and W.J. Becker. November 2001. Personal protective equipment. OSHA Standards 1910.132-137. AE271. Gainesville, Florida: Institute of Food and Agricultural Sciences, University of Florida. Available: <https://ufdc.ufl.edu/IR00002922/00001>.
- McCarty, L.B., and D.L. Colvin. 1990. Weeds of southern turfgrasses. Gainesville, Florida: University of Florida.
- Midwest Plan Service. Revised 1995. Designing facilities for pesticide and fertilizer containment. MWPS-37. Midwest Plan Service, 122 Davidson Hall, Iowa State University, Ames, IA 50011-3080. Tel.: (515) 294-4337. Available: <https://www-mwps.sws.iastate.edu/catalog/construction-farm/designing-facilities-pesticide-and-fertilizer-containment-pdf>.
- Mitra, S. 2006. Effects of recycled water on turfgrass quality maintained under golf course fairway conditions. WateReuse Foundation, 1199 North Fairfax Street, Suite 410, Alexandria, VA 22314.
- National Pesticide Information Center. 2008. Signal words. Topic Fact Sheet. Available: <http://npic.orst.edu/factsheets/signalwords.pdf>.
- Nesheim, O.N., and F.M. Fishel. September 2007, reviewed August 2013. Interpreting PPE statements on pesticide labels. PI16. Gainesville, Florida: Institute of Food and Agricultural Sciences, University of Florida.
- Nesheim, O.N., and F.M. Fishel. March 1989; revised November 2005. Proper disposal of pesticide waste. PI-18. Gainesville, Florida: Institute of Food and Agricultural Sciences, University of Florida. Available: <http://edis.ifas.ufl.edu/PI010>.
- Nesheim, O.N., F.M. Fishel, and M. Mossler. July 1993, revised 2005 and 2017, reviewed April 2020. Toxicity of pesticides. PI-13. Gainesville, Florida: Institute of Food and Agricultural Sciences, University of Florida. Available: <http://edis.ifas.ufl.edu/pdf/PI/PI00800.pdf>.
- O'Brien, P. July/August 1996. Optimizing the turfgrass canopy environment with fans. USGA Green Section Record, Vol. 34(4), 9-12 Available: <http://gsrpdf.lib.msu.edu/ticpdf.py?file=/1990s/1996/960709.pdf>.
- O'Brien, P., and C. Hartwiger. March/April 2003. Aerification and sand topdressing for the 21st century. USGA Green Section Record, Vol. 41(2), 1-7.
- Olexa, M.T., A. Leviten, and K. Samek. December 2008, revised December 2013. Florida solid and hazardous waste regulation handbook: Table of contents. FE758. Gainesville, Florida: Institute of Food and Agricultural Sciences, University of Florida.
- Otterbine Barebo, Inc. 2003. Pond and lake management. 3840 Main Road East, Emmaus, PA 18049.
- Peterson, A. 2000. Protocols for an IPM system on golf courses. University of Massachusetts Extension Turf Program. Available: <https://umassextensionbookstore.com/node/41>.

- LandStudies, Inc. and the Pennsylvania Environmental Council. June 2009. Golf course water resources handbook of best management practices. Pennsylvania Environmental Protection. Available: <http://pecpa.org/wp-content/uploads/Golf-Course-Water-Resources-Handbook-of-Best-Management-Practices.pdf>
- Pettinger, N.A. 1935. Useful chart for teaching the relation of soil reaction to availability of plant nutrients to crops. Virginia Agri. Ext. Bul. 136, 1-19.
- Portness, R.E., J.A. Grant, B. Jordan, A.M. Petrovic, and F.S. Rossi. 2014. Best management practices for New York State golf courses. Cornell Univ. Available: http://nysgolfbmp.cals.cornell.edu/ny_bmp_feb2014.pdf
- Rao, P.S.C., and A.G. Hornsby. May 1993, revised December 2001. Behavior of pesticides in soils and water. Fact Sheet SL40. Gainesville, Florida: Institute of Food and Agricultural Sciences, University of Florida. Available: https://www.fws.gov/PanamaCity/resources/Envirothon%20Study%20Guides/2017/Pesticides%201_Behavior%20of%20Pesticides%20in%20Soils%20and%20Water.pdf.
- Rao, P.S.C., R.S. Mansell, L.B. Baldwin, and M.F. Laurent. n.d. Pesticides and their behavior in soil and water. Ithaca, New York: Cornell Cooperative Extension. Available: <http://psep.cce.cornell.edu/facts-slides-self/facts/gen-pubre-soil-water.aspx>.
- Rodgers, J. n.d. Plants for lakefront revegetation. Invasive Plant Management, Florida Department of Environmental Protection, 3900 Commonwealth Blvd., MS 705, Tallahassee, FL 32399. Available: https://www.pinellascounty.org/environment/watershed/pdf/adoptapond/Lakefront_Revegetation.pdf.
- Shaddox, T.W. 2000. General recommendations for fertilization of turfgrasses on Florida soils. Fact Sheet SL-21. Gainesville, Florida: Institute of Food and Agricultural Sciences, University of Florida. Available: <http://edis.ifas.ufl.edu/LH014>.
- Shaddox, T.W. and J.B. Sartain. 2001. Soil testing and interpretation for Florida turfgrasses. SL-181. Gainesville, Florida: Institute of Food and Agricultural Sciences, University of Florida. Available: <http://edis.ifas.ufl.edu/SS317>.
- Shaddox, T. W. February 2002, revised 2007, 2013, and 2016. Recommendations for N, P, K, and Mg for golf course and athletic field fertilization based on Mehlich-I extractant. SL-191. Available: <https://edis.ifas.ufl.edu/ss404>.
- Shaddox, T.W. November 1980, revised 2008, 2015, and 2017.. The Florida fertilizer label. SL-3. Gainesville, Florida: Institute of Food and Agricultural Sciences, University of Florida. Available: <https://edis.ifas.ufl.edu/pdf/files/SS/SS17000.pdf>.
- Sartain, J.B., G.L. Miller, G.H. Snyder, and J.L. Cisar. 1999a. Plant nutrition and turf fertilizers: Best management practices for Florida golf courses. J.B. Unruh and M. Elliott (Eds.). SP-141 2nd ed. Gainesville, Florida: Institute of Food and Agricultural Sciences, University of Florida.
- Sartain, J.B., G.L. Miller, G.H. Snyder, and J.L. Cisar. 1999b. Liquid fertilization and foliar feeding: Best management practices for Florida golf courses. J.B. Unruh and M. Elliott (Eds.). SP-141 2nd ed.

Gainesville, Florida: Institute of Food and Agricultural Sciences, University of Florida.

Sartain, J.B., G.L. Miller, G.H. Snyder, and J.L. Cisar. 1999c. Fertilization programs: Best management practices for Florida golf courses. J.B. Unruh and M. Elliott (Eds.). P-141 2nd ed. Gainesville, Florida: Institute of Food and Agricultural Sciences, University of Florida.

Schueler, T.R. 2000. Minimizing the impact of golf courses on streams: The practice of watershed protection. Center for Watershed Protection, Ellicott City, MD., 673-675.

Schumann, G.L., P.J. Vittum, M.L. Elliott, and P.P. Cobb. January 1998. IPM handbook for golf courses. Indianapolis, Indiana: Wiley Publishing, Inc.

Seelig, B. July 1996. Improved pesticide application BMP for groundwater protection from pesticides. AE-1113. Fargo, North Dakota: North Dakota State University Extension Service.

Smajstrla, A.G., and B.J. Boman. April 2000. Flushing procedures for microirrigation systems. Bulletin 333. Gainesville, Florida: Institute of Food and Agricultural Sciences, University of Florida. Available: <http://irrigationtoolbox.com/ReferenceDocuments/Extension/Florida/WI01300.pdf>.

Staples, A.J. July 2009. Golf course energy use Part 2: Pump stations. Golf Course Management. <https://www.gcsaa.org/Uploadedfiles/Environment/Resources/Energy-Conservation/Golf-course-energy-use-Part-2-Pump-stations.pdf>.

Tennessee Department of Agriculture. n.d. Tennessee Handbook for Golf Course Environmental Management.

Thostenson, A., C. Ogg, K. Schaefer, M. Wiesbrook, J. Stone, and D. Herzfeld. 2016. Laundering pesticide-contaminated work clothes. PS 1778. Fargo, ND. North Dakota. Available: <https://www.ag.ndsu.edu/publications/crops/laundrying-pesticide-contaminated-work-clothes>.

State Univ. Cooperative Extension. <https://www.ag.ndsu.edu/pubs/plantsci/pests/ps1778.pdf>

Trautmann, N.M., K.S. Porter, and R.J. Wagenet. n.d. Pesticides and groundwater: A guide for the pesticide user. Fact Sheet. Ithaca, New York: Cornell Cooperative Extension.

University of Florida—Institute of Food and Agricultural Sciences. Center for aquatic and invasive plants website. Available: <http://plants.ifas.ufl.edu/>.

Buss, L.J. December 1993, revised 2013, and 2019. Insect Identification Service. Available: <http://edis.ifas.ufl.edu/SR010>.

Nematode Assay Laboratory Website. Available: <http://entnemdept.ufl.edu/nematology-assay-lab/>.

Pesticide Information Office Website. Available: <https://pested.ifas.ufl.edu/>.

Plant Disease Clinic Website. Available: <http://plantpath.ifas.ufl.edu/extension/plant-diagnostic-center/>

Rapid Turfgrass Diagnostic Service Website. Available: <http://turfpath.ifas.ufl.edu/rapiddiag.shtml>.

- Unruh, J.B. November 1993, revised 2006. Pesticide calibration formulas and information. Fact Sheet ENH-90. Gainesville, Florida: Institute of Food and Agricultural Sciences, University of Florida. Available: <http://edis.ifas.ufl.edu/WG067>.
- Unruh, J.B. 2012. University of Florida's pest control guide for turfgrass managers. Gainesville, Florida: Institute of Food and Agricultural Sciences, University of Florida. Available: https://turf.ifas.ufl.edu/pdf/2012_UF_Pest_Control_Guide.pdf.
- Unruh, J.B., and B.J. Brecke. Revised January 1998. Response of turfgrass and turfgrass weeds to herbicides. ENH-100. Gainesville, Florida: Department of Environmental Horticulture, University of Florida.
- Unruh, J.B., and M. Elliot. 1999. Best management practices for Florida golf courses, 2nd ed. UF-IFAS Publication SP-141. Gainesville, Florida.
- Unruh, J.B., J.L. Cisar, and G.L. Miller. 1999. Mowing: Best management practices for Florida golf courses. J.B. Unruh and M.L. Elliot (Eds.). 2nd ed. Gainesville, Florida: University of Florida Institute of Food and Agricultural Sciences.
- Unruh, J.B., A.E. Dudeck, J.L. Cisar, and G.L. Miller. 1999. Turfgrass cultivation practices: Best management practices for Florida golf courses. J.B. Unruh and M.L. Elliot (Eds.). 2nd ed. Gainesville, Florida: University of Florida Institute of Food and Agricultural Sciences.
- U.S. Environmental Protection Agency. 2005. GreenScapes: Environmentally beneficial landscaping. Washington, D.C. Office of Solid Waste and Emergency Response. Available: <https://archive.epa.gov/greenbuilding/web/pdf/brochure.pdf>
- United States Golf Association. 2004. Recommendations for a method of putting green construction. Available: <http://www.usga.org/content/dam/usga/images/course-care/2004%20USGA%20Recommendations%20For%20a%20Method%20of%20Putting%20Green%20Cons.pdf>.
- Van Es., H.M. October 1990. Pesticide management for water quality: Principles and practices. October 1990. Ithaca, New York: Cornell Cooperative Extension. Available: <http://psep.cce.cornell.edu/facts-slides-self/facts/pestmgt-water-qual-90.aspx>.
- Virginia Golf Course Superintendents Association. 2012. Environmental Best Management Practices for Virginia's Golf Courses. Available: https://pubs.ext.vt.edu/ANR/ANR-48/ANR-48_pdf.pdf
- White, C.B. 2000. Turfgrass manager's handbook for golf course construction, renovation, and grow-in. Sleeping Bear Press. Chelsea, MI.
- Witt, J.M. n.d. Agricultural spray adjuvants. Ithaca, New York: Cornell Cooperative Extension. Available: <http://psep.cce.cornell.edu/facts-slides-self/facts/gen-peapp-adjuvants.aspx>.
- Yergert, M.B. Austin, and R. Waskom. June 1993. Best management practices for turfgrass production. Turf BMP Fact Sheet. Colorado Department of Agriculture. Agricultural Chemicals and Groundwater Protection Program. Available: http://hermes.cde.state.co.us/drupal/islandora/object/co%3A3063/datastream/OBJ/download/Best_management_practices_for_turfgrass_production.pdf.

CHAPTER-SPECIFIC REFERENCES

Planning, Design, and Construction References

Maine Department of Environmental Protection. 2019. Wetland Types. Available: <https://www.maine.gov/dep/water/wetlands/types.html>

Irrigation References

Kostka, S., J.L. Cisar, C.J. Ritsema, L.W. Dekker, M.A. Franklin, S. Mitra, and S.E. McCann. 2005. Surfactants and soil water repellency in golf course soils—water use and environmental implications. Available: https://www.researchgate.net/publication/228433241_Surfactants_and_soil_water_repellency_in_golf_course_soils-water_use_and_environmental_implications

Landschoot, Peter. 2016. Irrigation Water Quality Guidelines for Turfgrass Sites. Pennsylvania State Extension. Available: <https://extension.psu.edu/irrigation-water-quality-guidelines-for-turfgrass-sites>

Golf Turf Fertilization and Nutrient Management References

Albrecht, W.A. 1975. The Albrecht papers. Vol. 1: Albrecht's Foundation Concepts. C. Walters, editor. Acres USA, Kansas City.

Bear, F.E., A.L. Prince, and J.L. Malcolm. 1945. Potassium needs of New Jersey soils. New Brunswick, N.H. New Jersey Agricultural Experiment Station. New Brunswick.

Bell, G.E., J.K. Kruse, and J.M. Krum. 2013. The Evolution of Spectral Sensing and Advances in Precision Turfgrass Management. *Turfgrass: Biology, Use, and Management*. p. 1151–1188. doi:10.2134/agronmonogr56.c30

Bosworth, S. 2017. Fertilization and Nutrient Management Guidelines for Golf Turf in Vermont. University of Vermont Extension. Available: http://pss.uvm.edu/ag_testing/Soil_Fertility_Recommendations_for_Vermont_Golf_Turf_UVM_2017.pdf

Carey, P., Q. Ketterings, and M. Hunter. 2006. Liming Materials. Agronomy Fact Sheet Series No. 7. Cornell University Cooperative Extension. Available: <http://cceonondaga.org/resources/liming-materials>.

Chaganti, V.N., and S.W. Culman. 2017. Historical Perspective of Soil Balancing Theory and Identifying Knowledge Gaps: A Review. *Crops, Forage & Turfgrass Management*. Vol. 3. Available: <https://access.onlinelibrary.wiley.com/doi/10.2134/cftm2016.10.0072>

Dest, W.M., and K. Guillard. 2001. Bentgrass response to K fertilization and K release rates from eight sand rootzone sources used in putting green construction. *International Turfgrass Society Research Journal*. 9. p. 375-381. Available: https://opencommons.uconn.edu/plsc_articles/13/

Dest, W.M., and K. Guillard. 1987. Nitrogen and Phosphorus Nutritional Influences on Bentgrass-Annual Bluegrass Community Composition. *Journal of the American Society of Horticultural Science*.

112. p. 769-773.

- Eckert, D.J. 1987. Soil test interpretations: Basic cation saturation ratios and sufficiency levels. p. 53–64. J.R. Brown, editor. Soil testing: Sampling, Correlation, Calibration, and Interpretation. SSSA Special Publication. 21. Soil Science Society of America, Madison, WI.
- Fitzpatrick, R.J.M, and K. Guillard. 2004. Kentucky bluegrass response to potassium and nitrogen fertilization. *Crop Science*. 44. p. 1721–1728. doi:10.2135/cropsci2004.1721. Available: https://opencommons.uconn.edu/plsc_articles/3/
- Geng, X., K. Guillard, and T.F. Morris. 2014. Relating turfgrass and quality to frequently measured soil nitrate. *Crop Science*. 54. p. 366–382. doi:10.2135/cropsci2013.03.0145. Available: <https://pdfs.semanticscholar.org/a48e/cd29667f450909384bc70682c350bd772e15.pdf>.
- Guillard, K., R.J.M. Fitzpatrick, and H. Burdett. 2016. Can frequent measurement of NDVI and soil nitrate guide nitrogen fertilization of Kentucky bluegrass sod? *Crop Science*. 56. p. 827–836. doi:10.2135/cropsci2015.06.0347.
- Guillard, K., and K.L. Kopp. 2004. Nitrogen fertilizer form and associated nitrate leaching from cool-season lawn turf. *Journal of Environmental Quality*. 33. p. 1822–1827. doi:10.2134/jeq2004.1822. Available: https://digitalcommons.usu.edu/cwel_pubs/13/.
- Hoffman, L. J.S. Ebdon, W.M. Dest, and M. DaCosta. 2010. Effects of nitrogen and potassium on wear mechanisms in perennial ryegrass: I. wear tolerance and recovery. *Crop Science*. 50. p. 357–366. doi:10.2135/cropsci2008.08.0473
- Hoskins, B.R. 1997. Soil testing handbook for professionals in agriculture, horticulture, nutrient and residuals management. Third edition. Maine Forestry & Agricultural Experiment Station. University of Maine, Orono, ME. Available: <https://umaine.edu/soiltestinglab/wp-content/uploads/sites/227/2016/07/handbook.pdf>
- Inguagiato, J.C., and K. Guillard. 2016. Foliar N concentration and reflectance meters to guide N fertilization for anthracnose management of annual bluegrass putting green turf. *Crop Science*. 56. p. 3328–3337. doi:10.2135/cropsci2015.12.0765.
- Jones Jr., J.B. 1980. Turf analysis. *Golf Course Management*. 48(1). p. 20–32.
- Karcher, D.E., and M.D. Richardson. 2013. Digital image analysis in turfgrass research. J.C. Stier, B.P. Horgan, and S.A. Bonos, editors, *Turfgrass: Biology, Use, and Management*. p. 1133–1149. doi:10.2134/agronmonogr56.c29.
- Kopittke, P.M., and N.W. Menzies. 2007. A review of the use of the basic cation saturation ratio and the ‘ideal’ soil. *Soil Science Society of America Journal*. 71. p. 259–265. doi:10.2136/sssaj2006.0186.
- Kopp, K.L., and K. Guillard. 2002. Clipping management and N fertilization of turfgrass: Growth, N utilization, and quality. *Crop Science*. 42. p. 1225–1231. doi:10.2135/cropsci2002.1225.
- Kopp, K.L, and K. Guillard. 2004. Decomposition rates and nitrogen release of turf grass clippings. In: T. Fischer, et al, editors. *New Directions for a Diverse Planet: Proceedings for the 4th International Crop Science Congress*. The Regional Institute Ltd., Gosford, Australia. Available: <http://www>.

- Mangiafico, S.S., and K. Guillard. 2006. Fall Fertilization Effects on Nitrate Leaching and Turfgrass Color and Growth. *Journal of Environmental Quality*. 35. p. 163–171. doi:10.2134/jeq2005.0061.
- Moore, D.B., K. Guillard, X. Geng, T.F. Morris, and W.F. Brinton. 2019a. Predicting cool-season turfgrass response with Solvita soil tests, Part 1: labile amino-nitrogen concentrations. *Crop Science*. 59. p. 1779–1788. doi:10.2135/cropsci2018.11.0706.
- Moore, D.B., K. Guillard, T.F. Morris, and W.F. Brinton. 2019b. Predicting cool-season turfgrass response with Solvita soil tests, Part 2: CO₂-burst carbon concentrations. *Crop Science*. 59. p. 2237–2248. doi:10.2135/cropsci2018.11.0707.
- Owen, M., J. Lanier, S. Ebdon, and J. Spargo. 2016. Soil & nutrient management. M.C. Owen and J.D. Lanier, editors. *Best Management Practices for Lawn and Landscape Turf*. Version 1.51. UMass Extension Turf Program. Available: http://ag.umass.edu/sites/ag.umass.edu/files/pdf-doc-ppt/lawn_bmp_establishment_2016_final.pdf.
- Petrovic, A.M. 1990. The fate of nitrogenous fertilizers applied to turfgrass. *Journal of Environmental Quality*. 19. p. 1–14. doi:10.2134/jeq1990.00472425001900010001x.
- Saunders, O. 2018. Understanding Your Soil Test Results. University of New Hampshire Cooperative Extension. Available: https://extension.unh.edu/resources/files/Resource000496_Rep518.pdf.
- Schlossberg, M., and J. Simmons. 2012. SportsTurf's Point–Counterpoint: SLAN vs. BCSR. *SportsTurf*. January. p. 24–31. Available: <https://sturf.lib.msu.edu/article/2012jan24.pdf>.
- Schmid, C.J., J.A. Murphy, B.B. Clarke. And M. DaCosta. 2016. Observations on the effect of potassium on winter injury of annual bluegrass in New Jersey in 2015. *Crops, Forage, & Turfgrass Management*. 2. p. 1–4. doi:10.2134/cftm2015.0170
- St. John, R.A. and N.E. Christians. 2013. Basic cation saturation ratio theory applied to sand-based putting greens. *International Turfgrass Society Research Journal*. 12. p. 581–592.
- St. John, R.A., and N.E. Christians. 2010. Special approaches are needed when testing calcareous sands. *Applied Turfgrass Science*. doi:10.1094/ATS-2010-0831-01-RS.
- Stowell, L., and M. Woods. 2013. Minimum levels for sustainable nutrition (MLSN). *Applied Turfgrass Science Proceedings: Constructed Rootzones 2012*. *Applied Turfgrass Science*. Available: <https://dl.sciencesocieties.org/publications/ats/abstracts/10/1/ATS-2013-0008BC>
- Webster, D.E., and J.S. Ebdon. 2005. Effects of nitrogen and potassium fertilization on perennial ryegrass cold tolerance during deacclimation in late winter and early spring. *HortScience*. 40. p. 842–849.
- Woods, M., L. Stowell, and W. Gelernter. 2014. Just what the grass requires: using minimum levels for sustainable nutrition. *Golf Course Management*. January. p. 132– 138.
- Woods, M.S., L.J. Stowell, and W.D. Gelernter. 2016. Minimum soil nutrient guidelines for turfgrass developed from Mehlich 3 soil test results. *PeerJ Preprints*. 4:e2144v1 Available: <https://doi.org/10.7287/peerj.preprints.2144v1>.

Cultural Practices References

- American Society for Testing and Materials. 2018. Standard test method for particle size analysis and sand shape grading of golf course putting green and sports field rootzone mixes. Annual Book of ASTM Standards. Designation: F 1632-03(2018). Vol. 15.07. Available: <https://www.astm.org/Standards/F1632.htm>.
- American Society for Testing and Materials. 2018. Standard Test Methods for Saturated Hydraulic Conductivity, Water Retention, Porosity, and Bulk Density of Athletic Field Rootzones. Annual Book of ASTM Standards. Designation: F 1815-11(2018). Vol. 15.07. Available: <https://www.astm.org/Standards/F1815.htm>.
- Beard, J.1972. Turfgrass science and culture. Prentice Hall, Englewood Cliffs, N.J.
- Calhoun, R.N., G.J. Rinehart, A.D. Hathaway, and D.D. Buhler. 2005. Maximizing cultural practices to minimize weed pressure and extend herbicide treatment Interval in a cool-season turfgrass mixture. International Turfgrass Society Research Journal. 10(Part 2). p. 1184-1188.
- Carrow, R.N. 2003. Turfgrass and environmental research online: Surface organic matter in bentgrass greens. USGA Turfgrass Environmental Research Online. 2(17). p. 1-12. Available: <https://usgatero.msu.edu/v02/n17.pdf>.
- Crider, F.J. 1955. Root-growth stoppage resulting from defoliation of grass. USDA Technical Bulletin No. 1102.
- Gardner, D.S., and R.M. Goss. 2013. Management of turfgrass in shade. J.C. Stier, B.P. Horgan, and S.A. Bonos, editors, Turfgrass: Biology, Use, and Management. p. 219-247. doi:10.2134/agronmonogr56.c6.
- Gaussoin, R.E., and B.E. Branham. 1989. Influence of cultural factors on species dominance in a mixed stand of annual bluegrass/creeping bentgrass. Crop Science. 29(2). p. 480-484.
- Juska, F.V., J. Tyson, and C.M. Harrison. 1955. The competitive relationship of merion bluegrass as influenced by various mixtures, cutting heights, and levels of nitrogen. Agronomy Journal. November 47(11). p. 513-518.
- Kopp, K.L., and K. Guillard. 2002. Clipping management and nitrogen fertilization of turfgrass growth, nitrogen fertilization, and quality. Crop Science. 42. p. 1225-1231. doi:10.2135/cropsci2002.1225.
- Krans, J.V., and J.B. Beard. 1985. Effects of clipping on growth and physiology of 'Merion' Kentucky bluegrass. Crop Science. 21(1). p. 17-20.
- Liu, X., and B. Huang, 2002. Mowing effects on root production, growth, and mortality of creeping bentgrass. 42. p. 1241-1250. doi:10.2135/cropsci2002.1241.
- Murphy, J.A., and P.E. Rieke. 1994. High pressure water injection and core cultivation of a compacted putting green. Agronomy Journal. 86(4). p. 719-724.
- Nikolai, T.A., P.E. Reike, J.N. Rogers III, and J.M. Vargas Jr. 2001. Turfgrass and soil responses to lightweight

rolling on putting green root zone mixes. *International Turfgrass Society Research Journal*. 9(Part 2). p. 604-609.

Steinke, K. and E. Ervin, E. 2013. *Turfgrass Ecology*. J.C. Stier, B.P. Horgan, and S.A. Bonos, editors, *Turfgrass: Biology, Use, and Management*. p. 355-357. doi:10.2134/agronmonogr56.c10.

Integrated Pest Management References

Cahill, J.V., J.J. Murray, N.R. O'Neil, and P.H. Dernoeden. 1983. Interrelationships between family and red thread fungal disease of turfgrass. *Plant Disease*. 67. p. 1080-1083.

Dernoeden, P.H. 1987. Management of take-all patch of creeping bentgrass with nitrogen, sulfur, and phenyl mercury acetate. *Plant Disease*. 17. p. 226-229.

Ellram, A., B. Horgan, and B. Hulke. 2007. Mowing strategies and dew removal to minimize dollar spot on creeping bentgrass. *Crop Science*. 47. p. 2129-2137.

Fidanza, M.A., and P.H. Dernoeden. 1996. Interaction of nitrogen source, application timing, and fungicide on rhizoctonia blight in ryegrass. *HortScience*. 31. p. 389-392.

Giordano P.R., et al. 2012. Timing and frequency effects of lightweight rolling on dollar spot disease in creeping bentgrass putting greens. *Crop Science*. 52. p. 1371- 1378.

Inguagiato, J. and K. Guillard. 2016. Foliar N concentration and reflectance meters to guide N fertilization for anthracnose management of annual bluegrass putting green turf. *Crop Science*. 56. p. 3328-3337.

Inguagiato, J.C., J.A. Murphy, and B.B. Clarke. 2008. Anthracnose severity on annual bluegrass influenced by nitrogen fertilization, growth regulators, and verticutting. *Crop Science*. 48. p. 1206-1216.

Inguagiato, J.C., J.A. Murphy, and B.B. Clarke. 2009. Anthracnose disease and annual bluegrass putting green performance affected by mowing practices and lightweight rolling. *Crop Science*. 49. p. 1454-1462.

Inguagiato, J.C., J.A. Murphy, and B.B. Clarke. 2012. Sand topdressing rate and interval effects on anthracnose severity of an annual bluegrass putting green. *Crop Science*. 52. p. 1406-1415.

Jiang, H., J.D. Fry, and N.A. Tisserat. 1998. Assessing irrigation management for its effects on disease and weed levels in perennial ryegrass. *Crop Science*. 38. p. 440-445.

McDonald, S.J., P.H. Dernoeden, and C.A. Bigelow. 2006. Dollar spot control in creeping bentgrass as influenced by spray volume and application timing. *Applied Turfgrass Science*. doi.:10.1094/ATS-2006-0531-01-RS.

Moore, L.D., H.B. Couch, and J.R. Bloom. 1963. Influence of environmental disease of turfgrasses. III. Effect of nutrition, pH, soil temperature, air temperature, and soil moisture on pythium blight of highland bentgrass. *Phytopathology*. 53. p. 53-57.

Roberts, J.A., et al. 2011. Irrigation quantity effects on anthracnose disease of annual bluegrass. *Crop*

Science. 51. p. 1244–1252.

- Roberts, J.A., et al. 2012. Lightweight rolling effects on anthracnose of annual bluegrass putting greens. *Agronomy Journal*. 104. p. 1176-1181.
- Rowell, J.B. 1951. Observations on the pathogenicity of *rhizoctonia solani* on bentgrasses. *Plant Disease Reporter*. 35. p. 240-242.
- Settle, D.M., J.D. Fry, and N.A. Tisserat. 2001a. Development of brown patch and pythium blight in tall fescue as affected by irrigation frequency, clipping removal, and fungicide application. *Plant Disease*. 85. p. 543-546.
- Thompson, D.C., B.B. Clarke, and J.R. Heckman. 1995. Nitrogen form and rate of nitrogen and chloride application for the control of summer patch in Kentucky bluegrass. *Plant Disease*. 79. p. 51-56.
- Williams, D.W., A.J. Powell, P. Vincelli, and C.T. Dougherty. 1996. Dollar spot on bentgrass influenced by displacement of leaf surface moisture, nitrogen, and clipping removal. *Crop Science*. 35. p. 1304-1309.

Pesticide Management References

- Dell, C.J., C.S. Throssell, M. Bischoff, and R.F. Turco. 1994. Estimation of sorption coefficients for fungicides in soil and turfgrass thatch. *Journal of Environmental Quality*. 23. p. 92-96.
- Doherty, J. 2017. Golfer exposure to pesticides. Doctoral dissertation. Available: https://scholarworks.umass.edu/dissertations_2/1146/.
- Gustafson, D.I. 1989. Groundwater ubiquity score: A simple method for assessing pesticide leachability. *Environmental Toxicology and Chemistry*. 8. p. 339-357. doi:10.1002/etc.5620080411.
- Kovach, J., C. Petzoldt, J. Degni, and J. Tette. 1992. A method to measure the environmental impact of pesticides. *New York's Food and Life Sciences Bulletin*. 139. p. 1–8. Available: <https://ecommons.cornell.edu/handle/1813/55750>
- Krutz, L.J., S.A. Senseman, R.M. Zablotowicz, and M.A. Matocha. 2005. Reducing herbicide runoff from agricultural fields with vegetative filter strips: A review. *Weed Science*. 53. p. 353-367. doi: 10.1614/WS-03-079R2.
- Lickfeldt, D.W., and B.E. Branham. 1995. Sorption of nonionic organic compounds by Kentucky bluegrass leaves and thatch. *Journal of Environmental Quality*. 24. p. 980-985.
- McCarty, L.B, I.R. Rodriguez, B.T. Bunnell, and F.C. Waltz. 2003. *Fundamentals of turfgrass and agricultural chemistry*. John Wiley & Sons, Hoboken, NJ.
- Portmess, R.E., J.A. Grant, B. Jordan, A.M. Petrovic, and F.S. Rossi. 2014. *Best management practices for New York state golf courses*. 1st edition. Cornell University.
- Putnam, R.A., J.J. Doherty, J.M. Clark. 2008. Golfer exposure to chlorpyrifos and carbaryl following application to turfgrass. *Journal of Agricultural and Food Chemistry*. 56. p. 6616-6622. doi:

10.1021/jf800359b.

United States Environmental Protection Agency. 2014. Label review manual. Available: <https://www.epa.gov/pesticide-registration/label-review-manual>.

USGA Green Section. 2016. Tremendous savings from GPS spray technology. Oct. 21. United States Golf Association. Available: <https://www.usga.org/content/usga/home-page/course-care/water-resource-center/bmp-case-studies/tremendous-savings-from-gps-spray-technology.html>.

Pollinator References

Brust, G. 2015. Plants that attract pollinators and natural enemies. University of Maryland Extension. Available: http://extension.umd.edu/sites/extension.umd.edu/files/_docs/articles/PollinatorAndNaturalEnemyAttractingPlants-AdvancedMG_June2015.pdf

Ceplo, M. 2017. Increasing pollinator populations by improving habitat. USGA. Available: <http://archive.lib.msu.edu/tic/usgamisc/cs/292023.pdf>

Couto, A.V., and A.L. Averill. 2016. A review on bees/northeast crops edition. UMass Extension. Available: https://ag.umass.edu/sites/ag.umass.edu/files/pdf-doc-ppt/2-1-17_a_review_on_bees.pdf.

Larson, J.L., C.T. Redmond, and D.A. Potter. 2013. Assessing insecticide hazard to bumble bees foraging on flowering weeds in treated lawns. PLOS ONE. 8(6). doi:10.1371/journal.pone.0066375.

Larson, J.L., A. Dale, D. Held, B. McGraw, D.S. Richmond, K. Wickings, and R.C. Williamson. R. 2017. Optimizing pest management practices to conserve pollinators in turf landscapes: Current practices and future research needs. Journal of Integrated Pest Management. 8(1). doi.org/10.1093/jipm/pmx012

Kammerer, S., and A. Moeller. 2019. Hormonal responses– re they helping or hurting your greens? USGA Green Section Record. 57(23). Dec. 6. Available: <https://www.usga.org/course-care/green-section-record/57/23/hormonal-responsesare-they-helping-or-hurting-your-greens-.html>.

Massachusetts Department of Agricultural Resources. Massachusetts Pollinator Protection Plan. Available: <https://www.mass.gov/files/documents/2017/06/zw/pollinator-plan.pdf>

Richmond, D., and A. Patton. 2014. Neonicotinoid insecticides and pollinators: what's all the buzz about? Golf Course Management. 82(10).

Richmond, D., and C. Sadof. 2016. Protecting pollinators in home lawns and landscapes. Purdue Extension. Available: <https://extension.entm.purdue.edu/publications/POL-1/POL-1.html>.

Sgolastra, F., Medrzycki, P., Bortolotti, L., Renzi, M.T., Tosi, S., Bogo, G., Teper, D., Porrini, C., Molowny-Horas, R. and Bosch, J. (2017), Synergistic mortality between a neonicotinoid insecticide and an ergosterol-biosynthesis-inhibiting fungicide in three bee species. Pest. Manag. Sci., 73: 1236-1243. <https://doi.org/10.1002/ps.4449>

Thompson, H.M., Fryday, S.L., Harkin, S. et al. Potential impacts of synergism in honeybees (*Apis mellifera*) of exposure to neonicotinoids and sprayed fungicides in crops. Apidologie 45, 545–553

(2014). <https://doi.org/10.1007/s13592-014-0273-6>

Whitford, F., et al. 2017. The complex life of the honey bee. Purdue Extension. Available: <https://ppp.purdue.edu/wp-content/uploads/2017/PPP-116.pdf>.

Williamson, R.C. 2018. Pollinators in the turfgrass ecosystem: best management practices. Golf Course Management. Available: <https://www.gcmonline.com/course/environment/news/pollinator-best-management-practices>.

Sustainable Landscaping in Out-of-Play Areas References

Benedict, M.A., and E.T. McMahon. 2002. Green infrastructure: Smart conservation for the 21st century. Renewable Resources Journal. 20(3). p. 12–17.

Brame, R.A. 2012. Tall grass rough or natural rough? USGA Green Section Record. 50(12). Available: <https://www.usga.org/course-care/2012/06/tall-grass-rough-or-natural-rough-21474847128.html>.

Colding, J., and C. Folke. 2009. The role of golf courses in biodiversity conservation and ecosystem management. Ecosystems. 12(2).

Dobbs, E.K., and D.A. Potter. 2014. Conservation biological control and pest performance in lawn turf: Does mowing height matter? Environmental Management. 53(3). p. 648–659.

Dobbs, E.K., and D.A. Potter. 2013. Operation pollinator for golf courses: Naturalized areas on golf courses can boost declining populations of native pollinators by providing habitat in out-of-play areas. Golf Course Management. 81(4).

Gillette, K. 2014. The Colorado golf carbon project. Doctoral dissertation: Colorado State University.

Gillette, K.L., R. Follett, Y. Qian, A. Koski, and S. Del Grosso. 2011. The carbon footprint of Colorado golf courses estimated by clubhouse energy use, trace gas emissions and modeling carbon sequestration. ASA-CSSA-SSSA International Annual Meetings. No. 65375.

Gross, P., and T. Eckenrod. 2012. Turf reduction template: A guideline for reducing turf acreage while maintaining golf course quality. USGA Green Section Record. 50(12). p. 1–5.

Lal, R., and B. Augustin, editors. 2011. Carbon sequestration in urban ecosystems. New York, New York: Springer.

Lyman, G.T., C.S. Throssell, M.E. Johnson, and G.A. Stacey. 2007. Golf course profile describes turfgrass, landscape, and environmental stewardship features. Applied Turfgrass Science.

Mader, E., M. Shepherd, M. Vaughan, S.H. Black, and G. LeBuhn. 2011. Attracting native pollinators: Protecting North America's bees and butterflies. Storey Publishing.

Mid-Atlantic Chapter of the Golf Course Superintendents Association of America, Eastern Shore Association of Golf Course Superintendents, and University of Maryland. 2017. Best management practices for Maryland golf courses.

- Porter, E.E., D.N. Pennington, J. Bullock, and R. B. Blair. 2004. Assessing the conservation value of golf courses for butterflies. *USGA Turfgrass and Environmental Research Online*. 3(14). p. 1-13.
- Schrum, H. 2018. Wildflower meadows for the busy superintendent (because aren't they all?). *Golfdom*. 74(8). p. 12-17.
- Selhorst, A.L., and R. Lal. 2012. Carbon sequestration in golf course turfgrass systems and recommendations for the enhancement of climate change mitigation potential. R. Lal and B. Augustin, editors. *Carbon Sequestration in Urban Ecosystems*. Springer.
- Shackelford, G. 2003. Go native: It can satisfy environmentalists – and be good for the bottom line, too. *Golfdom*. 59(11). p. 62, 64, 66, 68.
- Tallamy, D. 2009. *Bringing nature home: How you can sustain wildlife with native plants*. 2nd ed. Portland, OR.: Timber Press.
- Tanner, R.A., and A.C. Gange. 2005. Effects of golf courses on local biodiversity. *Landscape and Urban Planning*. 71(2-4). pp. 137-146.
- University of California at Davis. 2008. Designing wildlife corridors: Wildlife need more complex travel plans. *Science Daily*. Oct. 21. Available: <https://www.sciencedaily.com/releases/2008/10/081020135221.htm>
- United States Environmental Protection Agency. 2017. *Green infrastructure in parks: A guide to collaboration, funding, and community engagement*. U.S. EPA.
- Wissman, Jörgen. 2016. *Multifunctional golf courses*. Swedish University of Agricultural Sciences and Scandinavian Turfgrass and Environment Research Foundation.
- Zimmerman, C. 2010. *Urban and suburban meadows: Bringing meadowscaping to big and small spaces*. Silver Spring, Maryland: Matrix Media Press.

Energy Management References

- U.S. Department of Energy. 2011. *Clean cities guide to alternative fuel commercial lawn equipment*. Available: <https://afdc.energy.gov/files/pdfs/52423.pdf>.
- Hartwiger, C. 2010. *Course care: A penny saved is a penny earned*. United States Golf Association. July 5. Available: <http://www.usga.org/articles/2010/07/course-care-a-penny-saved-is-a-penny-earned-2147487886.html>